

Journal of Agricultural Chemistry and Biotechnology

Journal homepage: www.jacb.mans.edu.eg
Available online at: www.jacb.journals.ekb.eg

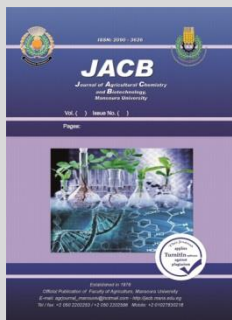
Using DNA Barcoding for Fingerprinting of Two Important Forage Crops Varieties (Alfalfa And Egyptian Clover)

Bondok, A. T.*

Department of Forage Crops Research, Field Crops Research Institute, Giza, Egypt



Cross Mark



ABSTRACT

Study was executed to differentiate and discriminate (*Medicago sativa* and *Trifolium alexandrinum*) using DNA barcoding genes [rbcL] and Cox1 genes. Identification of (*Medicago sativa* (Rammah 1) was completed via rbcL and Cox1 genes and was identified as *Medicago sativa* voucher G00199095 ribulose1,5 biphosphate carboxylase / oxygenase large subunit gene, partial cds; chloroplast Sequence ID: KJ204375.1 or *Medicago sativa* voucher Ahrendsen 23 for rbcL and Cox1 genes. Identity estimation were listed with 90% as alfalfa, Rammah 1 Genotype ribulose1 /5 biphosphate carboxylase / oxygenase large subunit gene sequences ID: KJ206375.1] also, identity values of 91.24% were recorded with for alfalfa Rammah 1 Genotype, cytochrome c oxidase subunit I gene cox 1 sequences ID: KJ 204375.1). *Trifolium alexandrinum* Helaly genotype was identified as *Trifolium alexandrinum* (Sequence ID: HM850407.1) and *Trifolium alexandrinum* voucher K-016Hv (Sequence ID: KU234213.1) by rbcL and Cox1 genes respectively. Affiliation of genetic source was revealed for *Trifolium alexandrinum* with 100 % match with origin which indicate rising possibility for applying discrimination through comparing with *Medicago sativa* which reflect the lowest genetic likeness with the source. Moreover, we might detect from the available data that we can use DNA Bar-coding Technique in Discriminating the local Egyptian Clover Genotypes and Protect Them internationally. Also, DNA Bar-coding can be used to determine the genetic polymorphism in identifying superior genotypes as source of parental genotypes in Egyptian clover breeding program in future.

Keywords: DNA Barcoding; rbcL; Cox 1; *Trifolium alexandrinum*; *Medicago sativa*; NCBI blast

INTRODUCTION

Sequences of DNA to organisms have planned as an elder path than traditional of taxonomic purist [Blaxter 2004; Tautz *et al.*, 2003]. Kress *et al.*, (2005) have pretend the performance DNA bar-coding angiosperms using nrDNA and non-coding sequences of cpDNA. Within trifolium, massive germplasm set utmost wild collected species occur [Morris & Greene, 2001]. Trifolium is member belong to the broad clad of legumes which lacks chloroplast copy repeat, IRLC (Lavin *et al.*, 1990 & Liston 1995). Phylogenetic studies had specific a strongly supported “vicioid of clad” within the IRLC the tribes of (Trifolieae & Fabeae). Molecular polymorphism with casual magnifies DNA (RAPD) polymorphic and the inter simple sequence repeat [ISSR] were employed to mark taxonomic relevance among 25 samples representing nine species of Orobanchaceae. Dendrogram generated by the evaluation of the molecular statistics (RAPD and ISSR) identify that structure by NJ dendrogram for the morphological distinction (Sahrawy and Karakishi 2015) estimated the use of two chloroplast regions, trnL and rpoC1, besides a nuclear internal transcriber region, ITS2, for their competence to barcode the master Mediterranean leguminous yields. Twenty-five legume species were deliberated. Species identification based the sequence uniformity tactic outright by the catalog of GenBank. DNA zone trnL & ITS2 positively 100% differentiate crop legume species in the Mediterranean region, whilst the rpoC.1 in particular around 72%. Likewise, the use of trnL zone vest the discrimination

of even mightily concerning species, like *Phaseolus lunatus* also *P. coccineus* & *Vicia faba* subsp foremost marginal of *V. faba* sub sp which is neatly united smooth of N.C.B.I they were both point out like *Phaseolus vulgaris* & *V. faba*, correspondingly. trnL and ITS2 are effective DNA bar-coding intention regions so as to select leguminous crops in Mediterranean and impart credible the effective tool for agricultural sciences likewise the community in artificial (Madesis *et al.*, 2012). Badr (2001) examined *Trifolium alexandrinum* utilize AFLP figures. In Syria and Egypt. The information backup adjacent connection *T. alexandrinum* accessions at *T. apertum* & *T. berytheum* or *T. salmoneum* strength of species to interbred blankly tick that is *T. salmoneum* & *T. berytheum* may be honor as the initial ancestors from whom the familiar Egyptian clover in Syria has meantime artificial selection. Dependent domestication could have been brought into the production of rain-fed crops in Palestine and irrigated to Egypt. In this respect, Egyptian clover urbanization may be the same as other crops such as wheat and barley, which were also domesticated in the fertile crescent and grown in the Nile Valley.

This imitates the genetic enhancement of the plant in backward cultivation in Egypt and varieties that were sophisticated in Egypt were later extended throughout the world. Parsimony and Bayesian phylogenetic analysis were conducted nuclear ribosomal of DNA inner spacer transcribed & chloroplast trnL intron acquired from 218 to ca. Number of 255 species of *Trifolium* envoy to 11 genera.

