

## Anatomical Studies of Some *Euphorbia* species

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### Abstract

This study include 32 species all belonging to the genus *Euphorbia* ( Euphorbiaceae) .These samples collected from different floristic regions in Egypt. The anatomical studies occur on the stems of the plant samples and the characters are recorded comparatively for each species. The observations showed that the stems are mostly angled, the epidermis is often simple with one layer, the cortical layers of many species contained storage parenchyma, cells like palisade tissue. Transfer tissue, air cavities and idioblast also calcium oxalate observed in the cortex. The pith is solid in most samples and hollow in a few species.

**Key word:** *Euphorbia*, floristic, cortex, hollow

### Introduction

Euphorbiaceae family is the sixth largest family, and one of the most several families of angiosperms, consisting of about 300 genera and over 8000 species (Radcliffe-Smith 1986). The largest genus is *Euphorbia* (spurge) consisting of over 1600 species (Evans and Taylor 1983), these species are more common in tropical areas all over the world. Its habitat ranges from herbs, shrubs to trees and cacti type. The genus *Euphorbia* is characterised by the presence of milky latex, being more or less toxic (Singia and Pathak 1990). This work aims to clear the variations between the stems structures of some *Euphorbia* species.

### Materials and methods

#### 1- Materials

Samples of plants representing genus *Euphorbia* were taken from 32 species. All materials were collected from various floristic regions in Egypt. The identification of the collected plants was achieved by comparing their morphological characters with the characters of the previously identified plants as published by Täckholm (1974), Migahid (1989) ) and Boulos (2000). The studied species arranged alphabetically in Table (1).

**Table 1.** Alphabetical list of (32) species representing the genus *Euphorbia*.

No.	Species	No.	Species
1	<i>Euphorbia ambovombensis</i> Rauh&Razaf	17	<i>E. mellifera</i> Aiton
2	<i>E. caputa-medusae</i> L.	18	<i>E. milii</i> var.splendens Des Moul
3	<i>E. cuneata</i> Vahl.	19	<i>E. obese</i> Hook.f.
4	<i>E. deciduas</i> Bally & Leach	20	<i>E. paralias</i> L.
5	<i>E. fasciculata</i> Thunb	21	<i>E. peplus</i> L.
6	<i>E. fimbriata</i> Scop	22	<i>E. polyacantha</i> Boiss.
7	<i>E. grandicornis</i>	23	<i>E. pulcherrima</i> Willd. ex Klotzsch.
8	<i>E. helioscopia</i> L.	24	<i>E. pulvinata</i> Marloth
9	<i>E. heterophylla</i> L.	25	<i>E. retusa</i> Forssk.
10	<i>E. hierosolymitana</i> Boiss.	26	<i>E. royleana</i> Boiss.
11	<i>E. hirta</i> L.	27	<i>E. serpens</i> Kunth
12	<i>E. hyssopifolia</i> L.	28	<i>E. suzannae</i> Marloth
13	<i>E. indica</i> Lam.	29	<i>E. tirccalli</i> Linné
14	<i>E. lacteacrestata</i> Haw	30	<i>E. tithymaloides</i> L.
15	<i>E. lacteacandelabra</i>	31	<i>E. trigona</i> Miller
16	<i>E. mammillaris</i> L.	32	<i>E. vansenvillrnsis</i>

#### 2- Methods

Parts of stem samples were fixed in F.A.A. for a minimum period of 48 hours. Stem samples were prepared by the method suggested by Sass (1958), sample of one centimeter long from the middle part of the technical length of the stem. Samples were

dehydrated in series solutions of ascending concentrations of ethyl alcohol varying from 50% to 100% ethyl alcohol. The samples were then embedded in paraffin wax [m. p. 58-61c°] using xylol as solvent. By using rotary microtome, sections were cut at the thickness of 15 microns and then

mounted on slides with the aid of egg- albumin as an adhesive. Wax dissolved in xylol and the slides were passed through descending series of ethyl alcohol solutions varying from 100% to 50% ethyl alcohol concentrations in descending order. The sections on the slides were stained with safranin and light green and then the colored sections were kept as permanent

preparations on the slides with Canada balsam as mounting medium.

Sections in such cases were microscopically explored for the different microphotographs, which can be explored for the different tissues and components.

**Table 2.** List of (41) characters recorded comparatively for (32) *Euphorbia* spp. The characters were distinguished into (38) qualitative and (3) numerical characters respectively.

