

EFFECT OF OZONE THERAPY ON ISCHEMIC REPERFUSION INJURY OF THE HEART AND VASCULAR REACTIVITY IN ADULT MALE DIABETIC ALBINO RAT

By

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ABSTRACT

Background: Ozone therapy is a form of complementary medicine treatment that aims to increase the amount of oxygen to the body through the introduction of ozone into the body. **Objective:** Studying the effects of diabetes mellitus, insulin and ozone on ischemic reperfusion (IR) injury on the heart and vascular reactivity in diabetic rat. **Material and Methods:** Two hundred and fifty adult male albino rats of local strain, weighing $120-150 \pm 10$ grams each, were used in this investigation and divided into:

Group I (Normal control group): 10 rats. Group II (Diabetic group -240 rats) were subdivided into 4 subgroups: Group II-a (Diabetic non-treated group - 40 rats), Group II-b (Diabetic insulin-treated group - 40 rats, and Group II-c (Diabetic ozone-treated group - 80 rats Group II-d (Diabetic ozone and insulin-treated group - 80 rats): Rats were submitted to ozone therapy with concomitant treatment with insulin. Supernatant serum was collected in a dry clean tube for estimation of fasting serum glucose, serum total cholesterol, triglycerides, HDL, LDL, LDH, catalase enzyme, glutathione peroxidase, and SOD. Rat aortic rings preparation were used for estimation of changes in vascular reactivity in response to norepinephrine (10^{-5}), vasopressin (10^{-6} M), indomethacin (10^{-6} M) and relaxation of aortic rings (precontracted by NE (10^{-5}) in response to ACh (10^{-6}) and Na^+ nitroprusside (10^{-6}) as estimated in different groups. The organ bath was washed out three times with fresh Krebs' solution before the next substance was added and the rings were allowed to stabilize for 1 hour. All results were presented as the mean \pm SEM. The data were analyzed using SPSS program version 12. For comparison of statistical significance between different groups, a one way ANOVA with the post hoc of Tukey's multiple comparison test was used. A value of $P \leq 0.05$ was considered statistically significant. **Results:** Ozone therapy caused significant decrease in fasting glucose, total cholesterol, triglycerides, LDL, and lactate dehydrogenase "LDH", and significant increase in HDL and myocardial antioxidants (catalase, superoxide dismutase "SOD" and glutathione peroxidase). There were a significant increase in cardiac contractility and heart rate during pre-ischemic and ischemic periods. There was a significant decrease in heart rate accompanied by significant increase in cardiac contractility during reperfusion period. Also, ozone treatment produced a significant decrease in vascular reactivity of aortic rings to norepinephrine, vasopressin and indomethacin, with significant increase in percentage of relaxation to acetyl choline "ACh". **Conclusion:** Diabetic complications are attributed to the oxidative stress in the body. Ozone activates the antioxidant system affecting the level of glycemia. Ozone prevents oxidative stress by normalizing the organic peroxide levels by activating superoxide dismutase.

Key words: Diabetes mellitus, insulin, ozone treatment, ischemic reperfusion, vascular reactivity.

INTRODUCTION

In spite of the current optimal therapy, the mortality of patients with

ischemic heart disease (IHD) remains high, particularly in cases with diabetes mellitus as a co-morbidity. Myocardial lipid metabolism, inflammation, oxidative

stress, myocardial fibrosis, myocardial apoptosis and mitochondrial damage are considered possible mechanisms for the development and progression of diabetic cardiomyopathy "DCM" (Wang et al., 2011).

Increased production of *reactive oxygen species (ROS)* seems to be an important biochemical modification in some pathological events that cause complications accompanying diabetes, e.g. atherosclerosis, ischemic heart disease, nephropathy, pulmonary disease, and fatty liver (Baynes, 1991). High glucose level can stimulate free radical production. Weak defence system of the body becomes unable to counteract the enhanced ROS generation. As a result, condition of imbalance between ROS and their protection occurs which leads to domination of the condition of oxidative stress (Pandey et al., 2010). Certain amount of oxidative stress/ROS is necessary for the normal metabolic processes since ROS play various regulatory roles in cells (Gomes et al., 2012).

Ozone (O₃) therapy can activate the antioxidant system by influencing the level of antioxidant enzymes and some markers of endothelial cell damage (Martínez-Sánchez et al., 2005).

The present work aimed to study the effects of diabetes mellitus, insulin and ozone on ischemic reperfusion injury in the heart and vascular reactivity in streptozotocin (STZ)-induced adult diabetic male rat model.

MATERIALS AND METHODS

Animals:

Two hundred and fifty adult male albino rats of local strain, weighing 120-

150 ±10 grams each, were used in this investigation. Rats were kept (each three in cage; 30× 30 × 30 cm) on a standard laboratory diet and water throughout the study period. Rats were divided into:

Group I (Normal control group): 10 rats.

Group II (Diabetic group -240 rats) were subdivided into 4 subgroups as follows:

Group II-a (Diabetic non-treated group - 40 rats).

Group II-b (Diabetic insulin-treated group - 40 rats): Rats were submitted to insulin treatment. Mixtard insulin was injected subcutaneously in a dose of 0.75 IU/100 gm B.W. once daily.

Group II-c (Diabetic ozone-treated group - 80 rats): Rats were submitted to ozone/oxygen mixture through rectal cannulae every day for 15 days at doses of 1 ml/kg. Ozone was produced using a medical ozone generator and used immediately after it has been generated.

Group II-d (Diabetic ozone and insulin-treated group - 80 rats): Rats were submitted to ozone therapy with concomitant treatment with insulin.

Methods:

- * After 12 hours fasting, blood samples were collected from the retro-orbital venous plexus of each rat, using a fine heparinized capillary tube introduced into the medial epicanthus of the rat's eye (Schermer, 1968). Two milliliters of blood were collected in a graduated centrifuge tube, left for clotting at room temperature in a water bath for 15 minutes, and then centrifuged at 3000 r.p.m for 15 minutes. The super-

nantant serum was collected in a dry clean tube for:

Estimation of fasting serum glucose (**Trinder, 1969**).

Estimation of serum total cholesterol and triglycerides (**Trinder, 1969**).

Estimation of serum high density lipoprotein (HDL - **Burstein, 1970**).

Estimation of serum low density lipoprotein (LDL - **Friedwald et al. (1972)**).

Estimation of lactate dehydrogenase (LDH) concentration (**Scientific Committee, 1982**).

Estimation of catalase enzyme activity(**Goth, 1991**).

Estimation of glutathione peroxidase (GPx) activity(**Hafeman et al., 1974**).

Estimation of superoxide dismutase activity (SOD - **Beauchamp and Fridovich, 1971**).

- * Hearts from different groups were excised and immediately placed in ice-cold heparinized normal saline. Ascending aorta was cannulated, and retrograde perfusion of the nonworking heart (Langendorff's method) was initiated with modified Krebs-Henseleit (KH) buffer solution maintained at 37°C. The perfusate was aerated with carbogen equilibrated at pH 7.4. After 20 minutes of perfusion and stabilization, perfusate flow was then stopped, creating a state of global ischemia, which was maintained for 40 minutes at 37°C. Hearts were then allowed to reperfuse for 30 minutes during which some hemodynamic parameters were tested (**OferMerin et al., 2007**), and perfusate was collected

for assaying lactate dehydrogenase enzyme (LDH) release. The heart was perfused at a controlled constant flow rate of 8 ml/ minute.

- * Thoracic aorta was cut through near the heart as possible, and dissected free as far as the diaphragm. The aorta was then transferred to a Petri-dish containing Krebs' solution at room temperature aerated with carbogen and then cut into 2.5– 3 -millimeter rings. Aortic rings were then suspended in a 10 ml organ bath, containing the freshly prepared Krebs' solution maintained at 37°C and continuously bubbled with carbogen gas. The preparations were attached to a force transducer, and isometric tension was recorded on a polygraph (**Shin et al., 2006**). Aortic rings were allowed to equilibrate for 60 minutes. A resting tension of 1 g was maintained throughout the experiment. The rat aortic ring preparation was used for estimation of changes in vascular reactivity in response to norepinephrine (10^{-5}), vasopressin (10^{-6} M), indomethacin (10^{-6} M) and relaxation of aortic rings (precontracted by NE (10^{-5}) in response to ACh (10^{-6}) and Na^+ nitroprusside (10^{-6}) as estimated in different groups. The organ bath was washed out three times with fresh Krebs' solution before the next substance was added and the rings were allowed to stabilize for 1 hour.

- *Statistical Analysis: All results were presented as the mean \pm SEM. The data were analyzed using SPSS program version 12. For comparison of statistical significance between

different groups, a one way ANOVA with the post hoc of Tukey's multiple comparison test was used. A value of $P \leq 0.05$ was considered statistically significant.

RESULTS

The mean value \pm SEM of fasting blood glucose of diabetic non-treated rats was 208.9 ± 1.6 mg/dl which was significantly higher when compared to the corresponding values in control rats which was 76.5 ± 1.43 mg/dl. That of fasting blood glucose of diabetic insulin-treated rats was 97.4 ± 1.47 mg/dl which was significantly lower when compared to the corresponding values in diabetic non-treated rats, and decreased to be within normal range but still significantly higher when compared to the corresponding values in control rats. In diabetic ozone-treated group, it was 149.1 ± 1.17 mg/dl which was significantly lower from the corresponding values in diabetic non-treated group, and significantly higher when compared to the corresponding values in control, diabetic insulin-treated and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 79.5 ± 1.71 mg/dl which was significantly lower when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig. 1).

