

Determination of Vitamin D₃ Content in High, Low and Zero Fat Food Using High Performance Liquid Chromatography

AHMAD M. FARAG, Ph.D.¹; MAHMOUD S. RIZK, Ph.D.²; HAMDY A. EL-BASSEL, Ph.D.³ and MONA H. YOUSSEF, M.Sc.⁴

The Departments of Organic Chemistry¹, Inorganic and Analytical Chemistry², Faculty of Science, Cairo University, The Department of Nutritional Biochemistry and Metabolism, National Nutrition Institute³ and The Department of Food Science, National Nutrition Institute⁴, Cairo

Abstract

Background: Vitamin D, has a significant role in bone metabolism and helps calcium absorption in the body. There are only few vitamin D assay methods available for zero fat food.

Aims of Study: (1) To develop an accurate and sensitive LC method for the quantification of vitamin D₃ in food by optimization of each step of the analytical method: Extraction, sample preparation, separation and detection. (2) To validate the developed method. (3) To apply the method to quantify the total vitamin D₃ in food from several species.

Material and Methods: In this study, a rapid, simple, and economical reversed phase liquid chromatographic method was described for the determination of vitamin D₃ in some high, low and zero fat samples (milk products, cereal and chewing gum samples). The isolation of fat soluble vitamins includes a saponification step and an extraction step with petroleum ether and diethyl ether. The vitamin D₃ content of samples was determined by reversed phase liquid chromatography. Ultra violet-Visible (UV-VIS) detector and C18 column were used for this purpose.

Results: The linearity of standard curves of vitamin D₃ were 10-200 µg/mL expressed of the correlation coefficient $r^2 = 0.9992$. The detection (LOD) and quantification (LOQ) limits were 5.09 and 15.42 µg/mL, respectively. The accuracy was 101.37 ± 4.37 .

Conclusion: The described reversed-phase HPLC method is favorable compared with other published HPLC-UV methods (20 and 21) because of its stability-indicating nature, short run time and wide analytical range with outstanding linearity, accuracy and precision. The proposed method allows the determination of vitamin D₃ in a single chromatographic run and is suitable for the analysis of the stability of vitamin D₃. The obtained results from the assay of vitamin D₃ in commercial nutrition supplements confirmed that the method is appropriate for the routine analysis of various food samples.

Key Words: Analytical measurement – Dietary supplements – Fortified foods – Infant – Vitamin D.

Correspondence to: Dr. Ahmad M. Farag, The Department of Organic Chemistry, Faculty of Science, Cairo University

Introduction

VITAMIN D is very important fat soluble vitamin in human and animal diet. It exists in two forms, Vitamin D₂ and D₃. Vitamin D₃ (cholecalciferol) is synthesized endogenously from 7 – dehydrocholesterol after ultraviolet irradiation or is absorbed from the diet [1]. Vitamin D plays an important role in the maintenance of normal levels of calcium and phosphorus in the blood stream and is essential for the proper development and maintenance of bone [2]. Both vitamins D₂ and D₃ are biologically inactive. In humans, they are metabolized in the liver to calciferol (25-hydroxyvitamin D₂ and 25-hydroxyvitamin D₃, collectively known as 25(OH) D), Vitamin D that is not metabolized in the liver is stored in the adipose tissue and skeletal muscle, then released during vitamin D deprivation [3]. It has also been suggested that vitamin D₃ is more efficiently absorbed by the intestine than vitamin D₂ [4]. Vitamin D is soluble in lipids. Therefore the amount of fat in a meal may affect bioavailability with potentially higher vitamin D absorption from consumption in a higher fat food or meal [5].

Vitamin D deficiency is not only related to muscle weakness and osteomalacia, but has also been associated with cardiovascular disease, cancer, autoimmune diseases, diabetes mellitus and hypertension [6]. Furthermore, newborn babies may be affected in their normal growth and development, putting them at risk of long-term physical deficits [7].

HPLC methods offer the best approach to accurate determination of vitamin D₃ content in foods and pharmaceuticals, as well as stability testing. In the last decade, high-performance liquid chromatography coupled to mass spectrometry has

become the technique of choice for vitamin D₃ determination in foods, feeds and pharmaceuticals [8].