#### A. Qualitative characters

- 1- Outline shape: rounded (+) / angled (-)
- 2- Bark present (+) / absent (-)
- 3- Cuticle smooth (+) / rough (-)
- 4- Epidermis, hypodermis present (+) / absent (-)
- 5- “ prismatic crystals present (+) / absent (-)
- 6- “ druses crystals present (+) / absent (-)
- 7- “ agglomerated crystals present (+) / absent (-)
- 8- Cortex, cells like palsied present (+) / absent (-)
- 9- “ storage parenchyma present (+) / absent (-)
- 10- “ transfer tissue present (+) / absent (-)
- 11- “ sclerenchyma cells present (+) / absent (-)
- 12- “ air cavities present (+) / absent (-)
- 13- “ laticifers canals present (+) / absent (-)
- 14- “ idioblast present (+) / absent (-)
- 15- “ prismatic crystals present (+) / absent (-)
- 16- “ druses crystals present (+) / absent (-)
- 17- “ agglomerated crystals present (+) / absent (-)
- 18- “ cortical vascular bundles present (+) / absent (-)
- 19- Pericycle, only parenchyma cells (+) / parenchyma with sclerenchyma-
- 20- “ idioblast present (+) / absent (-)
- 21- “ prismatic crystals present (+) / absent (-)
- 22- “ druses crystals present (+) / absent (-)
- 23- “ laticifers present (+) / absent (-)
- 24- “ agglomerated crystals present (+) / absent (-)
- 25- Phloem, idioblast present (+) / absent (-)
- 26- “ laticifers present (+) / absent (-)
- 27- “ druses crystals present (+) / absent (-)
- 28- xylem, vessels in series present (+) / in clusters (-)
- 29- “ prismatic crystals present (+) / absent (-)
- 30- pith, sclerenchymatus cells present (+) / absent (-)
- 31- “ aerenchyma cells present (+) / absent (-)
- 32- “ storage parenchyma present (+) / absent (-)
- 33- “ idioblast present (+) / absent (-)
- 34- “ druses crystals present (+) / absent (-)
- 35- “ agglomerated crystals present (+) / absent (-)
- 36- “ air cavities present (+) / absent (-)
- 37- “ laticifers present (+) / absent (-)

#### B - Numerical characters

- 38- Stem cuticle layer, average thickness in  $\mu$ .
- 39- “ epidermal layer, average thickness in  $\mu$ .
- 40- “ xylem vessels, average dimension in  $\mu$ .

### Results and Discussion

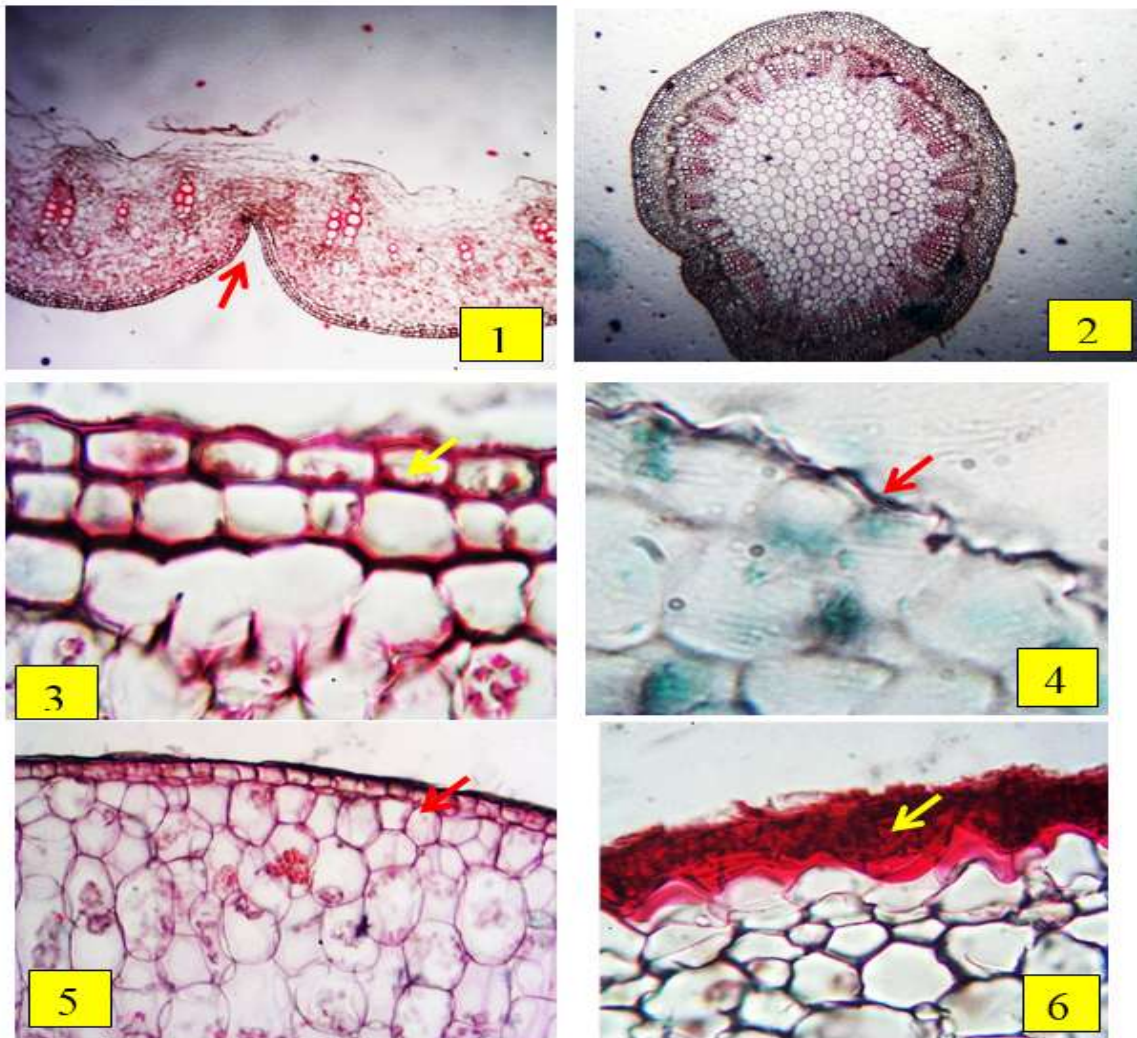
The investigation showed that, the stem varies in the outline shape, it is mostly angled e.g. *Euphorbia heterophylla* L (Fig.1), or rounded as in *E. hirta* L (Fig.2). The results are in harmony with the finding of Luz, et al (2015) who recorded that the stem