The mean value \pm SEM of fasting serum cholesterol of diabetic non-treated rats was 194.7 ± 2.43 mg/dl which was significantly higher when compared to the corresponding value in control rats which was 159.6 ± 2.05 mg/dl. That of diabetic insulin-treated rats was 167.8 ± 1.88 mg/dl which was significantly lower when compared to the corresponding value in

diabetic non-treated rats, and still significantly higher when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 167.4 ± 1.79 mg/dl which was significantly lower from the corresponding value in diabetic non-treated rats, and significantly higher when compared to the corresponding value in control rats. The same value showed an insignificant change from the corresponding values in diabetic insulin-treated and diabetic ozone and insulin treated rats. In diabetic ozone- and insulin-treated group, it was 160.4 ± 1.38 mg/dl which was significantly lower when compared to the corresponding value in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig. 2).

The mean value \pm SEM of fasting serum triglycerides of diabetic non-treated rats was 100.2 ± 1.29 mg/dl which was significantly higher when compared to the corresponding value in control rats which was 71.5 ± 1.02 mg/dl. That of diabetic insulin-treated rats was 84.1 ± 0.95 mg/dl which was significantly lower when compared to the corresponding value in diabetic non-treated rats, and still significantly higher when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 84.9 ± 1.15 mg/dl which was significantly lower from the corresponding value in diabetic non-treated rats, and significantly higher when compared to the corresponding values in control and diabetic ozone- and insulin -treated rats. In diabetic ozone- and insulin-treated group, it was 74.4 ± 1.05 mg/dl which was significantly lower when compared to the corresponding values in diabetic non-

treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig.2).

The mean value \pm SEM of fasting serum HDL of diabetic non-treated rats was 28.6 ± 0.93 mg/dl which was significantly lower when compared to the corresponding value in control rats which was 44.8 ± 0.74 mg/dl. That of diabetic insulin-treated rats was 32.4 ± 0.88 mg/dl which was significantly higher when compared to the corresponding value in diabetic non-treated rats, and still significantly lower when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 32.2 ± 0.69 mg/dl which was significantly higher from the corresponding value in diabetic non-treated rats, and significantly lower when compared to the corresponding values in control and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 43.8 ± 0.75 mg/dl which was significantly higher when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig.2).

The mean value \pm SEM of fasting serum LDL of diabetic non-treated rats was 65.9 ± 1.12 mg/dl which was significantly higher when compared to the corresponding value in control rats which was 43.3 ± 0.88 mg/dl. That of diabetic insulin-treated rats was 51.3 ± 0.79 mg/dl which was significantly lower when compared to the corresponding value in diabetic non-treated rats, and still significantly higher when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 50.3 ± 0.83 mg/dl which was significantly lower from the corresponding value in diabetic non-

treated rats, and significantly higher when compared to the corresponding values in control and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 44.4 ± 0.47 mg/dl which was significantly lower when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig. 2).

The mean value \pm SEM of LDH concentration in the collected perfusion fluid of diabetic non-treated rats was 804.9 ± 2.88 U/liter which was significantly higher when compared to the corresponding value in control rats which was 342.8 ± 10.69 U/liter. That of diabetic insulin-treated rats was 603.7 ± 3.1 U/liter which was significantly lower when compared to the corresponding value in diabetic non-treated rats, but still significantly higher when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 700.5 ± 2.05 U/liter which was significantly lower from the corresponding value in diabetic non-treated group, and significantly higher when compared to the corresponding values in control, diabetic insulin-treated and diabetic ozone – and insulin-treated rats. In diabetic ozone- and insulin -treated group, it was 352.6 ± 1.53 U/liter which was significantly lower when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig. 3).

The mean value \pm SEM of catalase of diabetic non-treated rats was 0.273 ± 0.0095 U/mg tissue of heart which was significantly lower when compared to the corresponding value in control rats which

was 0.697 ± 0.0093 U/ mg tissue of heart. In diabetic insulin-treated rats, it was 0.494 ± 0.0099 U/ mg tissue of heart which was significantly higher when compared to the corresponding value in diabetic non-treated rats, and still significantly higher when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 0.58 ± 0.0073 U/ mg tissue of heart which was significantly higher from the corresponding values in diabetic non-treated and diabetic insulin-treated group and significantly lower when compared to the corresponding values in control and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 0.66 ± 0.101 U/ mg tissue of heart which was significantly higher when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Table 1).

The mean value \pm SEM of glutathione peroxidase of diabetic non-treated rats was 0.183 ± 0.0065 U/ mg tissue of heart which was significantly lower when compared to the corresponding value in control rats which was 0.316 ± 0.006 U/ mg tissue of heart. That of glutathione peroxidase of diabetic insulin-treated rats was 0.223 ± 0.0042 U/ mg tissue of heart which was significantly higher when compared to the corresponding value in diabetic non-treated rats, and still significantly lower when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 0.262 ± 0.006 U/ mg tissue of heart which was significantly higher from the corresponding values in diabetic non-treated and diabetic insulin-treated group, and significantly lower when compared to the corresponding values in control and

diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 0.302 ± 0.0055 U/ mg tissue of heart which was significantly higher when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Table 1).

The mean value \pm SEM of superoxide dismutase (SOD) of diabetic non-treated rats was 22.2 ± 0.840 U/mg tissue of heart which was significantly lower when compared to the corresponding value in control rats which was 50.2 ± 0.663 U/mg tissue of heart. That of diabetic insulin-treated rats was 31.4 ± 0.933 U/mg tissue of heart which was significantly higher when compared to the corresponding value in diabetic non-treated rats, and still significantly lower when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 38.7 ± 0.578 U/ mg tissue of heart which was significantly higher from the corresponding values in diabetic non-treated and diabetic insulin-treated group, and significantly lower when compared to the corresponding values in control and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 48.1 ± 0.737 U/mg tissue of heart which was significantly higher when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Table 1).

The mean value \pm SEM of cardiac contractility in stabilization (pre-ischemic) period of diabetic non-treated rats was 1.78 ± 0.043 g tension which was significantly lower when compared to the corresponding value in control rats which

was 3.58 ± 0.071 g tension. That of diabetic insulin-treated rats was 2.58 ± 0.045 g tension which was significantly higher when compared to the corresponding value in diabetic non-treated rats, and still significantly lower when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 2.17 ± 0.041 g tension which was significantly higher from the corresponding value in diabetic non-treated group, and significantly lower when compared to the corresponding values in control, diabetic insulin-treated and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 3.45 ± 0.040 g tension which was significantly higher when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig. 4).

The mean value \pm SEM of cardiac contractility ischemic period of diabetic non-treated rats was 0.54 ± 0.034 g tension which was significantly lower when compared to the corresponding value in control rats which was 1.74 ± 0.037 g tension. That of cardiac contractility in ischemic period of diabetic insulin-treated rats was 1.23 ± 0.06 g tension which was significantly higher when compared to the corresponding value in diabetic non-treated rats, and still significantly lower when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 0.91 ± 0.031 g tension which was significantly higher from the corresponding value in diabetic non-treated group, and significantly lower when compared to the corresponding values in control, diabetic insulin-treated and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated

group, it was 1.59 ± 0.031 g tension which was significantly higher when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig. 4).

The mean value \pm SEM of cardiac contractility in reperfusion (post-ischemic) period of diabetic non-treated rats was 0.99 ± 0.066 g tension which was significantly lower when compared to the corresponding value in control rats which was 2.48 ± 0.059 g tension. That of diabetic insulin-treated rats was 1.87 ± 0.076 gm. tension which was significantly higher when compared to the corresponding value in diabetic non-treated rats, and still significantly lower when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 1.38 ± 0.052 g tension which was significantly higher from the corresponding value in diabetic non-treated group, and significantly lower when compared to the corresponding values in control, diabetic insulin-treated and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 2.27 ± 0.059 g tension which was significantly higher when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig. 4).

The mean value \pm SEM of heart rate (beat / minute) in stabilization (pre-ischemic) period of diabetic non-treated rats was 134.7 ± 1.32 beat / minute which was significantly lower when compared to the corresponding value in control rats which was 205.6 ± 1.29 beat / minute. That of diabetic insulin-treated rats was 184.9 ± 1.32 beat / minute which was significantly higher when compared to the

corresponding value in diabetic non-treated rats, and still significantly lower when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 165 ± 1.15 beat / minute which was significantly higher from the corresponding value in diabetic non-treated group, and significantly lower when compared to the corresponding values in control, diabetic insulin-treated and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group it was 203.1 ± 1.14 beat / minute which was significantly higher when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats. The same value showed an insignificant change from the corresponding value in control rats (Fig. 5).

The mean value \pm SEM of heart rate (beat / minute) in ischemic period of diabetic non-treated rats was 7 ± 0.63 beat / minute which was significantly lower when compared to the corresponding value in control rats which was 43.4 ± 0.72 beat / minute. That of diabetic insulin-treated rats was 26.7 ± 0.73 beat / minute which was significantly higher when compared to the corresponding value in diabetic non-treated rats, and still significantly lower when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 15.9 ± 0.48 beat / minute which was significantly higher from the corresponding value in diabetic non-treated group, and significantly lower when compared to the corresponding values in control, diabetic insulin-treated, and diabetic ozone- and insulin-treated rats. In diabetic ozone and insulin -treated group, it was 41.1 ± 0.67 beat / minute

which was significantly higher when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig. 5).