Vitamin D chemistry:

Vitamin D exists in a number of forms, where the major physiologically relevant forms are vitamin D₂ (ergocalciferol) and vitamin D₃ (cholecalciferol) (Fig. 1) [9]. Vitamin D₂ is produced by ultraviolet B (UVB) irradiation of the plant steroid ergosterol. Vitamin D₃ is synthesised in the skin of vertebrates through the action of UVB and 7-dehydrocholesterol. Vitamin D₂ is the less common form of vitamin D and has potentially lower bioavailability than Vitamin D₃ [10]. Vitamin D from sunlight or dietary sources is biologically inactive and is hydroxylated, via a two step process to 1,25-dihydroxyvitamin D (calcitriol), before it becomes metabolically active [9].

Vitamin D two major forms, cholecalciferol (vitamin D₃) and ergocalciferol (vitamin D₂).

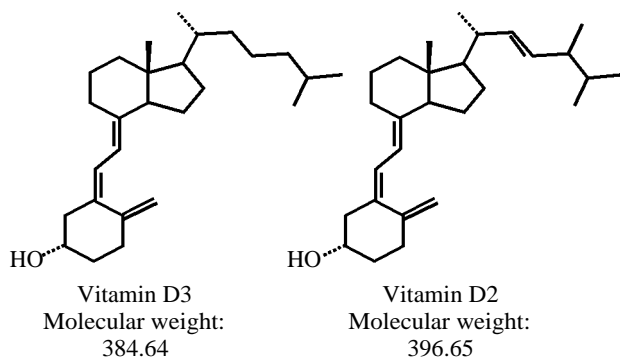


Fig. (1): Different forms of vitamin D.

Material and Methods

Samples selection:

In January 2017 In January 2017, products were purchased for this pilot study from the local supermarkets in Cairo governorate, Egypt, the selection included the majority of the available brands, and all products were brought within the best-before date. To study the potential difference in vitamin D content between labeled and measured concentration, triple of products of the same brand but each with a different best-before date were brought.

According to the South African Food Data System (SAFOODS), full cream milk contains at least 3.3g of fat per 100ml of which 2.1 g is saturated fat. Low fat contains not more than 1.9g of fat per 100ml of which 1.2g is saturated. On the other hand fat-free or skim milk contains not more than 0.2g of fat per 100ml and only a trace of saturated fat.

Vitamin D fortified foods were classified in the following two product groups:

- High fat food samples: Were included in baby milk powder and creamy cooking cheese.
- Low and Skimmed food samples: Low and zero fat food samples were selected, low fat samples included yogurt and baby food based on cereals (cerelac). While, zero fat vitamin D₃ food supplement in chewable forms were include in baby gum.

Labeled vitamin D concentration:

The labeled vitamin D concentration ($\mu\text{g}/100\text{gm}$) was obtained by the manufacturer's label declaration for comparison with the analyzed vitamin D concentration.

It is, however, not always possible for food or supplements to contain the exact micronutrient level specified on the label, due to natural and processing variations, as well as changes during storage. But on the other hand substantial deviation from what is labeled could mislead the consumer and should be prevented [11]. The actual vitamin D concentration of fortified foods and dietary supplements may deviate from the label due to potential overages to cover losses during shelf life [12].

Chemical analysis (Reagents and materials):

All materials and reagents used in this study were pure laboratory chemicals. Potassium hydroxide pellets, sodium sulphate anhydrous, ascorbic acid were purchased from Merck. The organic solvents used include, ethanol, methanol, petroleum ether (40-60°C), acetonitrile and diethyl ether peroxide free. All the used chemicals were of the highest purity available analytical grade and all solvents in this study were grade-HPLC and obtained from Merck, fluka, Fisher, sigma Aldrich Companies.

Experiments were carried out using a GBC HPLC system coupled to UV-VIS detector using single wavelength for this study, 264nm for Vitamin D₃. The compounds were separated with reversed phase (250mm x4.6mm, 5 μm) ACE 5 C18-300 column with a C 18 guard cartridge which maintained at room temperature. Mobile phase was (50%: 50%, v/v) methanol and acetonitrile HPLC grade and the flow rate was 1.00mL/min (isocratic elution). Total run time required was equal to 7 min.

Standard solutions:

Stocks and standards solution of Cholecalciferol (vitamin D₃): Stock solution was prepared by dissolving 1mg of cholecalciferol standard in 1ml

methanol. Serial standard solutions were prepared by using 20, 50, 100, 150 and 200 μ l of the stock standard solution in 10ml measuring flask then complete to the mark by methanol. The stock and standard solutions were stored in brown flask with screw cap to avoid exposure to light and air at -4°C in refrigerator.