transverse section showed a circular outline. The cuticle layer varies in thickness from 1.33  $\mu$  in *E. trigona* Miller to 17.33  $\mu$  in *E. obesa* Hook.f. The cuticle is usually smooth or straight this showed in all studied samples e.g. *E. heterophylla* L (Fig.3), except in *E. retusa* Forssk which has rough cuticle (Fig.4). The average thickness of epidermal layer

ranged between  $5.26\mu$  in *E. heterophylla* L to  $26.33\mu$  in *E. obesa* Hook.f. Epidermal layer is simple (present in one layer) in most of the examined species e.g. *E. tithymaloides* L (Fig.5), except two plant samples which have multiple epidermis (hypodermis) *E. grandicornis* and *E. heterophylla* L (Fig.3). Bark is observed in a few of the examined samples e.g. *E. milii* var.splendens Des Moul (Fig.6). These results are in agreement with those obtained by **Kakkar and Paliwal.(1972)** , **Watson and Dallwitz (1992)** **Gales & Toma (2007)**, who recorded that the epidermis is covered by a cuticle of variable thickness, the stem cork was present.

In cortex the storage parenchyma are recorded in most examined samples e.g. *E. mellifera* Aiton (Fig.7). The chlorenchymatous cells like palisade tissue are noticed only in two samples *E. tirucalli* Linné and *E. retusa* Forssk (Fig.8). Transfer tissue is

recorded in some examined samples as in *E. lactea cristata* Haw (Fig.9). And air cavities are also observed in the stem cortical. Idioblast present in some plants e.g. *E. ambovombensis* Rauh & Razaf and laticifer present in some plants e.g. *E. fasciculata* Thunb (Fig.10). Cortical vascular bundles present in some plants e.g. *E. caputa-medusae* L (Fig.11). Prismatic crystals observed in the cortex of some plants. Agglomerate and druses crystals observed in the cortex of some plants. The investigation are both idioblast and laticifer canals in the stem cortical. These results are in agreement with those obtained by **Jafari & Nasseh (2009)** , **Luković. et al (2009)** , **Sultana (2017)** who recorded that both the paranchyma and chlorenchyma cells were present in cortex of *Euphorbia* spp. Laticifer canals and storage parenchyma were also present in the cortex layer.



(Fig.1) T.S. of *Euphorbia heterophylla* L angled stem (x=40).