The mean value \pm SEM of heart rate (beat / minute) in reperfusion (post-ischemic) period of diabetic non-treated rats was 336 ± 7.29 beat / minute which was significantly higher when compared to the corresponding value in control rats which was 234 ± 5.76 beat / minute. That of diabetic insulin-treated rats was 269.5 ± 2.98 beat / minute which was significantly lower when compared to the corresponding value in diabetic non-treated rats, and still significantly higher when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 291 ± 3.02 beat / minute which was significantly lower from the corresponding value in diabetic non-treated group, and significantly higher when compared to the corresponding values in control, diabetic insulin-treated and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 249.5 ± 3.36 beat / minute which was significantly lower when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig. 5).

The mean value \pm SEM of vascular reactivity of aortic strip to 1×10^{-5} M nor-epinephrine of diabetic non-treated rats was 100.4 ± 1.02 mg tension which was significantly higher when compared to the corresponding value in control rats which was 45.9 ± 1.24 mg tension. That of diabetic insulin-treated rats was 62.4 ± 0.92 mg tension which was significantly

lower when compared to the corresponding value in diabetic non-treated rats, but still significantly higher when compared to the corresponding values in control rats. In diabetic ozone-treated group, it was 73.5 ± 1.07 mg tension which was significantly lower from the corresponding value in diabetic non-treated group, and significantly higher when compared to the corresponding values in control, diabetic insulin-treated and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 48.9 ± 1.16 mg tension which was significantly lower when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats. The same value was insignificant with the corresponding values in control rats (Fig. 6).

The mean value \pm SEM of vascular reactivity of aortic strip to 1×10^{-6} M vasopressin of diabetic non-treated rats was 102.4 ± 1.22 mg tension which was significantly higher when compared to the corresponding values in control rats which was 46.1 ± 0.93 mg tension. That of diabetic insulin-treated rats was 64.4 ± 1.24 mg tension which was significantly lower when compared to the corresponding value in diabetic non-treated rats, but still significantly higher when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 75.3 ± 1.03 mg tension which was significantly lower from the corresponding value in diabetic non-treated group, and significantly higher when compared to the corresponding

values in control, diabetic insulin-treated and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 49.7 ± 1.2 mg tension which was significantly lower when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats (Fig. 6).

The mean value \pm SEM of vascular reactivity of aortic strip to 1×10^{-6} M indomethacin of diabetic non-treated rats was 101.9 ± 1.16 mg tension which was significantly higher when compared to the corresponding value in control rats which was 45.7 ± 0.96 mg tension. That of diabetic insulin-treated rats was 63.9 ± 0.98 mg tension which was significantly lower when compared to the corresponding value in diabetic non-treated rats, but still significantly higher when compared to the corresponding value in control rats. In diabetic ozone-treated group, it was 74.5 ± 1.09 mg tension which was significantly lower from the corresponding value in diabetic non-treated group, and significantly higher when compared to the corresponding values in control, diabetic insulin-treated and diabetic ozone- and insulin-treated rats. In diabetic ozone- and insulin-treated group, it was 50.1 ± 1.31 mg tension which was significantly lower when compared to the corresponding values in diabetic non-treated, diabetic insulin-treated and diabetic ozone-treated rats. The same value was insignificant with the corresponding value in control rats (Fig. 6).

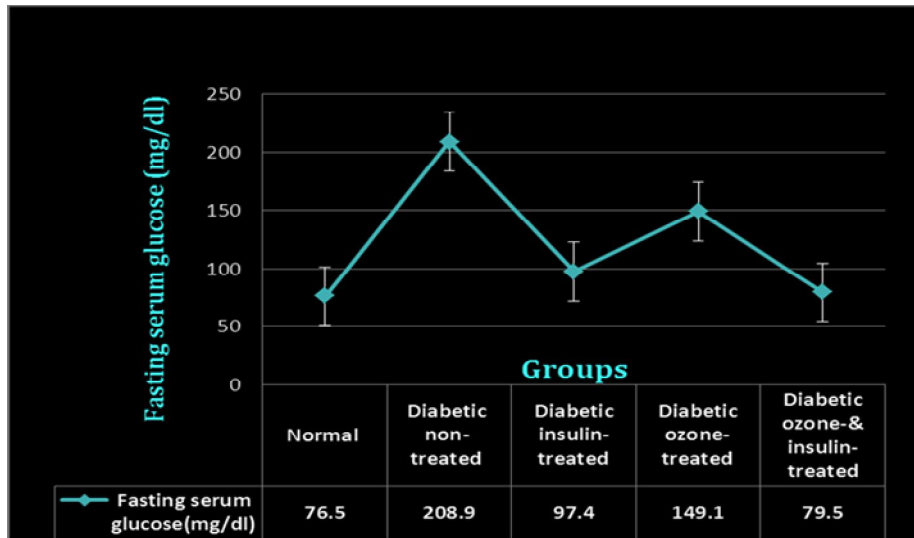


Figure (1): Fasting serum glucose (Mean ± S.E.)

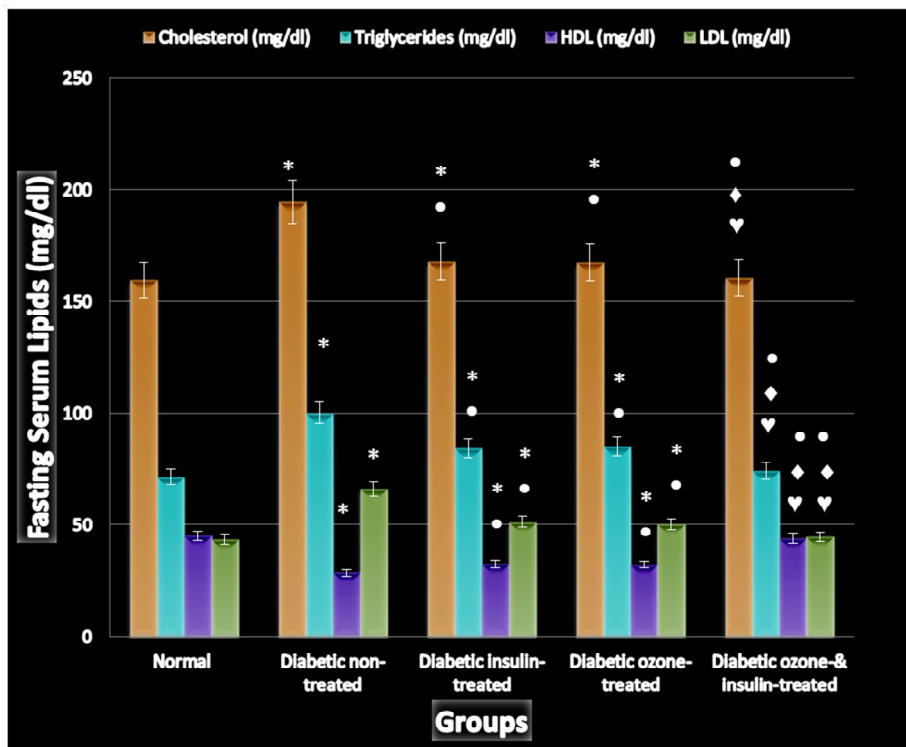


Figure (2): Fasting serum cholesterol (mg/dl), triglycerides (mg/dl) LDL (mg/dl) and HDL (mg/dl) in normal, diabetic non-treated, diabetic insulin-treated, diabetic ozone-treated and diabetic ozone-& insulin-treated groups (Mean ± S.E.).

- *Significant when compared to the corresponding values in normal group.
- Significant when compared to the corresponding values in diabetic non-treated group.
- ♦Significant when compared to the corresponding values in diabetic insulin-treated group.
- ♥ Significant when compared to the corresponding values in diabetic ozone-treated group.

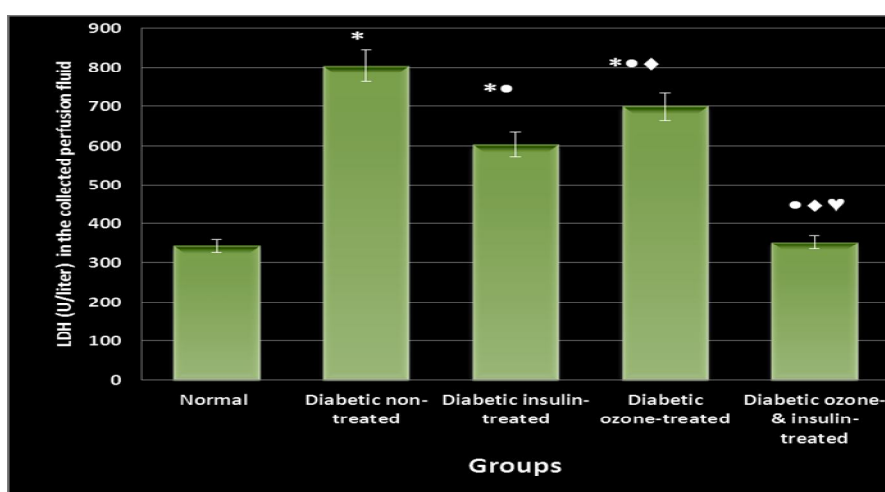


Figure (3): The mean values \pm S.E. of LDH (U/liter in the collected perfusion fluid) in control, diabetic non-treated, diabetic insulin-treated, diabetic ozone-treated and diabetic ozone- and insulin-treated rats.