Saponification:

Saponification step of high fat food: Five grams of powdered or 5ml liquid sample weighted in 10ml measuring flask then complete to the mark by warm deionized water at 40°C and mixed for 10min by vortex until complete homogenization occurred then 20ml 50% KOH solution and 1gram of ascorbic acid were added in 100ml measuring flask with covered foil and complete to the mark by ethanol with continuously shaking through over night in water bath at room temperature until the sample was completely saponified.

Saponification step of low and zero fat food: Five grams of powdered or 5ml liquid sample, 1 gram of rice bran oil (put in 100ml volumetric flask then add 20ml 50% KOH solution and one gram of ascorbic acid added and completed by ethanol to mark. The solution was covered by aluminum foil with continuously shaking through over night in water bath at room temperature until the sample completely saponified) and complete as the first saponification step.

Extraction step for high and low fat and non fat food: After shaking, the sample was transferred into 500ml separating funnel then add 50ml of (50%: 50%, v/v) diethyl ether: Petroleum ether, shake the mixture for 20min. The upper and organic layer was left in the same separating funnel. The aqueous layer was re-extracted with a further 50ml of the mixture and the mixture was shaken for 20 min in another separating funnel. The two organic

layers were joined and washed twice by shaking 2min with 250ml of deionized water. The organic layer washed until no change in red pH paper. The organic layer was collected and evaporated by furnace at 40°C . For reversed phase chromatography the residue dissolved in 2 ml of methanol for determination of vitamin D₃ was directly filtered through 0.45 μ m filter and 20 μ l of extract was injected into the HPLC system. The vitamin D₃ concentration in the samples was calculated in $\mu\text{g}/100\text{g}$ edible weight using peak area by comparison between standard and sample after injection.

Determination of fat crud:

0.5-2 g sample was weighted in 250ml volumetric flask, adding 10ml of distilled water, 1.5ml ammonium hydroxide was added then the flask was immersed on water bath at 70°C for 15min then cooled. 10ml of concentrated hydrochloric acid was added. The flask was heated on hot plate for 5 min then cooled. 25ml petroleum ether, 25ml diethyl ether and 10ml ethyl alcohol 95% were added and mix them and leave for 24 hour. Solvent with fat was separated in beaker (known weight). The beaker was leaved in the oven at 50°C . The beaker was put in desiccator until cool then the beaker weighted with fat.

- Weight of fat = Weight of beaker with fat – Weight of beaker

- % Crude fat = (Weight of fat / Weight of sample) x 100

Results

Table (1) shows the measured and labeled fat content in high fat food (represented by creamy cooked cheese and baby infant milk powder), low fat food (represented by yogurt and baby food based on cereals) and zero fat food (represented by baby gum).

Table (1): Determination of crud fat and comparison with label information.

Fat g/100 g	Label concentration g/100g	Found concentration g/100g	Mean	SD	RSD %	Recovery %
1- Fat in (creamy cooked cheese) cheese	25	24.08 24.14 24.75	24.32	0.37	1.52	97.29
2- Fat in baby infant milk powder	26.1	25.81 25.37 25.74	25.64	0.24	0.92	98.24
3- Fat in yogurt	3	2.83 2.59 2.70	2.71	0.12	4.44	90.22
4- Fat in baby food based on cereals	0.55	0.47 0.51 0.46	0.48	0.026	5.51	87.27
5- Fat in baby gum	0	0	0	0	0	0

The number of vitamin D fortified foods has increased. The food type with the greatest increase was yoghurt, Fortified skim milks and reduced fat milks [13]. Fortification can be achieved in multiple ways with varying efficiency. For example, vitamin D₃ was added by: Addition of water-soluble emulsion, or crystalline liposoluble vitamin D [14]. Vitamin D₃ was fortified into yoghurt in either a crystalline or emulsified form; both forms of vitamin D₃ were stable in yoghurt during storage for the expected shelf lives of the products [15]. In an evaluation of increasing the level of vitamin D₃ fortification in high temperature processed reduced fat milks and low-fat yoghurt serving, no loss of vitamin D₃ during processing was found [16]. Vitamin D₃ has also been shown to be stable in a non-fat food, with no change [17]. Study by [18] strongly recommend food industries to use opaque containers for storage of yogurt products or even any kinds of foodstuffs fortified with Vitamin D₃ to reduce light-related degradation of the vitamin during the products shelf-life.