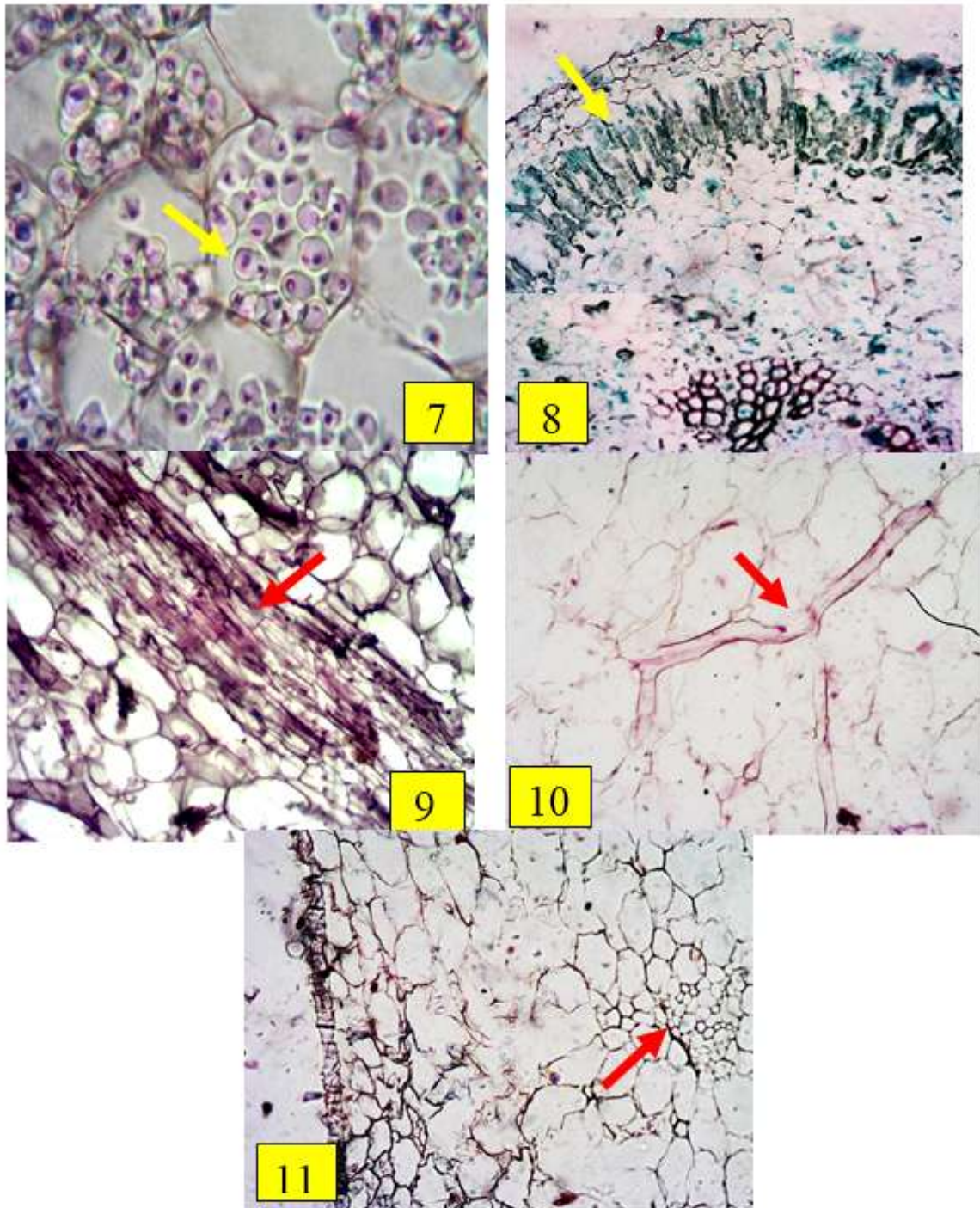
(Fig.2) T.S of *E. hirta* L round stem (x=40).

(Fig.3) T.S of *E. heterophylla* L smooth cuticle and hypodermis (x=400).

(Fig.4) T.S of *E. retusa* Forssk rough cuticle (x=400).

(Fig.5) T.S of *E. tithymaloides* L. simple epidermis layer (x=200).

(Fig.6) T.S of *E. milii* var.splendens Des Moul Bark. (x=400).



(Fig.7) T.S. of *Euphorbia mellifera* Aiton storage parenchyma (X=450)

(Fig.8) T.S of *E. retusa* Forssk Cells like palisade tissue (X=100)

(Fig.9) T.S. of *E. lactea cristata* Haw transfer tissue (X=100)

(Fig.10). T.S. of *Euphorbia fasciculata* Thunb laticifer(X=100)

(Fig.11). T.S. of *E. caputa-medusae* L cortical vascular bundles (X=100)