Incongruence via nrDNA and cpDNA tests demonstrate six cases of manifested hybrid speciation and describe th

* Corresponding author.
E-mail address: haae730a@yahoo.com
DOI: 10.21608/JACB.2019.58451

e fantastic allopolyploid progenitors. A common herb, *T. dubium*, and *T. Repens*, the most widely grown species of clover (Ellison *et al.*, 2006). Provenance ancestry of Egyptian clover was tested via AFLP information. A relative relationship of *T. alexandrinum* accessions from Syria and Egypt of *T. apertum*, *T. berytheum*, and *T. salmoneum*. Whilst cross ability and regional distribution indicate that is *T. apertum* is out of the way ancestor. In disparity, [*T. salmoneum*] look alike to be utmost prospective progenitor for Egyptian clover's Syrian substance though, close relevance to [*T. berytheum*] was revealed.

Such species ' openly crossing potency points out th at [*T. Salmoneum* & *T. Berytheum*] may be known as the f irst ancestors from whom Egyptian clover was domesticate d by man over Syrian selection. Kergoat *et al.*, (2004) reconstructed sequences in partial in three genes of mitochondrial [12S rRNA cytochrome. b and cytochrome c. oxidase subunit, I] phylogeny of seed beetles in Europe [Bruchidae] pertinence to Bruchus [Linnaeus & Bruchidius Schilsky]. In field the elder beetles were gained from larvae bred from seeds. Parsimony ultimate prospect and Bayesian reasoning were used to understand phylogenetic rapport between species. Genera [Bruchidius & Bruchus] formative monophyletic sets through whole analyses. Chloroplast-genome sequences encoded *rbcl*. *Medicago sativa* cv. gene Regen,S was confronted to pea. 94.1% of Alfalfa shares nucleotide sequence homology in pea 1721 bases cross beginning gene 213 bases upstream of the coding sequences over 83 bases into the 3' flanking region ending at locus 1508. Sequences of Peas are extremely ramosse alfalfa after that point. 94.3% of amino acid sequences are uniform to that of pea with about 56% (15/27) of the replacement non conservative (Aldrich *et al.*, 1987). Also, DNA barcodes from extreme herbal products about (91%) were recovered and whole leaf taking (100%) with (95%) species resolution utilize a tiered tactic (*rbcl* + *ITS2*). Utmost (59%), of the products examined include DNA barcodes from plant species not bound on labels. Whilst we were fancy to notarize roughly half about (48%) of the products one-third of these also rein contaminants and or fillers not recorded on the label. Product replacement appear in (30/44) of the products examined and only (2/12) of corporations had products without any swapping contamination or fillers. Somewhat of the contaminants was formed constitute critical health hazards to consumers (Newmaster *et al.*, 2013).

Table 1. Specific Primer sequence under study.

Forge crop	Coding genes	Primer sequence	Length	Tm	GC%
<i>Trifolium alexandrinum</i>	<i>Rbcl</i>	CAAGGCTTTGCGTGCTCTAC TATCGCGCAATAGTGAGCC	741	59.83 60.32	55.00 55.00
	<i>Cox1</i>	ATATTGCCCATAGAGGCCCTTC GCATAGTGATTGCTCCTGCT	289	59.69 58.04	50.00 50.00
<i>Medicago sativa</i>	<i>Rbcl</i>	CGGCTACCGATGGACTTACC GTTCCACCCTCTCCAGACG	339	59.97 60.04	60.00 60.00
	<i>Cox1</i>	TATGGTTTGCCGCGATGAT TTGTAATTGCCCTGCCAGT	759	60.18 59.89	50.00 50.00

RESULTS AND DISSCUSION

Specific gene detection technique:

Main purpose of this investigation is identifying and evaluating *Medicago sativa* and *Trifolium alexandrinum* probability discrimination Genotypes. Thus,

The aim of the present study was to:

- Use DNA barcoding (*rbcl* and *cox*) genes to identify and discriminate *Medicago Sativa* and *Trifolium alexandrinum* genotypes as two important forage crops.

MATERIALS AND METHODS

The seeds of alfafa and Egyptian clover were obtained from the Forage Crops Research Department (ARC) (*Medicago Sativa* – alfalfa, Rammah 1 and *Trifolium alexandrinum*, Egyptian clover Helaly) genotypes.

Methods:

Sequence Database for DNA Bar-coding:

It was applied for identification and comparing sequences under study was carried out at National Center for Biotechnology Information (NCBI) database.

Sampling Taxon and origin sequences.

Two Leguminosae samples (*Trifolium alexandrinum* and *Medicago sativa*) were studied including in reference database.

The extraction of DNA, amplification and sequencing

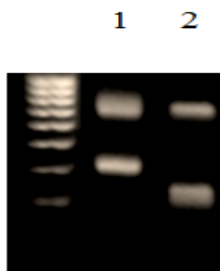
Samples in fresh stage collected and were stocked in silica prior to extraction. Extracted DNA by the Genomic of Gene JET purification kit following the protocol of manufacture. As shown in table (1), two plastid regions were amplified, [ribulose1, 5 bisphosphate carboxylase / oxygenase] large subunit gene (*rbcl*) and cytochrome c oxidase subunit 1 gene (*Cox1*) with specific primer according to [Kergoat *et al.*, 2004 and Cai *et al.*, 2008, Gurdon *et al.*, 2014, Young *et al.*, (2011)] into *Medicago sativa* and *Trifolium alexandrinum*. Magnify products were detached by gel electrophoresis (1.0%) Agarose. Obtained RT-PCR of products were purified from Agarose gel and quantities spectrophotometrically set for sequencing trial via ABI Prism 7000 instrument based on industrialist procedure

Nucleotide sequence accession numbers.