Calibration curve of vitamin D₃:

The linearity of standard curves (Table 2) was expressed in terms of the determination coefficient (*r*²) from plots of the integrated peak area versus concentration of the standard (µg/ml). These equations were obtained over a wide concentration range, in accordance with the levels of these compounds found in food samples. Linear equations were found, with satisfactory linearity (*r*² > 0.99).

Table (2): Vitamin D₃ Calibration curve analysis.

Concentration µg/ml	Beak area	Found concentration µg/ml	Recovery%
20	111	20.84827	104.2413
50	286	49.51563	99.03126
100	579	97.51298	97.51298
150	920	153.3734	102.2489
200	1197	198.7497	99.37486

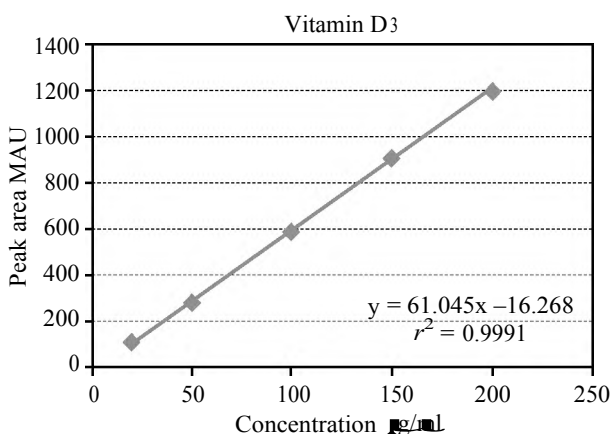


Fig. (2): Calibration curve of vitamin D₃.

Validation of vitamin D₃:

Validation studies were performed by measuring basic parameters such as precision, accuracy, linear region, limits of detection (LOD) and quantification (LOQ), and recovery.

Table (3): Validation sheet of vitamin D.

Parameter	Values
Accuracy	100.48±2.71
Slope	6.10
Intercept	-16.27
Linear Range	20-200 (µg/ml)
Correlation Coefficient (<i>r</i>)	0.9991
Standard Error	13.28
Variance	6.10
LOD	6.53
LOQ	21.76

Quality control chart of vitamin D₃:

10 samples of standard reference material® 1849 a prepared by this method.
 - Mean = 1.11 µg/100g, SD = 0.085

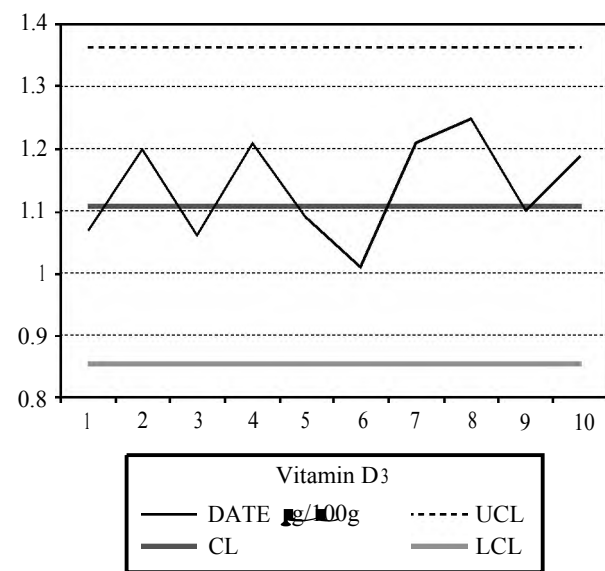


Fig. (3): Quality control chart of vitamin D₃.

Vitamin D₃ in Creamy cooked cheese:

Fig. (4) shows measured vitamin D₃ concentration (µg per 100g) in creamy cooked cheese compared to the vitamin D content declared on the label, the measured vitamin D₃ content ranged between 87.5% and 96.4% of the declared value.

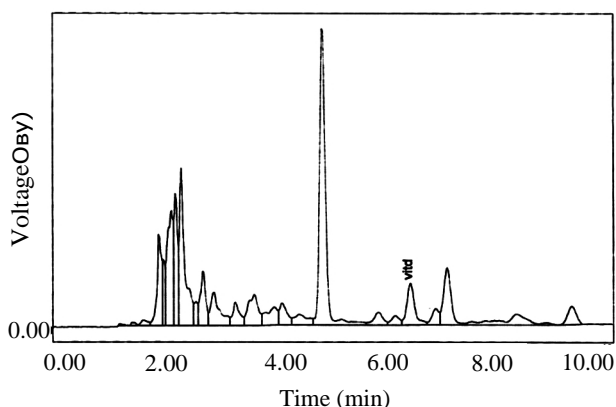


Fig. (4): Vitamin D₃ in creamy cooked cheese.