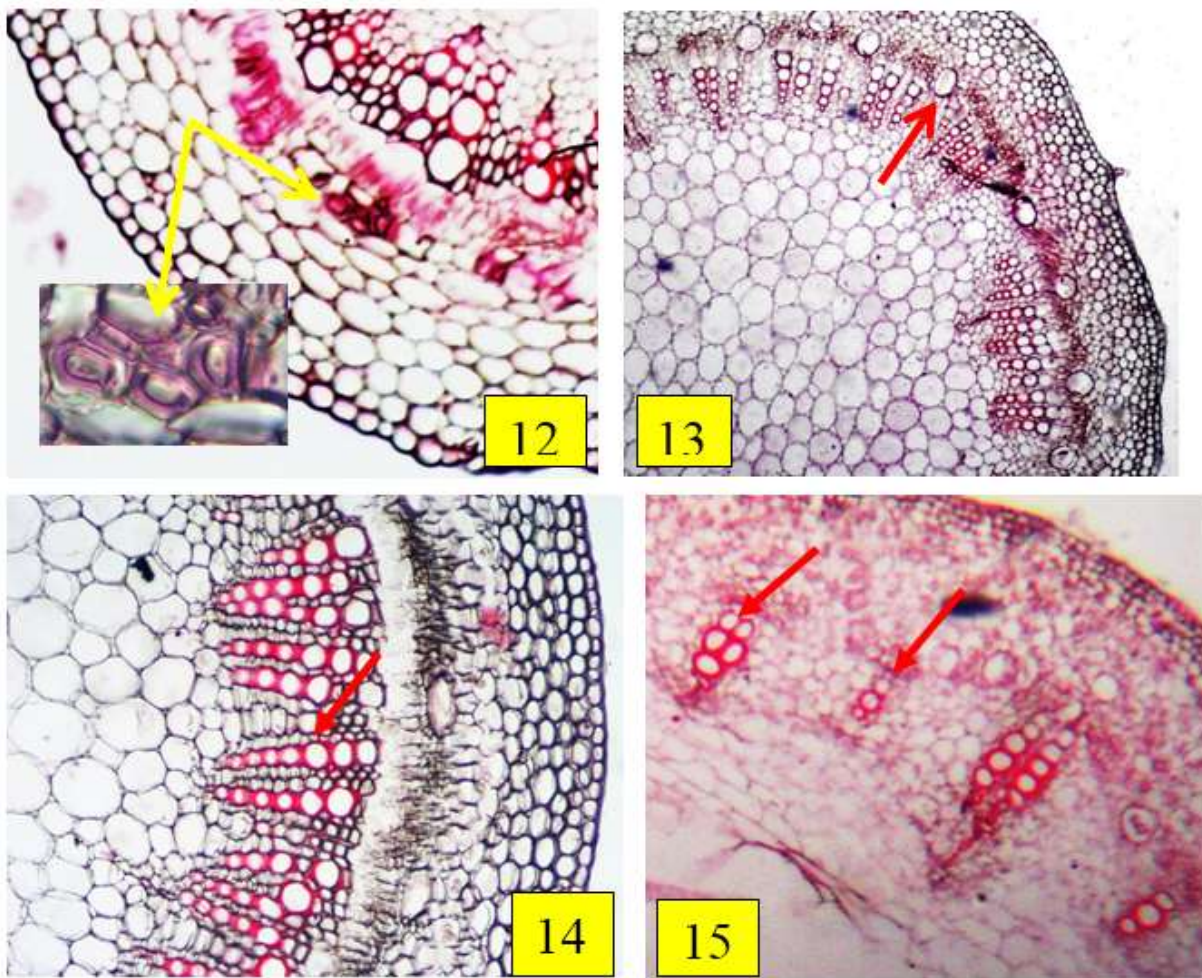
The pericycle layer is usually consists of parenchymatous cells only, in this study some plant samples pericycle layer consists of parenchymatous cells accompanied with sclerenchymatus cells e.g. *E. serpens* Kunth (Fig.12). Both Idioblast and laticifer canals are recorded in some examined samples e.g. *E. hirta* L (Fig.13). Also druses and prismatic

crystals are observed in some studied samples in this layer. Agglomerate crystals are observed in some studied samples in this layer. The previous results are in harmony with the finding of **Luković. et al (2009)** who recorded that pericyclic groups of sclerenchyma tissue occur above the phloem region of vascular bundles. Laticifer canals are recorded in the pericycle

layer. Idioblast cells and laticifer canals are recorded in the phloem of some examined samples also druse crystals are observed. The results are in harmony with the finding of **Bercu,R. (2016)** who recorded that in the phloem zone, few laticifers are present.