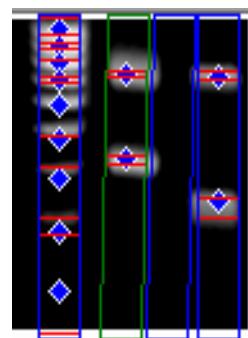
Nucleotide sequences of bar-coding genes (*rbcl* and *Cox1* genes) were submitted to be identified through program of NCBI BLAST - www. ncbi. nlm. nin. Gov / BLAST just as a single sense strand close sequence to each of Rammah 1 as well as Helaly genotypes. Products PCR were immediately sequenced at 2 sides to every fragment through a massive Dye v3.1 terminator Cycle sequencing kit (PE Applied Biosystems, City Foster, CA, USA.) in ABI 3730 an automated sequencer [P.E Applied Biosystems.]. CLUSTAL W program for sequences were used.

two bar-coding genes (*rbcl* and *Cox1* genes) were employed for identification. Based on alignment data with reference genes, genetic similarity and possibility for discrimination were evaluated for *Medicago sativa* and *Trifolium alexandrinum*.

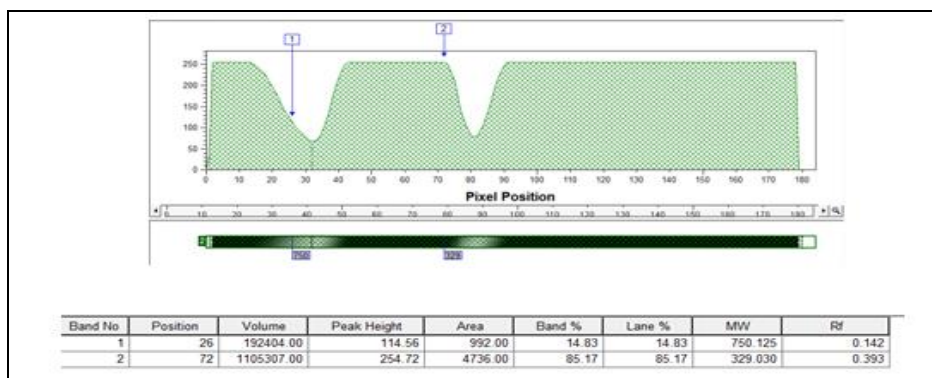
Photograph (1 and 2) show molecular weight parameters. Thus, specific fragments lengths were detected for each of *Medicago sativa* and *Trifolium alexandrinum*.



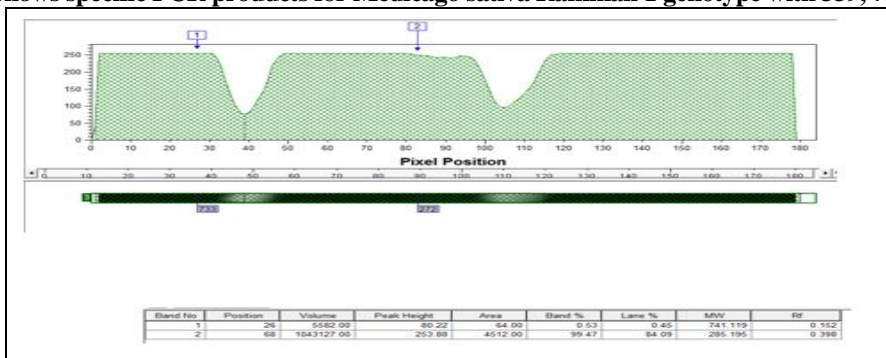
Photograph 1. shows specific PCR products for 1. *Medicago sativa* Rammah 1 genotype and 2. *Trifolium alexandrinum* Helaly genotype with 339, 759 bp and 741, 289 bp for *rbcl* marker gene plus cytochrome c oxidase subunit (1) gene respectively



Photograph 2. Detection of specific PCR products for *Medicago sativa* Rammah 1 genotype and *Trifolium alexandrinum* Helaly with 339, 759 bp and 741, 289 bp for *rbcl* marker gene in addition cytochrome c oxidase subunit (1) gene respectively



Photograph 3. Shows specific PCR products for *Medicago sativa* Rammah 1 genotype with 339, 759 bp.



Photograph 4. Shows specific PCR products for *Trifolium alexandrines* Helaly with 741, 289 bp.

Identification of *Medicago sativa* Rammah 1 genotype was performed through *rbcl* and genes *Cox1*. Figure (1) shows *rbcl* marker gene for *Medicago* indicating it as *Medicago sativa* voucher G00199095 ribulose 1, 5 bisphosphate carboxylase / oxygenase large subunit gene (*rbcl*) partial cds; chloroplast sequence ID; KJ204375. 1.