*Significant when compared to the corresponding values in normal group.

• Significant when compared to the corresponding values in diabetic non-treated group.

♦ Significant when compared to the corresponding values in diabetic insulin-treated group.

♥ Significant when compared to the corresponding values in diabetic ozone-treated group.

Table (1): Mean \pm S.E. of catalase, glutathione peroxidase and SOD (U / mg of heart tissue) level in control, diabetic non-treated, diabetic insulin-treated, diabetic ozone- and insulin-treated rats.

Groups		Normal (N)	Diabetic non-treated (DNT)	Diabetic insulin-treated (DIT)	Diabetic ozone-treated (DOT)	Diabetic ozone- and insulin- treated (DOIT)
Catalase (U/mg)		0.697 \pm 0.009	0.273 \pm 0.0095	0.494 \pm 0.0099	0.58 \pm 0.0073	0.66 \pm 0.101
P values	N		<0.01	<0.01	<0.01	>0.05
	DNT	<0.01		<0.01	<0.01	<0.01
	DIT	<0.01	<0.01		<0.01	<0.01
	DOT	<0.01	<0.01	<0.01		<0.01
	DOIT	>0.05	<0.01	<0.01	<0.01	
Glutathione peroxidase (U/mg)		0.316 \pm 0.006	0.183 \pm 0.0065	0.223 \pm 0.0042	0.262 \pm 0.006	0.302 \pm 0.0055
P values	N		<0.01	<0.01	<0.01	>0.05
	DNT	<0.01		<0.01	<0.01	<0.01
	DIT	<0.01	<0.01		<0.01	<0.01
	DOT	<0.01	<0.01	<0.01		<0.01
	DOIT	>0.05	<0.01	<0.01	<0.01	
Superoxide dismutase activity (U/mg)		50.2 \pm 0.663	22.2 \pm 0.840	31.4 \pm 0.933	38.7 \pm 0.578	48.1 \pm 0.737
P values	N		<0.01	<0.01	<0.01	>0.05
	DNT	<0.01		<0.01	<0.01	<0.01
	DIT	<0.01	<0.01		<0.01	<0.01
	DOT	<0.01	<0.01	<0.01		<0.01
	DOIT	>0.05	<0.01	<0.01	<0.01	

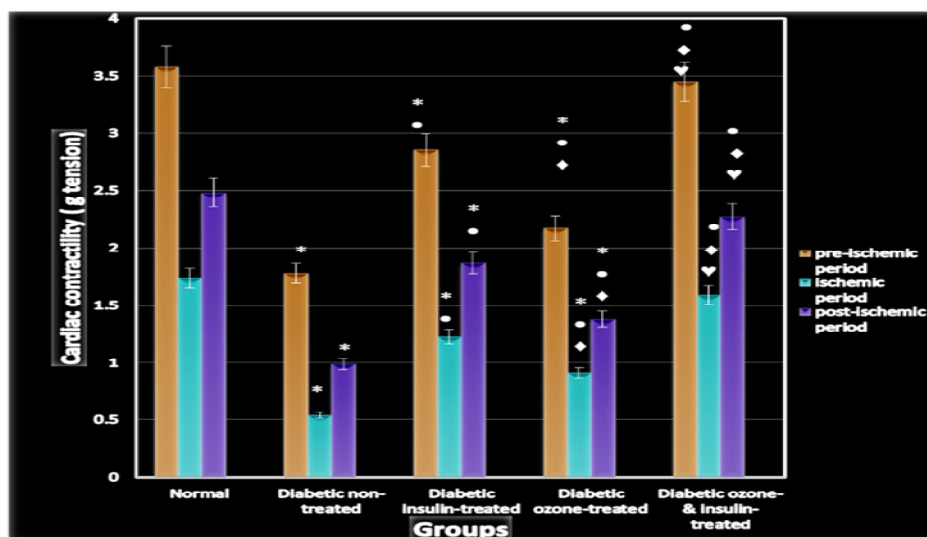


Figure (4): The Mean values \pm S.E. of cardiac contractility (g tension) in stabilization (pre-ischemic), ischemic and reperfusion (post-ischemic) periods in normal, diabetic non-treated, diabetic insulin-treated, diabetic ozone-treated and diabetic ozone- and insulin-treated groups.

- * Significant when compared to the corresponding values in normal group (p < 0.05)
- Significant when compared to the corresponding values in diabetic non treated group (p < 0.05)
- ◆ Significant when compared to the corresponding values in diabetic insulin treated group (p < 0.05)
- ▼ Significant when compared to the corresponding values in diabetic ozone treated group (p < 0.05)

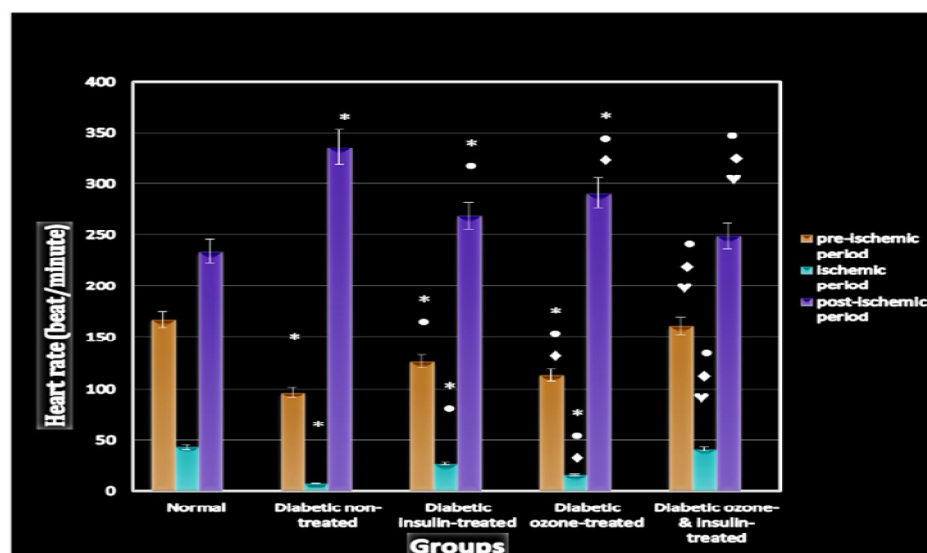


Figure (5): The Mean values \pm S.E. of heart rate (beat/minute) in stabilization (pre-ischemic), ischemic and reperfusion (post-ischemic) periods in normal, diabetic non-treated, diabetic insulin-treated, diabetic ozone-treated and diabetic ozone- and insulin-treated groups.

- * Significant when compared to the corresponding values in normal group (p < 0.05)
- Significant when compared to the corresponding values in diabetic non treated group (p < 0.05)
- ◆ Significant when compared to the corresponding values in diabetic insulin treated group (p < 0.05)
- ▼ Significant when compared to the corresponding values in diabetic ozone treated group (p < 0.05)

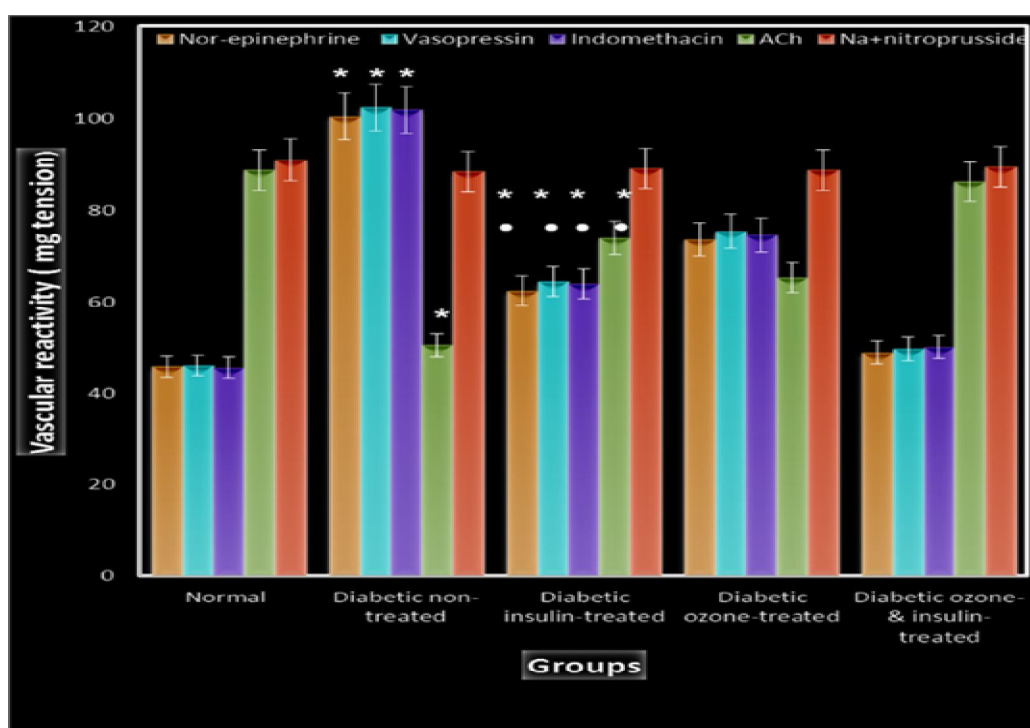


Figure (6): Mean \pm S.E. of vascular reactivity of aortic strip (mg tension) to nor-epinephrine, vasopressin, indomethacin, ACh and Na⁺ nitroprusside in normal, diabetic non-treated, diabetic insulin-treated, diabetic ozone-treated and diabetic ozone- & insulin-treated groups.