Vitamin D₃ in Baby infant milk powder:

Fig. (5) shows measured vitamin D₃ concentration (g g per 100g) in baby infant milk powder compared to the vitamin D content declared on the label, the measured vitamin D₃ content ranged between 91.5% and 96.2% of the declared value.

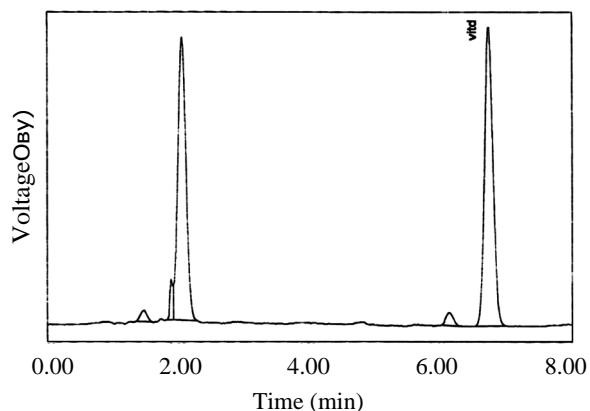


Fig. (5): Vitamin D₃ baby in infant milk powder.

Vitamin D₃ in Yogurt:

Fig. (6) shows measured vitamin D₃ concentration (g g per 100g) in yogurt compared to the vitamin D content declared on the label, the measured vitamin D₃ content ranged between 82.3% and 89% of the declared value.

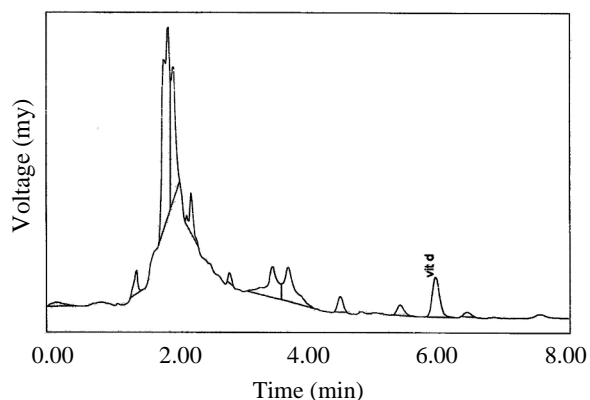


Fig. (6): Vitamin D₃ in yogurt.

Vitamin D₃ in Baby food based on cereals:

Fig. (7) shows measured vitamin D₃ concentration (g g per 100g) in yogurt compared to the vitamin D content declared on the label, the measured vitamin D₃ content ranged between 91.4% and 96.4% of the declared value.

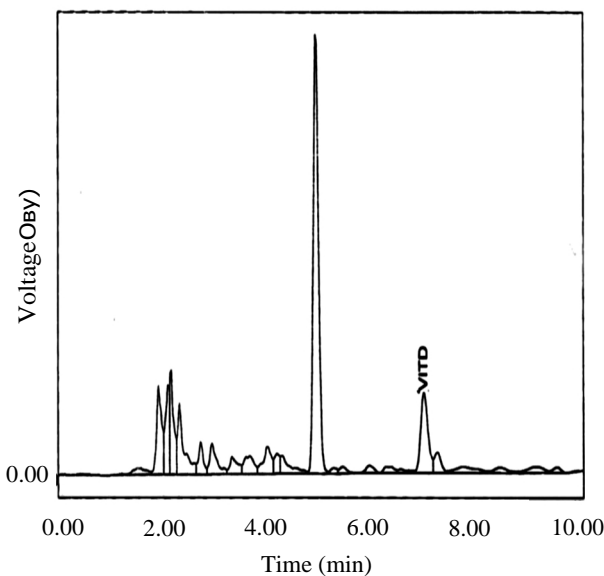


Fig. (7): Vitamin D₃ in baby food based on cereals.

Vitamin D₃ in Baby gum:

Fig. (8) shows measured vitamin D₃ concentration (g g per 100g) in herba land kids gummy compared to the vitamin D content declared on the label, the measured vitamin D₃ content ranged between 93.5% and 96.5% of the declared value.