To evaluate genetic similarity for *Medicago sativa* sample, *rbcl* marker gene for *Medicago sativa* and *rbcl* original sequence were compared. Interestingly, comparison of data showed, 90 % of genetic similarity which was detected between *rbcl* marker gene for *Medicago sativa* and *rbcl* reference sequence (Fig. 2). For further confirmation the cytochrome c. oxidase subunit 1 gene (*Cox1*) marker gene were applied to identification

of *Medicago sativa* Rammah 1 genotype (Fig.3) and indicated it as *Medicago sativa* voucher Ahrendsen_23 cytochrome c. oxidase subunit 1 gene complete cds; mitochondrial.

For further confirmation the cytochrome c. oxidase subunit 1 gene *Cox 1*. marker gene was applied to identification of *Medicago sativa* Rammah 1 Genotype (Fig.3) and indicated it as *Medicago sativa* voucher Ahrendsen_23 cytochrome c. oxidase subunit (1) gene complete cds; mitochondria.

Highly genetic similarity (91.24) was found between the cytochrome c oxidase subunit 1 gene (*Cox1*.) for *Medicago sativa* and c oxidase subunit (1) gene (*Cox1*) for reference sequence and estimated with 91.24% (Fig.4).



Figure 1. Comparison alignments between *rbcL* marker gene for *Medicago sativa* Rammah 1 genotype and *rbcL* reference sequence.

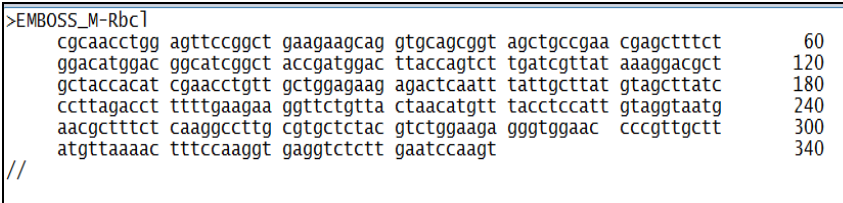


Figure 2. Shows *rbcL* marker gene sequence for *Medicago sativa* Rammah 1 genotype. (DNA Barcoding of alfalfa Rammah I Genotype (*rbcL*) gene).

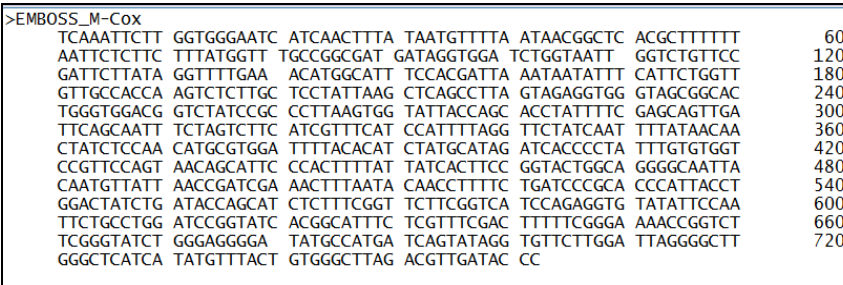


Figure 3. Shows cytochrome c oxidase subunit 1 gene (*Cox1*) marker gene sequence for *Medicago sativa* Rammah 1 genotype.

(DNA Barcoding of alfalfa Rammah 1 Genotype (*Cox 1*) gene).

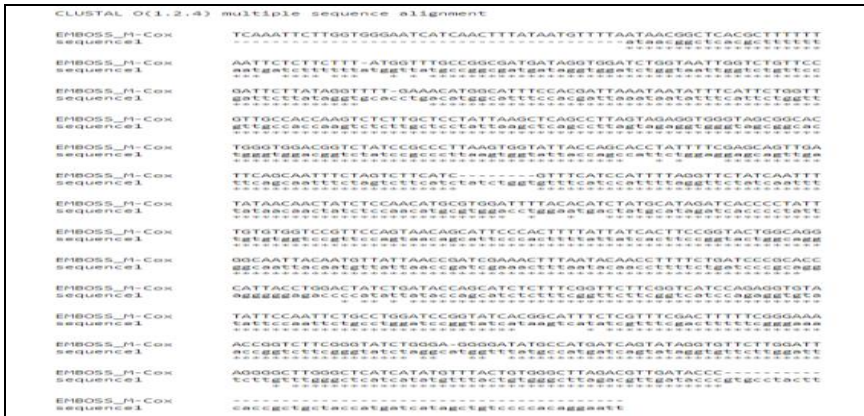


Figure 4. Comparing alignments between the cytochrome c oxidase subunit (1) gene to *Medicago sativa* Rammah 1 genotype and *cox 1* reference sequence.

Suspected *Trifolium alexandrinum* Helaly genotype was identified based [ribulose 1, 5 bisphosphate carboxylase / oxygenase large subunit (*rbcL*) gene] *Trifolium* sample was identified as *Trifolium alexandrinum* [ribulose 1, 5 bisphosphate carboxylase / oxygenase large subunit (*rbcL*) gene] partial cds: chloroplast. [Sequence ID: HM850407.1] with 100% of genetic identity (fig.5).

To estimate genetic relationship between *Trifolium alexandrinum* and genetic origin of *Trifolium alexandrinum*, alignment results were analyzed between [ribulose 1, 5 bisphosphate carboxylase / oxygenase large subunit (*rbcL*) gene for *Trifolium alexandrinum*. Helaly genotype in addition [ribulose 1, 5 bisphosphate carboxylase / oxygenase large subunit (*rbcL*) reference of

gene. Thus, 95.92 % of genetic similarity was recorded (fig.6).