DISCUSSION

Clinical studies suggested that diabetic patients have a significantly greater incidence and severity of several cardiopathies (e.g. angina, acute myocardial failure, and atherosclerosis) and approximately 80% of all patients with diabetes die of cardiovascular diseases (Feuvray and Lopaschuk, 1997). Ischemia–reperfusion injury may occur as damage to the myocardium following blood restoration after a critical period of coronary occlusion (Bolli and Marban, 1999). Oxidative stress explains the pathogenesis of ischemia–reperfusion injury (Griendling and Alexander, 1997). Oxidative stress is usually

associated with increased formation of reactive oxygen species (ROS), modifies phospholipids and proteins leading to lipid peroxidation and oxidation of thiol groups. These changes are considered to alter membrane permeability and configuration in addition to producing functional modification of various cellular proteins (Suzuki et al., 1997).

Because ozone therapy can activate the antioxidant system, and improve some markers of endothelial cell damage (Martínez-Sánchez et al., 2005), medical ozone treatment could be used as a complementary therapy in the treatment of diabetes and its complications.

Marked fluctuations in glucose levels contribute to more oxidative stress in the diabetic condition, even in patients being treated with insulin (**Gao et al., 2012**), and with time results in the development of diabetic complications (**Ceriello and Testa, 2009**). In the present investigation, administration of ozone (1mg/kg B.W.) for 15 days revealed a significant decrease of fasting serum glucose when compared to the corresponding values in diabetic-non treated rats. This indicated that ozone therapy potentially improved the glycemic control during diabetes. **Eman et al. (2013)** recorded a significant increase in β -cell number in diabetic ozone-treated group when compared with diabetic non-treated group. **Gergorio et al. (2005)** observed a significant decrease in the percentage of damaged islets for diabetic rats treated with ozone with regard to STZ group.

In the present study, STZ injected animals exhibited a significantly higher fasting total cholesterol, triglycerides and LDL levels and a significant decrease in fasting serum HDL level when compared to the normal group. These results indicated a significant dyslipidemia in untreated diabetic rats. **Sout (2005)** considered diabetic dyslipidemia and hyperglycemia to be predictors of cardiovascular complications. The elevation of lipid profiles in STZ-diabetic rats may be attributed to an increase in the rate of lipolysis with a decrease in lipogenesis leading to release more fatty acids into the blood circulation (**Agardh et al., 1999**). Elevation of serum lipids indicates either the defective removal or overproduction (or both) of one or more lipoproteins (**Akula et al., 2003**). Diabetic insulin-treated rats exhibited significantly lower

fasting total cholesterol, triglycerides and LDL levels, and a significant increase in fasting serum HDL level when compared to the diabetic non-treated group reflecting a partial improvement of dyslipidemia as still there is significant difference when compared to normal rats. As insulin has a profound role in the regulation of key enzymes involved in the lipid and lipoprotein metabolism, its deficiency causes major changes in the activity of these enzymes and thereby affecting overall lipid metabolism and lipid profile of various tissues (**Mironava et al., 2000**). The altered lipid and lipoprotein pattern observed in diabetic rats could be due to defect in insulin secretion and/or action (**Krishnaswami, 1996**). Diabetic ozone-treated rats exhibited significantly lower fasting total cholesterol, triglycerides and LDL levels and a significant increase in fasting serum HDL level. **Udupa et al. (2012)** reported that antioxidants showed an improvement in insulin sensitivity in patients with type 2 diabetes mellitus. So, the improvement of lipid profile with ozone treatment in the present study may be attributed to a relative improvement of insulin level and decreased insulin resistance as a result of controlling the redox state. **Haobo et al. (2013)** stated that the release of plasma LDH significantly increased in diabetic rats. This indicates that myocardial cellular injury is more severe in diabetic than that in the control rats during reperfusion. Hyperglycemia enhances oxidative stress, and reduces antioxidant defenses (**Kain et al., 2011**). Compared to the diabetic non-treated group, insulin-treated group exhibited a significantly lower LDH level in the collected perfusion fluid following IR reflecting a

cardio-protective role for insulin. In the rat, insulin reduces the infarct size, plasma creatine kinase and lactate dehydrogenase (both markers of myocardial injury), and apoptosis following IR (Xing et al., 2009).

Ozone-treated group exhibited a significantly lower LDH level in the collected perfusion fluid following IR reflecting a cardio-protective role for ozone treatment on cardiac cell damage after IR. These results were in agreement with Lamiaa et al. (2012) who concluded that ozone therapy can afford significant cardio-protection against biochemical and histological changes associated with IR injury by oxidative preconditioning as it appeared in reducing creatine kinase-MB release, oxidative stress, lactate accumulation, as well as preserving myocardial adenine nucleotides. Histological examination also revealed better improvement with ozone therapy compared to the non-treated I/R group. Filippo et al. (2015) found that ozone treatment led to an evident increase of both systolic and diastolic functions and then of the myocardial performance index with decrease of LDH levels confirming the important role of ozone in the cardio-protection already seen in the present study.

Oxidative stress is the major mechanism that triggers IR injury. Ozone maintains cellular antioxidant systems including glutathione, SOD, and enzymatic reactions, preparing the host to confront the pathophysiologic conditions mediated by oxidative stress (Kesik et al., 2009). The hyperglycemia-induced increase of IRI in diabetes can be prevented by treatment with antioxidants (Akhtar et

al., 2012). Several studies revealed that the addition of antioxidants or scavengers, such as SOD and catalase, could reduce infarct size (Chi et al., 1989 and Kilgore et al., 1994). Ozone oxidative preconditioning appears to restore the oxidant balance, minimizing tissue injury caused by IR injury (Bhalla et al., 1999).

Regarding antioxidant enzymes in the present investigation, catalase, glutathione peroxidase and SOD levels in heart tissue partially reversed in insulin-treated diabetic rats when compared to diabetic non-treated rats to a level that was significantly different from control rats, reflecting partial improvement in oxidative stress state with preservation of antioxidant enzymes. Ryuichi et al. (2000) stated that insulin protects cardiomyocytes from oxidative stress-induced apoptosis. Seiichi et al. (2004) indicated that GSH and its related enzyme activities are impaired in diabetic endothelial cells; and these impairments are prevented by treatment with insulin. Insulin can prevent oxidative damage by reducing the formation of peroxynitrite (ONOO) free radicals after myocardial ischemia and reperfusion in the rats (Koo and Vaziri, 2003). In diabetic rat hearts, insulin restores the activity of glutathione peroxidase, a key antioxidant enzyme, correspondingly reduces the toxic process of lipid peroxidation and increases eNOS expression (Zobali et al., 2002).

Regarding antioxidant enzymes in the present investigation, catalase, glutathione peroxidase and SOD levels in heart tissue were partially reversed in ozone-treated diabetic rats. Gregorio et al. (2005) found that blood oxidative stress was controlled by ozone as shown in increased antioxi-

dant endogenous systems (superoxide dismutase, catalase, glutathione peroxidase and reduced glutathione). Ozone helps to alleviate oxidative stress associated with diabetes mellitus (**Bocci, 2006**). The capacity of ozone to enhance antioxidant endogenous systems, in front of oxidative stress by oxidative preconditioning or adaptive mechanisms has been demonstrated (**Le?n et al., 1998**). Therefore, these results suggest that ozone protective effects on antioxidant endogenous defenses improve glucose metabolism.

In the present study, STZ injected animals exhibited a significantly lower cardiac contractility and heart rate in stabilization (preischemic) period which reflected a significant cardiomyopathy. These results were in agreement with **Giulianna et al. (2006)**, who found that basal cardiac contractility and heart rate decrease in STZ-diabetic rats. Reduction of heart rate is an early event in type 1 diabetes mellitus in a number of species including rats, rabbits, and humans (**Wali et al., 2013**). Diabetic hearts are chronically subjected to hyperglycemia and hyperlipidemia, both thought to contribute to oxidizing conditions and contractile dysfunction (**Niraj et al., 2014**). Myocardial triglycerides and cholesterol content significantly increase in diabetic rats (**Sharma et al., 2004**). Diabetic insulin treated rats exhibited a significantly higher cardiac contractility and heart rate in stabilization (pre-ischemic) period when compared to diabetic non-treated rats reflecting a partial reversal of diabetic cardiomyopathy with insulin treatment and still there is significant difference when compared to normal rats. **Giulianna et al. (2006)** found that treatment with insulin prevented the occurrence of

alterations caused by diabetes, i.e. bradycardia, hypotension and attenuated basal inotropism. The decrease in cardiac contractility induced by chronic diabetes results in part from decrease in expression and alteration in function of ryanodine type 2 calcium release channel in cardiac myocyte, and these changes can be reversed by insulin treatment (**Bidasee et al., 2013**). Diabetic ozone-treated rats exhibited a significantly higher cardiac contractility and heart rate in stabilization (preischemic) period when compared to diabetic non-treated rats reflecting a partial reversal of diabetic cardiomyopathy with ozone treatment and still there is significant difference when compared to normal rats.