Xylem in most examined samples stems occur in series or arms such as *E. hyssopifolia* (Fig.14), while in other samples the vessels are in clusters, and few samples have the two states e.g. *E. heterophylla* L(Fig. 15). The average dimension of xylem vessels ranged between  $9.4\mu$  in *E. suzannae* Marloth and  $80.8\mu$  in *E. heterophylla* L. Most examined samples have solid stem e.g. *E. tithymaloides* L. (Fig. 16), while few species have hollow stem e.g. *E. heterophylla* L. (Fig.17). The stem pith is usually consists of parenchyma cells e.g. *E. hirta* L. (Fig.18),

except in *E. cuneata* Vahl. (Fig. 19) has only sclernchyma cells. Also aerenchyma cells are recorded only in the pith of *E. helioscopia* L. (Fig.20). Many samples have storage cells in their pith e.g. *E. tithymaloides* L. Air cavities are also observed in the pith of some studied samples, both idioblast and laticifer canals are recorded in some species such as *E. hirta* (Fig.21). Prismatic oxalate crystals present in the pith of many samples e.g. *E. milii* var.splendens Des Moul (Fig.22). The results are in harmony with the finding of **GALEŞ &TOMA (2006), Gales & Toma (2007), Zokian (2011)** who recorded that the pith is parenchyma cells, having air cavities. In some samples pith often becomes hollow. Also laticiferes canals were observed in the pith.

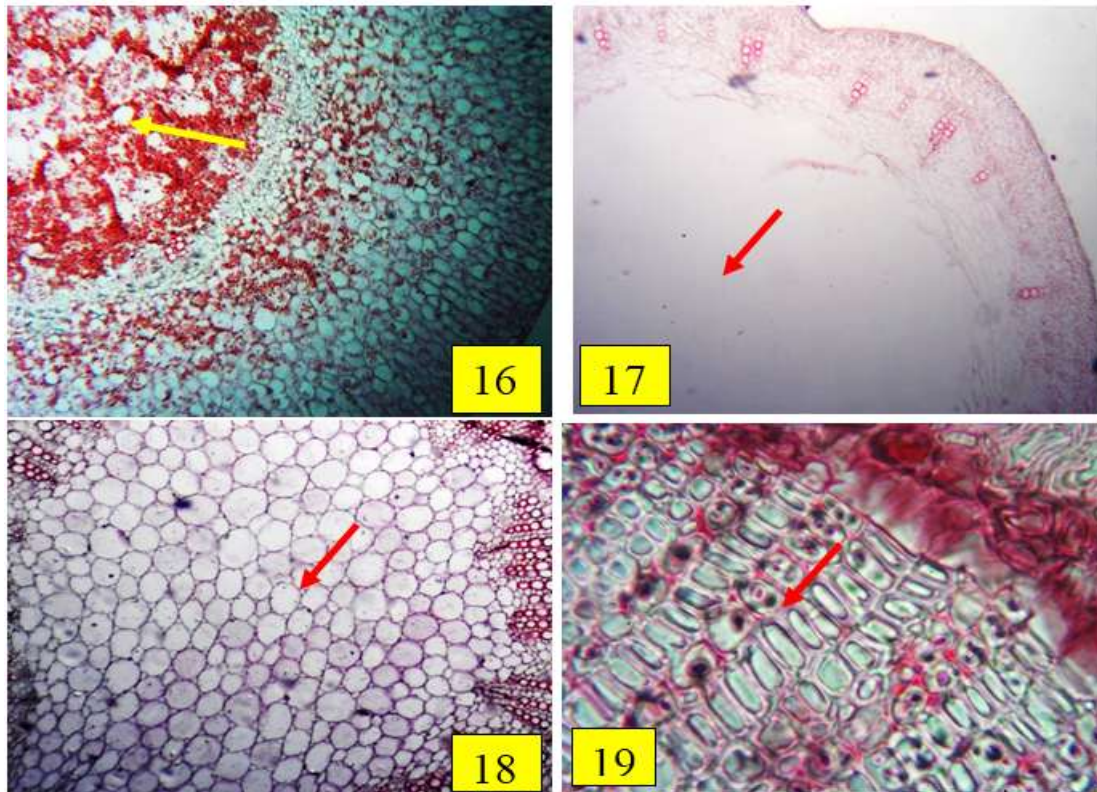


(Fig.12). T.S. of *E. serpens* Kunth sclerenchymatous cells(X=100 - X=400)

(Fig.13) T.S. of *E. hirta* L laticifer canals in pericycle layer (X=100)

(Fig.14) T.S. of *Euphorbia hyssopifolia* L xylem vessels in clusters (X=100)

(Fig.15) T.S. of *E. heterophylla* vessels in clusters and agglomeration (X=100)