In the light of *rbcl* marker identification gene, comparing the cytochrome c. oxidase subunit 1. gene Cox1 gene marker to *Trifolium alexandrinum* indicate uniformity

as *Trifolium alexandrinum* voucher K-016Hv cytochrome c. oxidase (C.O.I) gene, partially CDS; mitochondrial (Sequence of ID: KU234213.1) with 100 % at genetic similarity (figure 7).

```
>EMBOSS_Tri-rbc1
ACCACATCGA GCCGGTTGCT GGAGAAGAAA CTC AATTTAT TGCTTATGTA GCTTATCCCT      60
TAGACCTTTT TGAAGAAGGT TCTGTACTA ACATGTTTAC CTCCATTGTA GGTAATGTAT      120
TTGGGTTCAA GGCTTTGCGT GCTCTACGCC TTGGAAGATTT GCGAATCCCC GTTGCTTATG      180
TTAAAACCTT CCAAGGTCCT CCTCACGGAA TCCAAGTTGA GAGAGATAAA TTGAACAAGT      240
ATGGACGTCC CCTATTGGGA TGTACTATTA AACCTAAATT GGGTTTTATCC GCTAAGAATT      300
ACGGTAGAGC AGTTTATGAA TGTCTACGCG GTGGACTTGA TTTTACAAA GATGATGAAA      360
ATGTGAACTC CCAACCATTT ATGCGTTGGA GAGACCGTTT CTTATTTTGT GCCGAAGCTA      420
TTTATAAATC ACAGGCCGAA ACGGGTGNNN TCACGGAAAT NNNNNNNNNN NNNNNNNNNN      480
NNNTTCCGGT GCGGTTGTTT GGCTGTATT GCAAGAGAA TGGGC GTTCC TATAGGCCAC      540
TAATGCAGGA CTACCTAACA GGCGATTCA CTGCAAATC TACCCTGGCT CACTATTGCC      600
GCGATAATGG TCTACTTCTT CATATCCACC GTGCAATGCA TGCAGTTATC GATAGACAGA      660
AAAATCATGG TATGCACCTT CGTGTATTAG CTAAGCGTT ACGTTTGTCT GGTGGAGATC      720
ATATTCACGC CGTACTGTA G                                     741
//
```

Figure 5. Shows the [ribulose 1,5-bisphosphate carboxylase/oxygenase large subunit (*rbcl*)] marker gene sequence for [*Trifolium alexandrinum*] Helaly Genotype. (DNA Barcoding of Egyptian clover Helaly Genotype (*rbcl*) gene).

```
EMBOSS_Tri-rbc1 -----ACCCACATCGAAGCCGTTGCTGGAGAAGAAA
HM558487.1 CCAGTCTTGGATCGTTATAAAGSACGCTGCTACCCACATCGAAGCCGTTGCTGGAGAAGAAA

EMBOSS_Tri-rbc1 -----CTCAAAATTTATTTGCTTATGTAGSCTTATCCCTTAGACCTTTTGAAGAAGGTTCTGTTACTA
HM558487.1 CTCAAAATTTATTTGCTTATGTAGSCTTATCCCTTAGACCTTTTGAAGAAGGTTCTGTTACTA

EMBOSS_Tri-rbc1 -----ACATGTTTACCTCCATTGTAGSCTTATTTGGGTTCAAGGCTTTGCGTGTCTCACGCC
HM558487.1 ACATGTTTACCTCCATTGTAGSCTTATTTGGGTTCAAGGCTTTGCGTGTCTCACGCC

EMBOSS_Tri-rbc1 -----TGAAGAATTTGCGAATCCCGTTGCTTATGTTAAAACCTTTCCAAGGTCCTCCCTCACGGAA
HM558487.1 TGAAGAATTTGCGAATCCCGTTGCTTATGTTAAAACCTTTCCAAGGTCCTCCCTCACGGAA

EMBOSS_Tri-rbc1 -----TCCAAGTTGAGAGAGATAAAATGAAACAAGTATGGACGTCCTCCATTGGGATGTACTATTA
HM558487.1 TCCAAGTTGAGAGAGATAAAATGAAACAAGTATGGACGTCCTCCATTGGGATGTACTATTA

EMBOSS_Tri-rbc1 -----AACCTAAATTTGGGTTTATCCGCTAAGAAATACGGTAGAGCAGTTTATGAATGTCTACGCC
HM558487.1 AACCTAAATTTGGGTTTATCCGCTAAGAAATACGGTAGAGCAGTTTATGAATGTCTACGCC

EMBOSS_Tri-rbc1 -----GTGGACTTGAATTTACAAAAGATGATGAAAATGTGAAGCTCCCAACCATTTATGCGTTGGA
HM558487.1 GTGGACTTGAATTTACAAAAGATGATGAAAATGTGAAGCTCCCAACCATTTATGCGTTGGA

EMBOSS_Tri-rbc1 -----GAGACCGTTTCTTATTTTGTGCCGAAAGCTATTTATAAATCACAGGCCGAAACGGGTGNNN
HM558487.1 GAGACCGTTTCTTATTTTGTGCCGAAAGCTATTTATAAATCACAGGCCGAAACGGGTGNNN

EMBOSS_Tri-rbc1 -----TCACGGAAATNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
HM558487.1 TCACGGAAATNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN

EMBOSS_Tri-rbc1 -----TATTTGCAAGAGAAATTTGGGCTTCTTATAGGCCCACTAATGCAAGGACTACCTAACAGGCGG
HM558487.1 TATTTGCAAGAGAAATTTGGGCTTCTTATAGGCCCACTAATGCAAGGACTACCTAACAGGCGG

EMBOSS_Tri-rbc1 -----ATTCACTGCAAAATACTACCCCTGCTCACTATTGCGCGGATAAATGGTCTACTTCTTCATAT
HM558487.1 ATTCACTGCAAAATACTACCCCTGCTCACTATTGCGCGGATAAATGGTCTACTTCTTCATAT

EMBOSS_Tri-rbc1 -----CCACCGTGAAGTGCATGCAATTTATGATAGACAGAAAAATCATGGTATGCACTTTGCGTGT
HM558487.1 CCACCGTGAAGTGCATGCAATTTATGATAGACAGAAAAATCATGGTATGCACTTTGCGTGT

EMBOSS_Tri-rbc1 -----ATTAGCTAAGAGCGTTTACGTTTGTCTGGTGGAGATCATATTCAAGCCGCTACTGTAGTAGG
HM558487.1 ATTAGCTAAGAGCGTTTACGTTTGTCTGGTGGAGATCATATTCAAGCCGCTACTGTAGTAGG

EMBOSS_Tri-rbc1 -----TAAACTTGAAGAGAGAAAGGGAGATAAATTTAGGTTTGTGACTTACTACGATGATGATTA
HM558487.1 TAAACTTGAAGAGAGAAAGGGAGATAAATTTAGGTTTGTGACTTACTACGATGATGATTA

EMBOSS_Tri-rbc1 -----TGTTGAAAAAGATAGAAAGTCGCGGATTTTTTTTCACTCAGGATTGGGTTTCTTTACCGGG
HM558487.1 TGTTGAAAAAGATAGAAAGTCGCGGATTTTTTTTCACTCAGGATTGGGTTTCTTTACCGGG

EMBOSS_Tri-rbc1 -----TGTTCTGCGCTTGTGCTTTCAGGGGATATCCACGTTTGGCATATGCCCGCTCTGACGAGSAT
HM558487.1 TGTTCTGCGCTTGTGCTTTCAGGGGATATCCACGTTTGGCATATGCCCGCTCTGACGAGSAT

EMBOSS_Tri-rbc1 -----TTTTGGAGATGATTCTGTACTTCAATTCGGCGGAGGAACTGTAGGACACCCCTTGGGAGAA
HM558487.1 TTTTGGAGATGATTCTGTACTTCAATTCGGCGGAGGAACTGTAGGACACCCCTTGGGAGAA

EMBOSS_Tri-rbc1 -----TGAC
HM558487.1 TGAC
```