In ischemic period, in the present study, STZ injected animals exhibited a significantly lower cardiac contractility and heart rate when compared to the normal group, reflecting severe decrease in cardiac performance. **Jiung-Pang et al. (2009)** found that acute myocardial ischemic injury in diabetic rats markedly reduced cardiac output subsequent to bradycardia and reduction of contractility. **Ryuko et al. (2007)** reported that the hearts of spontaneously diabetic rats were found to be more susceptible to ischemic insult. Cardiac function is critically dependent on substrate utilization, and changes in myocardial fuel selection can have a major impact both positively and negatively (**Lopaschuk, 2001**). In myocardial ischemic injury, the blood flow in the heart is reduced, decreasing the substrates which are essential to myocardium workload. This pathological condition favors an imbalance between ROS production and the protective antioxidant defense system, thereby

increasing ROS mediated oxidative stress (Hill and Singal, 1996). Moreover, increased oxidative stress plays a critical role in diabetes complications as demonstrated by increased levels of oxidized DNA, proteins and lipids in diabetic subjects (Wiernsperger, 2003). Diabetic insulin-treated rats exhibited a significantly higher cardiac contractility and heart rate when compared to the diabetic non-treated group, reflecting improvement in cardiac performance which still significantly lower when compared to normal rats. The presence of hyperglycemia during myocardial ischemia is closely associated with insulin resistance, which in turn attenuates cardiac sensitivity to exogenous insulin administration (Moriscoet al., 2007). In the present study, *diabetic ozone-treated rats* exhibited a significantly higher cardiac contractility (mg tension) and heart rate (beat /minute), when compared to the diabetic non-treated group, in ischemic period, reflecting improvement in cardiac performance which still significantly lower when compared to normal rats. Ozone therapy leads to the activation of glycolysis with an increase in ATP and 2,3- diphosphoglycerate and increases the release of oxygen in the ischemic tissues (Bocci et al., 2009). So, ozone is used in complementary treatment of hypoxic and ischemic syndromes (Clavo et al., 2003).

In reperfusion (post-ischemic) period, in the present study, STZ injected animals exhibited a significantly lower cardiac contractility and higher heart rate when compared to the normal group, in post-ischemic (reperfusion) period, reflecting development of severe contractile dysfunction and ventricular tachyarrhy-

thmia. Reperfusion of myocardium subjected to a transient ischemia rapidly induces ventricular arrhythmias including VT and VF in both animals and human (Lu et al., 1999). Reperfusion arrhythmias and transient mechanical dysfunction are components of myocardial ischemia-reperfusion injuries (Buja and Weerasinghe, 2010). Oxidative stress caused by reactive oxygen species has a considerable role in ischemia/reperfusion injury, which impairs cardiac function (Inafuku et al., 2013). Diabetic insulin treated rats exhibited a significantly higher cardiac contractility and lower heart rate in reperfusion (post-ischemic) period when compared to diabetic non-treated rats reflecting a partial improvement in recovery of cardiac contractility and decrease in rate of tachyarrhythmia with insulin treatment as still there is significant difference when compared to normal rats. The impairment in functional recovery of diabetic mice after ischemia/reperfusion could be ameliorated by insulin and glucose in perfusates which increased glucose use and enhanced cardiac efficiency (Dragoy et al., 2007).

Diabetic ozone-treated rats exhibited a significantly higher cardiac contractility and lower heart rate in reperfusion (post-ischemic) period when compared to diabetic non-treated rats reflecting a partial improvement in recovery of cardiac contractility and decrease in rate of tachyarrhythmia with ozone treatment as still there is significant difference when compared to normal rats. Ofer et al. (2007) reported a significantly better post-ischemic hemodynamic recovery in ozone-treated rats.

Administration of streptozotocin to rats caused higher vascular reactivity of aortic strips to norepinephrine, vasopressin and indomethacin, significantly lower percentage of relaxation to acetylcholine (endothelium-dependent relaxation), and insignificant changes to Na⁺ nitroprusside (endothelium-independent relaxation) in aortic strips when compared to the corresponding values in normal rats reflecting endothelial dysfunction and preserved vascular smooth muscles. **Naowaboot et al. (2009)** reported that vascular responses of the diabetic rats to acetylcholine significantly suppressed, whereas those to phenylephrine significantly increased as compared to normal rats. **Wang et al. (2009)** concluded that maximum contraction to NA increases significantly in diabetic aorta, and significant decrease in relaxation to ACh in diabetic group compared with controls. **Keenoy et al. (2005)** also reported a decline of total antioxidant status in diabetes mellitus. It has been shown that vessels from diabetic animals exhibited abnormal endothelium dependent vascular relaxation to acetylcholine. This endothelium-dependent vasodilatation is reduced in diabetes largely due to excessive oxidative stress and decreased bioavailability of nitric oxide (**Majithiya et al., 2005**). Administration of insulin caused significant decrease in vascular reactivity of aortic strips to norepinephrine, vasopressin and indomethacin and significant increase in percent of relaxation to acetylcholine (endothelium-dependent relaxation) and insignificant changes to Na⁺nitroprusside (endothelium-independent relaxation) in aortic strips when compared to the corresponding values in diabetic-non

treated rats. **Lembo et al. (1995)** showed that insulin has the ability to modulate the vascular contractile response evoked by various vasoactive substances. The vascular actions of insulin are mediated chiefly through the regulation of endothelium-derived factors. In this regard, insulin can stimulate the production of nitric oxide (NO) (**Potenza et al., 2009**). Administration of ozone caused significant decrease in vascular reactivity of aortic strips to norepinephrine, vasopressin and indomethacin and significant increase in percent of relaxation to acetylcholine (endothelium-dependent relaxation) and insignificant changes to Na⁺nitroprusside (endothelium-independent relaxation) in aortic strips when compared to the corresponding values in diabetic-non treated rats. **Clavo et al. (2004)** showed that ozone therapy could decrease the vasoconstriction. These results were also supported by **Al-Dalain et al. (2001)**, who showed that with ozone treatment there was improvement in aortic relaxation and decreased micro-vessel reactivity, in STZ-experimental model of diabetes. Our results indicated that the combination of insulin treatment with ozone revealed more significant enhancement in oxidative state and improvement in cardiac functions in STZ-induced diabetic rats. According to **Sindhu et al. (2004)**, combined therapy with insulin and antioxidants normalized all measured antioxidant enzyme protein expression and activities. Thus, diabetes-associated reductions in antioxidant enzymes can be ameliorated by combined insulin and antioxidant therapy.

The ozone treatment, by means of its oxidative preconditioning effect, normalizes glucose levels and consequently

restores the concentrations of organic peroxides. So, it contributes to the control of the vascular complications of diabetes.

REFERENCES

1. **Agardh CD., Bjorgell P. and Nilson EP. (1999):** The effect of tolbutamide on lipoproteins and lipoprotein lipase and hormone sensitive lipase. *Diab Res Clin Pract.*, :4699-708.
2. **Akhtar S, Yousif M, Chandrasekhar B and Benter IF (2012):** Activation of EGFR/ERBB2 via Pathways Involving ERK1/2, P38 MAPK, AKT and FOXO Enhances Recovery of Diabetic Hearts from Ischemia-Reperfusion Injury. *PLoS ONE*, 7 (6): e39066.
3. **Akula A., Kota M.K. and Gopisetty S.G. (2003):** Biochemical, histological and echocardiographic changes during experimental cardiomyopathy in STZ-induced diabetic rats. *Pharmacological Research*, (48):429-435.
4. **Al-Dalain M S., Martinez G., Alil E. C.J., Menendez S., Re L., Giuliani A. and Leon O.S (2001):** Ozone treatment reduces markers of oxidative and endothelial damage in an experimental diabetes model in rats. *Pharmacological Research*, 44 (5): 391–396.
5. **Baynes JW. (1991):** Role of oxidative stress in development of complications in diabetes. *Diabetes*,40: 405–412.
6. **Beatch G. and McNeill J. (1988):** Ventricular arrhythmias following coronary artery occlusion in the streptozotocin diabetic rat. *Can. J. Physiol. Pharmacol.*, 66: 312 – 317.
7. **Beauchamp, C. and Fridovich, I. (1971):** Superoxide dismutase: improved assays and an assay applicable to acrylamide gels. *Anal. Biochem.*, 44: 276–287.
8. **Bhalla S. Gupta K., and Reinhart P. (1999):** Alteration of epithelial integrity, alkaline phosphatase activity, and fibronectin expression in lungs of rats exposed to ozone. *Journal of Toxicology and Environmental HealthA*, 57(5): 329–343.
9. **Bhimji S., Godin D. and McNeill J. (1986):** Coronary artery ligation and reperfusion in alloxan-diabetic rabbits: ultrastructural and haemodynamic changes. *Br. J. Exp. Pathol.*, 67: 851 – 863.
10. **Bidasee KR, Nallani K, Henry B, Dincer UD and Besch HR (2013):** Chronic diabetes alters function and expression of ryanodine receptor calcium-release channels in rat hearts. *Mol Cell Biochem.*, 249 (1-2):113-23.
11. **Bocci VA (2006):** Scientific and medical aspects of ozone therapy. *State of the art. ArchMed Res.*, 37:425–435.
12. **Bocci V, Borrelli E, Travagli V and Zanardi I (2009):** The ozone paradox: Ozone is a strong oxidant as well as a medical drug. *Medicinal Res Rev.*, 29:646-682.
13. **Bolli R and Marban E. (1999):** Molecular and cellular mechanisms of myocardial stunning. *Physiol Rev.*, 79: 609–634.
14. **Buja L. M. and Weerasinghe P. (2010):** Unresolved issues in myocardial reperfusion injury. *Cardiovascular Pathology*, 19 (1) : 29–35.
15. **Burstein M (1970):** Rapid method for isolation of lipoproteins from human serum by precipitation with polyanions. *J Lipid Res.*, 11: 585- 590.
16. **Ceriello A. and Testa R. (2009):** Antioxidant anti- Inflammatory treatment in type 2 diabetes. *Diabetes Care*, 32 (2): S232 –S236.
17. **Chi L.G., Tamura Y., Hoff P.T., Macha M., Gallagher K.P., Schork M.A. and Lucchesi B.R. (1989):** Effect of superoxide dismutase on myocardial infarct size in the canine heart after 6 hours of regional ischemia and reperfusion: a demonstration on myocardial salvage. *Circulation Research*, 64 (4):665–675.
18. **Clavo B., Catal? L., Pérez J. , Rodr?guez V and RobaiL. (2004):** Ozone Therapy on Cerebral Blood Flow: A Preliminary Report *eCAM*, 1(3):315-319.
19. **Dragoy Hafstad A, Khalid AM, How OJ, Larsen TS and Aasum E. (2007):** Glucose and insulin improve cardiac efficiency and post-ischemic functional recovery in perfused hearts from type 2 diabetic (db/db) mice. *Am J Physiol Endocrinol Metab.*, 292:E1288–E1294.