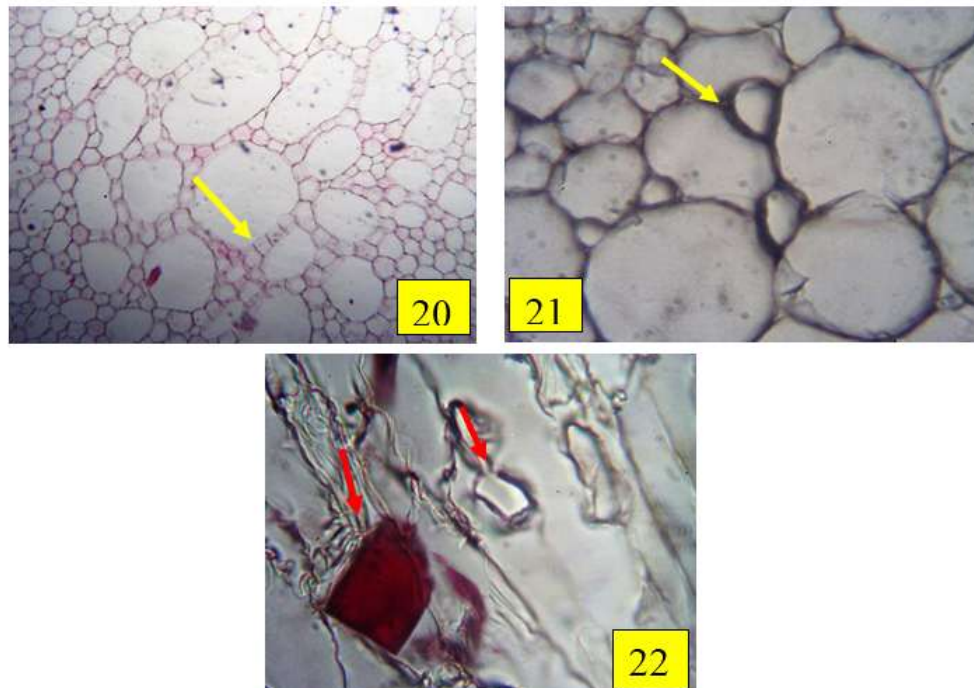


(Fig.16) T.S. of *E. tithymaloides* L solid stem (X=40)

(Fig.17) T.S. of *E. heterophylla* L hollow stem (X=40)

(Fig.18) T.S. of *Euphorbia hirta* L parenchyma cells in the pith (X=40)

(Fig. 19) T.S. of *E. cuneata* Vahl sclerenchyma cells in the pith (X=400)



(Fig.20) T.S. of *E. helioscopia* L aerenchyma cells in the pith (X=40)

(Fig.21) T.S. of *E. hirta* L laticifer canals in the pith . (X=400)

(Fig.22) T.S. of *E. milii* var. *splendens* Des Moul L. Agglomerate and prismatic crystals in the pith (X=400)

**Table (3).** Data matrix of the (41) observed characters recorded comparatively for (32) species from genus Euphorbia. The characters were distinguished into (38) qualitative and (3) numerical. Missing characters denoted by (\*)