Figure 6. Comparison alignments between *rbcl* marker gene for *Trifolium alexandrinum* Helaly genotype and *rbcl* reference sequence

```
>EMBOSS_Tri-Cox
TCTTTCAGCT AATATTGCC ATAGAGGCC TTCTGTTGAT TTAGCTATTT TTAGATTACA      60
TTTAGCTGGT GTATCATCAA TTTTAGGAGC AATTAATTTT ATTACTACCA TGATTAATAT      120
ACGACCTATT GGTATACAAT TAGATAAACT TCCTTTATTT GCTTGGTCAG TTTTAATTAC      180
TGCTATTTTA CTCTGCTTT CCCTCCCTGT ATTAGCAGGA GCAATCACTA TGCTTTAAAC      240
AGATCGAAAT ATTAATACTT CATTTTTTGA CCCTGCAGGA GGTGGGGAT      289
```

Figure 7. Shows the cytochrome c. oxidase subunit (1) gene (Cox1) marker gene sequence to *Trifolium alexandrinum* Helaly genotype.

(DNA Barcoding of Egyptian clover Helaly Genotype (Cox 1) gene).

Preserve the originality was detected (fig.8) through comparing the cytochrome c. oxidase (C.O.I) gene CDS: sequence of mitochondrial with cytochrome c oxidase

(COI) gene partial cds: reference of mitochondrial sequence and showed completely identical similarity with 100 % of genetic similarity.



Figure 8. Comparison of alignments between cytochrome c oxidase subunit 1 gene for *Trifolium alexandrinum* Helaly genotype and cox 1 reference sequence

It is significant to consider that DNA-based classification by *Trifolium* would be challenging without any accessibility of universal comprehensive monograph & biological data to utmost genus (Gillett and Taylor, 2001).

The results obtained in this study for identification and evaluation of Similarity with the original genetic base are in agreement with the results of Ganopoulos *et al.*, (2012). Who applied barcode of DNA high resolution melting

system employ the global nuclear power plant DNA barcoding area ITS2 for meadow species uniformity, quantification and deception reveals *Medicago lupulina* deceit as low as (1: 100) in *Trifolium pratense* seeds. Their results indicated that [Bar-HRM] analyses could be a faster with extreme resolution and cost-effective replacement method to back up forage and meadow species and quantitatively disclose the purity of their seeds and their feed products. More light was added to our findings as obtained by Gillett and Taylor, (2001). They applied DNA-based identification in *Trifolium* and reported that its potential confronts without availability of an overall international monograph, (Zohary and Heller 1984) biological noticed of ultimate genus.