20. **Eman G.H.E., Samia M. and Anwaar A. (2013):** Comparison between the effect of ozone and vitamin C in treatment of diabetes mellitus. *The Egyptian Journal of Hospital Medicine*, 51:434–447.
21. **Feuvray D and Lopaschuk GD (1997):** Controversies on the sensitivity of diabetic heart to ischemic injury: The sensitivity of the diabetic heart to ischemic injury is decreased. *Cardiovasc Res.*, 34: 113–120.
22. **Filippo T., Maisto S., Luongo M., Alfano A., Rossi F. and Amico L. (2015):** Daily Oxygen/O₃ Treatment Reduces Muscular Fatigue and Improves Cardiac Performance in Rats Subjected to Prolonged High Intensity Physical Exercise. *Oxidative Medicine and Cellular Longevity*, 2015: 190640.
23. **Friedwald WT., Leve RI. and Fredrickson DS. (1972):** Estimation of the concentration of the LDL-C without the use of preparative ultracentrifuge. *Clin Chem.*, 18: 499-501.
24. **Giulianna B., Mauro O., Helio S. and Rubens F. (2006):** Myocardial performance in conscious streptozotocin diabetic rats. *Cardiovascular Diabetology*, 5:1-26.
25. **Gomes M., Domenech E. and Vi? a J. (2008):** Moderate exercise is an antioxidant: upregulation of antioxidant genes by training. *Free Radical Biology and Medicine*, 44, (2): 126–131.
26. **Gregorio M., Said M., Silvia M., Attilia G. and Olga S. (2005):** Ozone Treatment Reduces Blood Oxidative Stress and Pancreas Damage in a Streptozotocin-Induced Diabetes Model in Rats. *Acta Farm. Bonaerense*, 24(4): 491-7.
27. **Griendling KK and Alexander RW (1997):** Oxidative stress and cardiovascular disease. *Circulation*, 96:3264–3265.
28. **Hafeman D.G., Sunde R.A. and Hoekstra W.G. (1974):** Effect of dietary selenium on erythrocyte and liver glutathione peroxidase in the rat. *Nutrition*, 104: 580–587.
29. **Hill M. and Singal P. (1996):** Antioxidant and oxidative stress changes during heart failure subsequent to myocardial infarction in rats. *Am J Pathol.*, 148:291–300.
30. **Inafuku H., Kuniyoshi Y., Yamashiro S., Katsuya A., Takaaki N., Yuji M. and Yuya K. (2013):** Determination of oxidative stress and cardiac dysfunction after ischemia reperfusion injury in isolated rat hearts. *Annals of Thoracic and Cardiovascular Surgery*, 19(3): 186–194.
31. **Kain V., Kumar S. and Sitasawad SL. (2011):** Azelnidipine prevents cardiac dysfunction in streptozotocin-diabetic rats by reducing intracellular calcium accumulation, oxidative stress and apoptosis. *Cardiovasc Diabetol.*, 10:97-110
32. **Keenoy B., Campenhout A., Aerts P., Vertommen J., Abrams P., Gaal L., Van G. and Leeuw I. (2005):** Time Course of Oxidative Stress Status in the Postprandial and Post-absorptive States in Type 1 Diabetes Mellitus: Relationship to Glucose and Lipid Changes. *Journal of the American College of Nutrition*, 24 (6): 474-485.
33. **Kesik B., Uysal B., Kurt E., Kismet and Koseoglu V. (2009):** Ozone ameliorates methotrexate-induced intestinal injury in rats. *Cancer Biology and Therapy*, 8(17): 1623–1628.
34. **Kilgore S., Friedrichs G.S., Johnson C.R. Schasteen C.S., Riley D.P., Weiss R.H., Ryan U. and Lucchesi B.R. (1994):** Protective effects of the SOD-mimetic SC-52608 against ischemia/reperfusion damage in the rabbit isolated heart. *Journal of Molecular and Cellular Cardiology*, 26(8): 995–1006.
35. **Koo JR and Vaziri ND. (2003)** Effects of diabetes, insulin and antioxidants on NO synthase abundance and NO interaction with reactive oxygen species. *Kidney Int.*, 63:195–201.
36. **Krishnaswami S. (1996):** The relevance of lipids in Indians. *Lipid India*. 1:5–7.
37. **Kusama Y., Hearse D. and Avkiran M. (1992):** Susceptibility to reperfusion-induced ventricular arrhythmias in diabetes. *J. Mol. Cell. Cardiol.*, 24: 411 – 421.
38. **Lamiaa A., Hesham A., Mohamed N., Amina S. and Azza M. (2012):** Cardio-protective effects of ozone oxidative preconditioning in an in vivo model of

- ischemia/reperfusion injury in rats. *Scandinavian Journal of Clinical and Laboratory Investigation*, 72(5): 345-354.
39. **Lembo G., Iaccarino G., Vecchione C., Rendina V. and Trimarco B. (1995):** Insulin Modulation of Vascular Reactivity Is Already Impaired in Prehypertensive Spontaneously Hypertensive Rats. *Hypertension*, 26:290-293.
 40. **Le?n OS, Menéndez S, Merino N, Castillo R, Sam S, Pérez L, Cruz E and Bocci V. (1998):** Ozone oxidative preconditioning: a protection against cellular damage by free radicals. *Mediators Inflamm*, 7: 289-294.
 41. **Lopaschuk GD. (2001):** Optimizing cardiac energy metabolism how can fatty acid and carbohydrate metabolism be manipulated? *Coron Artery Dis.*, 12 (Suppl 1): S8–11.
 42. **Lu R., Yang P., Remeysen P., Saels A., Dai D., and Clerck, DF. (1999):** Ischemia reperfusion induced arrhythmias in anaesthetized rats: a role of Na⁺ and Ca²⁺ influx. *European Journal of Pharmacology*, 365: 233–239.
 43. **Majithiya J. B., Paramar A. N. and Balaraman R. (2005):** Pioglitazone, a PPAR agonist, restores endothelial function in aorta of streptozotocin-induced diabetic rats. *Cardiovascular Research*, 66(1):150-161.
 44. **Martinez G., AL- Dalain M.S., Menendez S., Giuliani and Leon O.S., (2005):** Ozone Treatment Reduces Blood Oxidative Stress and Pancreas Damage in a Streptozotocin Induced Diabetes Model in Rats. *Acta Farm. Bonaerense*, 24 (4): 491-7.
 45. **Martinez- Sanchez G., Al- Dalain S., Silvia M., Lamberto R., Attilia G., Eduardo C., Hector A., Ignacio F. and Sonia L . (2005):** Therapeutic efficacy of ozone in patients with diabetic foot. *European Journal of Pharmacology*, 523: 151-161.
 46. **Mironava M., Klein R., Virella G. and Lopes M. (2000):** Antimodified LDL antibodies, LDL-containing immune complexes and susceptibility of LDL to in vitro oxidation in patients with type2 diabetes. *Diabetes*, 49:1033–1049.
 47. **Naowaboot J, Pannangpetch P, Kukongviriyapan V, Kukongviriyapan U Nakmareong S, Nakmareong S and Itharat A (2009):** Mulberry leaf extract restores arterial pressure in streptozotocin induced chronic diabetic rats. *Nutr Res.*, 29(8):602-8.
 48. **Niraj M. , Miguel A., Carlo G. , Xiaoxu S., Swati D., Genaro R., Brian O, Wei D. and Sonia C. (2014):** Mechanisms of ROS Balance and Cardiac Energy Metabolism in Diabetes Mellitus. *American Journal of Physiology - Heart and Circulatory Physiology*, 308 (4): H291-H302.
 49. **Nishikawa T., Edelstein D. and Du X.L., (2000):** Normalizing mitochondrial superoxide production blocks three pathways of hyperglycaemic damage. *Nature*, 404: 787-790.
 50. **Ofer M., Eyal A., Deborah E., Herzl S., Dani B., Ari Z. and Shuli S. (2007):** Ozone Administration Reduces Reperfusion Injury in an Isolated Rat Heart Model. *J Card Surg.*, 22:339-342.
 51. **Pandey N.M. and Rizvi S.I. (2010):** Protein oxidation biomarkers in plasma of type 2 diabetic patients. *Clinical Biochemistry*, 43(4-5): 508–511.
 52. **Potenza M. A., Addabbo F. and Montagnani M. (2009):** Vascular actions of insulin with implications for endothelial dysfunction. *Am J PhysiolEndocrinolMetab.*, 297: E568- E577.
 53. **Ryuichi A., Masao N., Yaping G., Hideki K., Tomoichiro A., Weidong Z., Ryoza N. and Issei K. (2000):** Insulin Prevents Cardiomyocytes From Oxidative Stress–Induced Apoptosis Through Activation of PI3 Kinase/Akt. *Circulation*, 102:2873-2879.
 54. **Ryuko A., Shingo S., Kazuaki H., Masayuki T. and Seibu M. (2007):** Exacerbation of acidosis during ischemia and reperfusion arrhythmia in hearts from type 2 Diabetic Otsuka Long-Evans Tokushima Fatty rats. *Cardiovascular Diabetology*, 6:17-29.
 55. **Schermer S (1968):** Rats Haemopoietic system In: Blood morphology of laboratory animals. 1st edition, Pb1. Davis. F.A. Co., Philadelphia Chap.10, p. 112.
 56. **Scientific Committee (1982):** Recommendations for measurement of lactate dehydro-