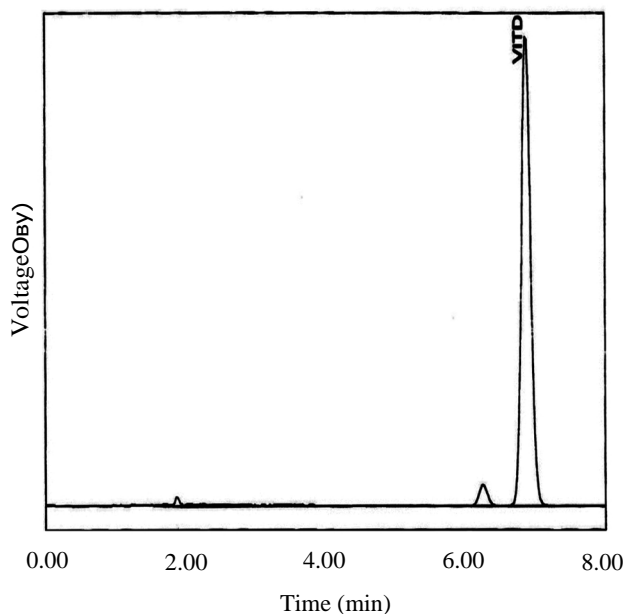


Fig. (8): Vitamin D₃ in baby gum.

Table (4): Overview of labeled and measured vitamin D₃ contents in fortified foods and dietary supplements.

Vitamin D ₃ μg per 100 g	Label concentration $\mu\text{g}/100\text{g}$	Found concentration $\mu\text{g}/100\text{g}$	Mean	SD	RSD %	Recovery %
1- Vitamin D ₃ in creamy cooked cheese	2.5	2.18 2.32 2.41	2.32	0.116	5	92.8
2- Vitamin D ₃ in baby infant milk powder	10	9.62 9.58 9.15	9.45	0.26	2.76	94.5
3- Vitamin D ₃ in yogurt	3	2.67 2.47 2.52	2.55	0.104	4.08	93.33
4- Vitamin D ₃ in baby food based on cereals	5	4.61 4.57 4.82	4.67	0.134	2.88	90.59
5- Vitamin D ₃ in baby gum	170	165 159 164	154	3.53	2.96	

Discussion

The measured vitamin D concentrations were compared with the labeled values. The products for which the measured vitamin D value deviated significantly from the declared value were re-analysed

Figs. (4-8) and Table (4) show the concentrations of vitamin D₃ in food samples (creamy cooked cheese, baby infant milk powder, yogurt, baby food based on cereals and baby gum, it was found to be 2.32 ± 0.12 , 9.45 ± 0.26 , 2.55 ± 0.10 , 4.67 ± 0.13 and 154 ± 3.53 $\mu\text{g}/100\text{g}$ respectively and label vitamins content were 2.5, 10, 3, 5 and 170. The recoveries rang 90.6%-94.5%. The relative standard deviations were 2.8-5%.

In this study, some parameters were changed in order to recommend a specific method for determination of Vitamin D₃ in high, low and zero fat content food in the both extraction and quantification stages.

In the saponification step, previous studies mainly have focused on the effect of varied temperature and time with the most confirming 70-85°C and 25 to 30min conditions [19].

However, some studies stated that using high temperature water bath can negatively affect the saponification process [20,21]. In this study, warm deionized water at 40°C and mixed for 10min. was applied.

In the extraction step, Formation of intractable emulsion layer in this step was considered as a common problem in this step, the substitution of hexane with a more polar solvent such as ethyl

ether and petroleum ether, detained emulsion formation using the wide range of food matrices.

During the washing step, ethanol addition to the distilled water used for washing solvents, which had a significant impact on separation which confirmed the results of [20] who mentioned the positive impact of ethanol in stabilizing the solution state of solvents. In the post extraction treatment, the results of this study in terms of using micro-column, confirmed the results [23] stated that solid phase extraction helped distinguishing two types of vitamin D₂ and D₃.

As another changing variable in this study, changing the ratio, type and amount of solvent that was used in SPE had significant effects on the performance of this treatment.

Recent studies in the field recommend micro-extraction as a comparable method to the classic assay which was used in the present study.

In HPLC determination of vitamin D₃, it was shown in this study that substitution of the mobile phase was (50%: 50%, v/v) methanol and acetonitrile instead of the pure methanol, led to better separation of vitamin D₂ and D₃ peaks, though, the vitamin D₃ retention time became longer.

Structural similarity between vitamin D₂ and D₃ could interfere with the vitamin D₃ calculation as D₂ can appear in the chromatogram with the same retention time as vitamin D₃, when the applied protocol does not allow to differentiate the two components from together. Investigating the vitamin D groups in foods, [22] reported that vitamin D₂ may appear along with vitamin D₃ peak leading to false positive results. The observed peak in Figs.