Efficacy of various genes (cox1, *Rbcl*, 18S and I.T.S. rDNA) were assessed for recognize species of cryptic in the morphospecies model Cox1. divergence was usually much greater than *rbcl* variance and always extremely variable than 18S rDNA. I.T.S. rDNA sequences were significant variable than cox1, but well-known problems with regard to variability of intragenomic caution against its use in identification. More information and less sequencing effort mean that is the cox1. can benefit aid in identification diatom. Advantages of cox1. for crucial phylogenetic relationships between tree topologies were very identical, even though back up values were generally decrease for cox1 (Evans *et al.*,2007). With agreements to our findings, Hawkins *et al.*, (2015) metabarcoding DNA plus melissopalynological fitted to most numerous floral honey compounds and plant Taxa. there were 92% harmony for taxa had plentiful over 20%.

Whereby, when all taxa were comparable, the rate of classification decreased from 2245 in addition there was little agreement among the relative abundance of taxa found using the two techniques. DNA of meta-barcoding given more repeatability 64% taxa compared to 28% melisspalynology.

Altschul *et al.*, (1990) introduced BLAST tool for finding sequence similarity (Basic Local Alignment Tool). BLAST border constancy that optimize a measure of local identification the maximal segment pair score. Such a border may be concept of as decrease the evolutionary distance or maximizing the uniformity between the two sequences compared. BLAST employs a magnitude based on well – defined mutation scores to compare two sequences, whether DNA or amino acid sequences to discover sequence homology. Pairwise alignment is deciding if a pair of sequences is evolutionary related or not. Pairwise uniformity mark for the sequences that be fed into a cluster analyses, or program of tree calculating. The tree program is calculated to place uniformity pairs of sequences closer altogether on the tree than sequences that are less identical.

CONCLUSION

This work aims at discrimination and identifying *Medicago sativa* and *Trifolium alexandrinum* (two important Forage crops) via two DNA bar-coding genes (*rbcl* and *Cox1* genes).

Identification of *Medicago sativa* Rammah 1 genotype was performed through *rbcl* and *Cox1* genes identified it as *Medicago sativa* voucher G00199095 ribulose1, 5 biphosphate carboxylase / oxygenase large subunit gene. (*rbcl*) CDS; chloroplast. Sequence ID: KJ204375.1 and *Medicago sativa* voucher Ahrendsen 23_ for *rbcl* and *Cox1* genes respectively. Moreover, *Trifolium alexandrinum* Helaly genotype was identified as *Trifolium alexandrinum* (Sequence ID: HM850407.1) and *Trifolium alexandrinum* voucher K-016Hv (Sequence ID: KU234213.1) as *rbcl* and *Cox1* genes respectively.

Trifolium alexandrinum showed more success for genetic similarity comparing with [*Medicago sativa*] as output of genetic similitude eminence with origin sequences.

REFERENCES

Aldrich J, B. Cherne, E. Merlin and J. Palmer 1987. Sequence of the *rbcl* gene for the large subunit of ribulose biphosphate carboxylase-oxygenase from alfalfa. *Nucleic Acids Res*, 15 (2): 868.
 Altschul, S.F., W. Gish., W.Miller., E.W.Myers and D. Lipman 1990. "Basic Local Alignment Search Tool", *Journal of Molecular Biology*, Vol. 215 : PP.403-410.

- Badr A, and H. El-Shazly 2001. Molecular approaches to origin, ancestry and domestication history of crop plants: Barley and clover as examples. Journal of Genetic Engineering and Biotechnology. Volume 10 : 1–12.
- Blaxter, M.L.,(2004. The promise of a DNA taxonomy. Philos. Trans. R. Soc. Lond. Biol. Sci. 359, 669–679.
- Ellison N, W. A., A.B. Liston, J.C. Steiner, W. Williams, N.L. Taylor 2006. Molecular phylogenetics of the clover genus (*Trifolium*—Leguminosae), Molecular Phylogenetics and Evolution 39, 688–705.
- Evans M; Wortley AH., Mann DG. 2007. An Assessment of Potential diatom "Barcode" genes (*cox1*, *rbcl*, 18S and ITS rDNA) and their effectiveness in determining Relationships in Taxa Protist, 158(3) : 344 – 364. Erup 207, Jun 19.
- Gael J. A. Kergoat, Alex Delobel, b and Jean-Francois Silvaina 2004. Phylogeny and host-specificity of European seed beetles (Coleoptera, Bruchidae), new insights from molecular and ecological data. Molecular Phylogenetics and Evolution 32, 855–865.
- Ganopoulos I., P. Madesis and A. Tsaftaris 2012. Universal ITS2 Barcoding DNA Region Coupled with High-Resolution Melting (HRM) Analysis for Seed Authentication and Adulteration Testing in Leguminous Forage and Pasture Species. Plant Molecular Biology Reporter, 30(6) : pp 1322–1328
- Gillett JM, and NL. Taylor 2001. The world of clovers. Ames, IA, Iowa State University Press. 457 p
- Gurdon C and P. Maliga 2014. Two distinct plastid genome configurations and unprecedented intraspecies length variation in the *accD* coding region in *Medicago truncatula*. DNA Res. 21(4):417-27.
- Hawkins, J., Devere, N. Griffith, A. Ford, C. Allain guillaume, J. Hegarty, M. Baillic, L. and Adams – Groomy B. 2015. Using DNA metabarcoding to Identify the Floral Composition of Plant Taxa and Honey. A new Tool for Investigating plant and Honey bee. Foraging Preferences – Plos ONE, 10(8). E0134735. Iss N. 1932 6203 Available from : <http://eprints.uwe.ac.uk/25992>.
- Kergoat *et al.*, 2004. The Genus *Trifolium*. Israel Academy of Sciences and Humanities.
- Kress, W.J., K.J. Wurdack, E.A. Zimmer, L.A. Weigt, D.H. Janzen 2005. Use of DNA barcodes to identify Xowering plants. Proc. Natl. Acad. Sci. USA 102, 8369–8374.
- Lavin, M., J.J. Doyle, and J.D. Palmer 1990. Evolutionary significance of the loss of the chloroplast-DNA inverted repeat in the Leguminosae subfamily Papilionoideae. Evolution 44, 390–402.
- Liston, A., 1995. Use of the polymerase chain reaction to survey for the loss of the inverted repeat in the legume chloroplast genome. In: Crisp, M.D., Doyle, J.J. (Eds.), Advances in Legume Systematics, Part 7. Royal Botanic Gardens, Kew, UK, pp. 31–40.
- Madesis P. I., P. Ralli Ganopoulos and A. Tsaftaris 2012. Barcoding the major Mediterranean leguminous crops by combining universal chloroplast and nuclear DNA sequence targets. Genet. Mol. Res. 11 (3): 2548-2558.
- Morris, J.B., and S.L. Greene 2001. Developing a multiple-use germplasm collection for the genus *Trifolium*. Crop Sci. 41, 893–901.
- Newmaster S. G., Email author, Grgruric M. Shanmug Hanandh and S. Ramalingam and S. Ragupathy 2013. DNA barcoding detects contamination and substitution in North American herbal products. *BMC Medicine* 2013:11:222. DOI: 10.1186/1741-7015-11-222.
- Sharawy S and E. Karakishi 2015. Taxonomic Relationships of Some Species of Orobanche L. Evidence from Rapid-pcr and Issr Markers. Pak. J. Bot., 47(2): 437-452.
- Tautz, D., P. Arctander, A. Minelli, R.H. Thomas, and A.P. Vogler 2003. A plea for DNA taxonomy. Trends Ecol. Evol. 18 : 70–74.
- Young ND, F. Debellé, G. Oldroyd, R. Geurts, S.B. Cannon, M.K. Udvardi, V.A. Benedito, K. Mayer, J. Gouzy and H. Schoof 2011. The *Medicago* genome provides insight into the evolution of rhizobial symbioses. Nature 480: 520–524.
- Zohary and Heller, 1984. The Genus *Trifolium*. Israel Academy of Sciences and Humanities. Jerusalem, Israel.