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	
1	+	+	+	-	+	-	-	-	-	-	-	+	+	-	+	-	-	+	-	+	+	-	-	-	-	-	±	-	-	-	+	-	+	-	-	-	*	*	13.4		
2	-	-	+	-	-	-	-	-	-	-	-	-	+	-	-	-	-	+	+	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	3.6	18	8.1	
3	+	-	-	-	-	-	-	-	-	-	-	-	+	+	-	-	-	+	+	-	-	-	-	-	-	-	+	-	+	-	-	-	-	-	-	-	-	1.86	10.4	17.66	
4	-	-	+	-	-	-	-	-	-	+	-	-	+	-	-	-	-	+	+	-	-	-	+	-	-	-	±	-	-	-	-	-	-	-	-	-	+	3.8	19.1	9.8	
5	-	-	+	-	-	-	-	-	-	+	-	-	+	+	-	-	+	+	+	+	-	-	-	-	+	-	-	-	-	-	-	-	+	-	-	-	+	6.13	18.66	14.93	
6	-	-	+	-	+	-	-	-	-	+	-	-	+	+	-	-	-	+	+	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	+	2.6	17.13	16.66
7	-	-	+	+	-	-	-	-	-	-	-	+	+	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	+	+	2.53	13.13	17.26	
8	+	-	+	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	+	-	-	-	-	-	-	-	+	-	-	+	-	-	-	-	-	-	-	5.33	23	18.53	
9	-	-	+	+	-	-	-	-	+	-	+	+	+	-	-	-	-	-	+	-	-	-	+	-	-	-	±	+	-	-	-	-	-	-	-	-	-	2.6	5.26	80.8	
10	+	+	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	+	-	-	-	-	-	-	-	-	-	2	10	21.2	
11	+	-	+	-	-	-	-	-	-	-	-	-	+	+	-	-	-	-	+	-	-	-	+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	+	1.86	12.86	22.6
12	+	-	+	-	-	-	-	-	+	-	-	-	+	+	-	-	-	-	+	-	-	-	+	-	-	+	-	+	-	-	-	-	-	-	-	-	+	+	2.73	12.6	12.46
13	+	-	+	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	+	-	-	-	-	-	-	-	1.4	5.53	22.13
14	-	-	+	-	-	-	-	-	+	+	-	+	+	+	-	-	-	+	+	+	-	-	+	-	-	+	-	±	-	-	-	-	+	-	-	-	-	+	2.26	23.4	10.86
15	-	-	+	-	-	-	-	-	+	-	-	-	+	+	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	3.13	25	15.4
16	-	-	+	-	-	-	-	-	+	+	-	-	+	+	-	-	-	+	+	+	-	-	+	-	-	-	±	-	-	-	-	-	-	-	-	-	-	+	2.73	23.8	14.4
17	+	-	+	-	-	-	-	-	+	+	-	-	+	+	+	-	+	+	+	-	-	-	+	-	-	+	-	±	-	-	-	+	-	-	-	-	+	*	*	35.6	
18	-	+	-	-	-	-	-	-	-	-	-	-	+	+	-	+	+	+	+	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	+	*	*	25.33	
19	-	-	+	-	-	-	-	-	+	+	-	-	+	+	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	+	17.33	26.33	20.4
20	+	-	+	-	-	-	-	-	-	-	-	-	+	+	-	-	-	-	+	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	5.7	18.3	32.5
21	+	-	+	-	-	-	-	-	-	-	-	-	+	+	-	-	-	-	+	-	-	-	-	-	-	-	±	-	-	-	-	-	-	-	-	-	-	-	1.7	18.7	22.8
22	-	-	+	-	-	-	-	-	+	-	-	+	+	+	-	-	-	+	+	+	-	-	+	-	-	-	-	-	-	-	-	-	-	+	-	-	-	+	4	18.93	13.8
23	+	-	-	-	-	-	+	-	-	-	-	-	+	+	-	-	+	-	+	+	-	-	-	-	-	-	±	-	-	-	-	-	-	-	-	-	-	-	1.93	12.93	43.73
24	-	-	+	-	-	-	-	-	+	+	-	+	+	+	-	-	+	-	+	-	-	-	-	-	-	-	±	-	-	-	+	-	-	-	-	-	+	+	6.86	16.86	15
25	+	-	-	-	-	-	-	+	-	-	-	+	-	+	-	-	-	+	+	-	-	-	-	-	-	-	±	-	-	-	-	-	-	-	-	-	-	-	1.86	19.53	29.93
26	-	-	-	-	-	-	-	-	+	-	-	+	+	+	-	-	-	+	+	-	-	-	-	-	-	-	±	-	-	-	-	-	-	-	-	-	-	+	1.6	16.86	27.66
27	+	-	-	-	-	-	-	-	-	-	+	-	+	-	-	-	-	-	+	-	-	-	-	-	-	-	±	-	-	-	-	-	-	-	-	-	-	-	1.93	13.86	24.53
28	-	-	+	-	-	-	-	-	+	+	-	-	+	+	-	-	-	+	+	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	4	14.73	9.4
29	+	-	-	-	-	-	-	+	-	-	-	+	-	+	-	-	-	-	+	-	-	-	-	-	-	-	±	-	-	-	-	-	-	-	-	-	-	-	2.8	11	19.6
30	+	-	+	-	-	-	-	-	+	-	-	-	+	+	-	-	-	-	+	-	-	-	+	-	-	-	±	-	-	-	-	-	-	-	-	-	+	+	1.93	13.8	18.73
31	-	-	+	-	-	-	-	-	-	+	-	-	+	+	-	-	-	+	+	-	-	-	-	-	-	-	±	-	-	-	-	-	-	-	-	-	-	+	1.33	9	10.26
32	-	-	+	-	-	-	-	-	+	+	-	+	+	+	-	-	-	-	+	-	-	-	+	-	-	-	±	-	-	-	-	-	-	-	-	-	+	+	7.46	17.53	19.6

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## الملخص العربي

احتوت هذه الدراسة على 32 نوع نباتي تنتمي إلى العائلة السوسيبية جنس الأيوفوريبيا وقد تم تجميع العينات من مناطق مختلفة بجمهورية مصر العربية . حيث تم عمل قطاعات تشريحية من سيقان النباتات محل الدراسة ثم حصر الصفات الخاصة بها ومقارنتها لكل نوع. وقد بينت النتائج أن معظم النباتات مزلعة وكانت خلايا البشرة عادة تتكون من صف واحد من الخلايا في حين احتوت منطقة البشرة في العديد من العينات على خلايا بارنثيمية مخزنة وأخرى عمادية وأخرى ناقلة بالإضافة إلى فجوات هوائية وخلايا إفرازية وبللورات من أكسالات الكالسيوم. النخاع مصمت في معظم العينات والقليل منها مجوف.