(4-8) 7min correspond to vitamin D₃ which is higher than vitamin D₂ at 6min. This confirms the results of Mattila in that using SPE column a clear differentiation was observed with about 2 minutes time difference in vitamin D₂ and vitamin D₃ retention time. This confirms the results of Mattila in that using SPE column a clear differentiation was observed with about 2 minutes time difference in vitamin D₂ and vitamin D₃ retention time.

Comparing the claimed and experimentally obtained values of vitamin D₃, less than 18% difference was observed for all samples. These results are satisfactory considering the vitamin D content tends to decrease with time and changes in storage conditions.

Recommendations:

Based on the obtained result, application of some changes in the general vitamin D₃ measurement method can result in a more efficient assay with adequate accuracy for vitamin D₃ in fortified food.

References

- SUBODH K., DIWESH C. and ASHOK K.: An Improved and Sensitive Method for Vitamin D₃ Estimation by RP-HPLC. *Pharm. Anal. Acta.*, 6: 8, 2015.
- WEAVER, C. and FLEET J.: Vitamin D requirements: current and future. *Am. J. Clin. Nutr.*, 80: 1735S-9S, 2004.
- Institute of Medicine, Food and Nutrition Board: Dietary Reference Intakes for Calcium and Vitamin D. Washington, DC: National Academy Press, 2010.
- LAMBERG-ALLARDT C.: Vitamin D in foods and as supplements. *Prog. Biophys. Mol. Biol.*, 92 (1): 33-38, 2006.
- TRIPKOVIC L., LAMBERT H., HART K., SMITH C. and BUCCA G.: Comparison of vitamin D₂ and vitamin D₃ supplementation in raising serum 25-hydroxyvitamin D status: A systematic review and meta-analysis. *Am. J. Clin. Nutr.*, 95 (6): 1357-1364, 2012.
- ROSS A., TAYLOR C., YAKTINE A. and DEL VALLE H.: Dietary reference intakes for calcium and vitamin D. Washington (DC): Institute of Medicine Committee to Review Dietary Reference Intakes for Vitamin D, Calcium, p6, 2011.
- PRINCIPI N., BIANCHINI S., BAGGI E. and ESPOSITO S.: Implications of maternal vitamin D deficiency for the fetus, the neonate and the young infant. *Eur. J. Nutr.*, 52 (3): 859-67, 2013.
- SCHADT H., GOSSL R., SEIBEL N. and AEBISCHER C.: Quantification of vitamin D₃ in feed, food, and pharmaceuticals using high-performance liquid chromatography/tandem mass spectrometry; *Journal of AOAC International*, 95 (5): 1487-1494, 2012.
- TRUSWELL A.: *Essentials of human nutrition*. Oxford: Oxford University Press, 1998.
- BORRADALE D. and KIMLIN M.: Vitamin D in health and disease: an insight into traditional functions and new roles for the 'sunshine vitamin'. *Nutrition Research Reviews*, 22: 118-136, 2009.
- GARG S., SABRI D., KANJI J., RAKKAR P., LEE Y. and NAIDOO N.: Evaluation of vitamin D medicines and dietary supplements and the physicochemical analysis of selected formulations. *The Journal of Nutrition, Health and Aging*, 17 (2): 158-161, 2013.
- VERKAJK J., SEVES S. and OCKÉ M.: Vitamin D concentrations in fortified foods and dietary supplements intended for infants: Implications for vitamin D intake. *Food Chemistry*, 221 (15): 629-635, 2017.
- ENGELSEN O.: The relationship between ultraviolet radiation exposure and vitamin D status. *Nutrients*, 2(5): 482-495, 2010.
- BANVILLE C., VUILLEMARD C. and LACROIX C.: Comparison of different methods for fortifying Cheddar cheese with vitamin D. *International Dairy Journal*, 10 (5-6): 375-382, 2000.
- KAZMI S., VIETH R. and ROUSSEAU D.: Vitamin D₃ fortification and quantification in processed dairy products. *International Dairy Journal*, 17 (7): 753-759, 2007.
- HANSON A. and METZGER L.: Evaluation of increased vitamin D fortification in high-temperature, short-time-processed 2% milk, UHT-processed 2% fat chocolate milk, and low-fat strawberry yogurt. *Journal of Dairy Science*, 93 (2): 801-807, 2010.
- BIANCUZZO R., YOUNG A., BIBULD D. and AMERI A.: Fortification of orange juice with vitamin D₂ or vitamin D₃ is as effective as an oral supplement in maintaining vitamin D status in adults. *American Journal of Clinical Nutrition*, 91 (6): 1621-1626, 2010.
- TINA J., SHAGHAYEGH H., ELHAM F. and AZIZ A.: Stability of Vitamin D₃ in fortified yoghurt and yoghurt drink (Doogh). *Adv. Biomed Res.*, 5: 52, 2016.
- JAKOBSEN J., CLAUSEN I., LETH T. and OVESEN L.: A new method for the determination of vitamin D₃ and 25-hydroxyvitamin D₃ in meat. *J. Food Compos. Anal.*, 17 (6): 777-787, 2004.
- MATTILA P., PIIRONEN V., BÄCKMAN C., ASUNMAA A., UUSIRAUVA E. and KOIVISTOINEN P.: Determination of vitamin D₃ in egg yolk by high-performance liquid chromatography with diode array detection. *J. Food Compos. Anal.*, 5 (4): 281-90, 1982.
- THOMPSON J., HATINA G., MAXWELL W. and DUVALL S.: High performance liquid chromatographic determination of vitamin D in fortified milks, margarine, and infant formulas. *J. AOAC Int.*, 65 (3): 624-31. 14, 1982.
- KATHERINE M., WAYNE R. and JAMES M.: Liquid chromatography with ultraviolet and dual parallel mass spectrometric detection for analysis of vitamin D in retail fortified orange juice. *Journal of Food Composition and Analysis*, 24: 299-306, 2011.
- QIAN H. and SHENG M.: Simultaneous determination of fat-soluble vitamins A, D and E and pro-vitamin D₂ in animal feeds by one-step extraction and high performance liquid chromatography analysis. *J. Chromatogr.*, 825 (2): 127-33, 1998.