استخدام تقنية DNA Barcoding في عمل البصمة الوراثية لنوعين من محاصيل العلف

عبد العزيز طلعت بندق

قسم بحوث محاصيل العلف ، معهد بحوث المحاصيل الحقلية ، مركز البحوث الزراعية ، الجيزة ، مصر

في هذه الدراسة تم عمل شفرة كودية مميزة لكلاً من البرسيم المسقاوي صنف الهاللي والبرسيم الحجازي صنف رماح 1 وكذلك تم تمييز وتعريف كل من البرسيم المسقاوي صنف الهاللي والبرسيم الحجازي صنف رماح 1 باستخدام تقنية الباركود DNA وهذا عن طريق جينين يستخدمان في إعطاء شفرة كودية مميزة لكل منهما وهما (*rbcl* and *Cox1 genes*). ولقد أظهرت النتائج أن استخدام جين (*rbcl*) في تعريف البرسيم الحجازي صنف رماح 1 بأنه *Medicago sativa* voucher G00199095 (*Sequence ID* : KJ204375.1) وأيضاً أمكن تعريف البرسيم الحجازي صنف رماح 1 باستخدام جين (*Cox 1*) وأوضحت النتائج أنه *Medicago Sativa* Voucher Ahrendsen-23 (*Sequence ID* : HM850407.1) بأنه *Trifolium Alexandrinum* (*Sequence ID* : HM850407.1) وأعطى نسبة تشابه بلغت 91.24% بالمقارنة بالأصل الوراثي (*Sequence ID* : KJ 204375.1) أما بالنسبة للبرسيم المسقاوي صنف الهاللي فقد أظهرت النتائج تعريفه باستخدام جين (*rbcl*) بأنه *Trifolium Alexandrinum* (*Sequence ID* : HM850407.1) وأيضاً أظهرت النتائج تعريفه باستخدام (*Cox 1 Gene*) بأنه (*Sequence ID* : HM850407.1) *Trifolium alexandrinum* voucher K-016 HV (*Sequence ID* : KU234213.1) وأعطى نسبة تشابه 100% بالمقارنة بالأصل الوراثي (*Sequence ID* : HM850407.1) وفي ضوء النتائج المتحصل عليها تم استخدام التشابه الوراثي مع الأصل الوراثي كمقياس للمتماثل الوراثي وأوضحت النتائج التماثل الوراثي العالي للبرسيم المسقاوي صنف الهاللي كنتيجة لتطابقه بنسبة 97.9% مع الأصل الوراثي بالمقارنة بالبرسيم الحجازي رماح 1 الذي بلغت نسبة تطابقه مع الأصل الوراثي 90.6%. وبناء على النتائج المتحصل عليها يصبح من الممكن استخدام تقنية DNA Barcoding في التمييز بين الأصناف والأنواع والأجناس للبرسيم المصري وكذا حمايتها دولياً، وتخلص الدراسة أيضاً إلى أهمية استخدام DNA Barcoding في تحديد الاختلافات الوراثية ودرجات التماثل الوراثي التي تستخدم كأساس في برامج التحسين الوراثي مستقبلاً.