تحديد محتوى فيتامين د₃ فى الأطعمة الغنية ، المنخفضة والخالية من الدهون باستخدام كروماتوجرافيا السوائل عالية الأداء

أجريت هذه الدراسة لاستحداث طريقة أكثر دقة وسهولة من الطرق السابقة لتقدير فيتامين د₃ باستخدام كروماتوجرافيا السوائل عالية الأداء فى الأغذية المدعمة بفيتامين د₃.

وقد شملت الدراسة اختيار أغذية عالية الدهون والتي شملت حليب الأطفال المجفف والجبن الدسم المطبوخ، أغذية منخفضة الدهون والتي شملت الزيادى وأغذية الأطفال التى أساسها الحبوب. كما شملت الدراسة أيضاً تقدير فيتامين د فى الأغذية خالية الدهون والمدعمة بفيتامين د₃ كطريقة جديدة وقد تم اختيار لعقة الأطفال الخالية من الدهون والمدعمة بفيتامين د₃ لهذه الدراسة.

فى هذه الدراسة تم تعديل فى بعض مراحل التحديد الكمى لفيتامين D₃ فى الأغذية عالية ومنخفضة الدهون كالتالى :

مرحلة التصبن : تم تقليل درجة الحرارة والوقت ليصبح ٤٠م والمدة ١٠ دقائق بدلاً من ٧٠-٨٥م ولمدة ٣٠ دقيقة فى الطرق السابقة.

مرحلة الاستخلاص : تم استبدال الهاكسين بمذيب أكثر قطبية وهو الأثير البترولى أو الأثير الأيثيلى.

- تم أيضاً تغيير نسبة ونوع وكمية المذيب المستخدم فى مرحلة الاستخراج الصلب حيث تم استخدام الميثانول مع اسيتونيتريل بنسبة ١:١ بدلاً من الميثانول لوحده فقط.

- فى مرحلة الغسيل : تم إضافة الإيثانول إلى الماء المقطر المستخدم فى غسل المذيبات.

ولقد ساهمت هذه التعديلات فى استخدام هذه الطريقة لتقدير فيتامين د₃ فى الأغذية الخالية من الدهون. كما أدى إلى فصل أفضل من قمم فيتامين D₂ و D₃، كما أن هذه التعديلات أصبح مدة فصل فيتامين د₃ من ٦-٧ دقائق بدلاً من ١٢-١٥ دقيقة.

على ضوء هذه النتائج فإن هذه الطريقة مناسبة للتحليل الروتينى للعينات الغذائية المختلفة لتحديد فيتامين D₃.