- genase enzyme in human serum .Ann BiolClin., 40: 87-164.
57. **Seiichi T., Takahito K., Kazuhiro Y., Junichi H. , Yoshinori O. and Yoshikazu K. (2004):** Effect of insulin on impaired antioxidant activities in aortic endothelial cells from diabetic rabbits. *Science Direct*, 41(10):1053–1058.
 58. **Sharma S, Adroque JV, Golfman L, Uray I, Lemm J, Youker K, Noon GP, Frazier OH and Taegtmeier H (2004):**Intramyocardial lipid accumulation in the failing human heart resembles the lipotoxic rat heart. *FASEB J.*, 18: 1692–1700.
 59. **Shin I., Sohn J., Park K., Chang K., Choi J., Lee H. and Chung Y. (2006):** A supraclinical dose of tramadol stereoselectively attenuates endothelium- dependent relaxation in isolated rat aorta. *Anesth Analg.*, 103 (2): 366-371.
 60. **Sindhu J., Christian K., and Nosratola D. (2004):** Dysregulation of Hepatic Superoxide Dismutase Catalase and Glutathione Peroxidase in Diabetes Response to Insulin and Antioxidant Therapies. *clinical and experimental hypertension*, 26(1): 43–53.
 61. **Sout RW. (2005):** Diabetes and atherosclerosis. *Biomed. Pharmacother.*, 47: 1-2.
 62. **Suzuki O., Matsubara T., Kanashiro M., Nakao M., Nishimura H., Haruta K., Ikeda T. and Sakamoto N. (1993).** Are diabetic hearts more resistant to ischaemia reperfusion injury? *Jpn. Circ. J.*, 57:328 – 334.
 63. **Suzuki S., Kaneko M., Chapman D. and Dhalla N. (1997):** Alterations in cardiac contractile proteins due to oxygen free radicals. *Biochim Biophys Acta*, 1074:95–100.
 64. **Trinder P (1969):** Enzymatic colorimetric method or determination of blood glucose using an oxidase-peroxidase system, a non-carcinogenic chromagen, cholesterol and triglycerides. *Am ClinBiochem.*, 6: 24-30.
 65. **Udupa AS., Nahar PS., Shah SH., Kshirsagar MJ. and Ghongane BB. (2012):** Study of Comparative Effects of Antioxidants on Insulin Sensitivity in Type 2 Diabetes Mellitus. *J Clin Diagn Res.*, 6(9):1469-73.
 66. **Wali S, Saidu Y., Ladan M., Bilbis L. and Ibrahim N. (2013):** Antioxidant Status and Lipid Profile of Diabetic Rats Treated With Antioxidant Rich Locally Prepared. *Nutriceutical International Journal of Diseases and Disorders*, 1 (2): 033-038.
 67. **Wang S. L., Shang D.J., Wang X.L., You Z.L. and Li H.B. (2011):** Antihyperglycemic and neuroprotective effects of one novel Cu-Zn SOD mimetic. *Bioorganic and Medicinal Chemistry Letters*, 21(14): 4320–4324.
 68. **Wang S.B., Yang X.Y., Tian S., Yang H.G. and Du G.H. (2009):**Effect of salvianolic acid A on vascular reactivity of streptozotocin- induced diabetic rats. *Life Sciences*, 85 (13-14):499-504.
 69. **Wiernsperger NF. (2003):** Oxidative stress as a therapeutic target in diabetes revisiting the controversy. *Diabetes Metab.*, 29:579–585.
 70. **Will C., Catherine L., Robert O. and Maarten K. (2009):** Reduction of heart rate, but not heart rate variability, in diabetic rats. *FASEB J.*, 23: 1019.17.
 71. **Xing W., Yan W., Fu F., Jin Y., Ji L., Liu W., Wang L., Lv A., Duan Y., Zhang J., Zhang H. and Gao F. (2009):** Insulin inhibits myocardial ischemia induced apoptosis and alleviates chronic adverse changes in post-ischemic cardiac structure and function. *Apoptosis*, 14:1050 –1060.
 72. **Zobali F., Avci A., Canbolat O. and Karasu C. (2002):** Effects of vitamin A and insulin on the antioxidative state of diabetic rat heart: a comparison study with combination treatment. *Cell BiochemFunct.*, 20:75– 80.

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خلفية البحث: تعتبر أمراض الشريان التاجي من الأسباب المؤدية للوفاة في مرضي السكر . ولذلك فإن العلاجات الدوائية الحديثة يجب أن توجه ليس فقط لضبط مستوي الجلوكوز بالدم ولكن أيضا إلي منع مضاعفات الجهاز الدوري. و محاولة إعادة الإمداد الدموي لعضلة القلب بعد إصابتها باحتشاء عادة ما تكون مصحوبة بنوع من الإصابة أثناء إعادة التغذية الدموية لعضلة القلب حيث تنتج الشقائق الحرة والتي تلعب دورًا مهما في إحداث عدم التوازن بين المواد المؤكسدة والمواد المضادة للأكسدة. وغاز الأوزون لديه القدرة علي زيادة قدرة مضادات الأكسدة.

الهدف من البحث: دراسة تأثير العلاج بالأوزون و/أو الإنسولين علي تفاعلية الأوعية الدموية وعلي إصابة عضلة القلب بعد إعادة التغذية الدموية لها في الفئران المصابة بمرض البول السكري.

مواد وطرق البحث: أجري هذا البحث علي 250 من الفئران الذكور البيضاء البالغة التي تتراوح أوزانهم بين 120-150+10 جرام وقد قسم هذا العدد إلي: مجموعة ضابطة غير مصابة بمرض السكر (10 فئران) ومجموعة مصابة بمرض البول السكري (240 فأرا). وقد تم حقن الفئران بمادة الاسترنتوزوتوسين . وقد تم تقسيم المجموعة المصابة بمرض البول السكري إلي:

مجموعة مصابة بمرض البول السكري غير معالجة - مجموعة مصابة بمرض البول السكري معالجة بالإنسولين - مجموعة مصابة بمرض البول السكري معالجة بالأوزون - مجموعة مصابة بمرض البول السكري ومعالجة بالأنسولين والأوزون.

وفي كل مجموعة من المجموعات السابقة: تم قياس نسبة السكر والدهون بالدم. - قياس قوة إنقباض عضلة القلب قبل وأثناء وقف التغذية الدموية وبعد إعادة التغذية الدموية. - تسجيل عدد نبضات القلب قبل وأثناء وقف التغذية الدموية وبعد إعادة التغذية الدموية.-- قياس تفاعلية الأوعية الدموية

الإنقباضية للأدرينالين والإندوميثاسين والفازوبرسين وكذلك نسبة الإرتخاء تحت تأثير الأسيتيل كولين ونيتروبروسيد الصوديوم. - قياس إنزيمات مضادات الأكسدة داخل نسيج عضلة القلب. - قياس مستوى إنزيم اللاكتيت ديهيدروجينيز الناتج والمفرز بواسطة عضلة القلب.

النتائج: لوحظ مع العلاج بالأوزون إنخفاض بمستوي السكر والكوليستيرول الإجمالي والدهون الثلاثية والكوليستيرول منخفض الكثافة بالدم وكذلك إنخفاض إنزيم اللاكتيت ديهيدروجينيز، كما لوحظ إرتفاع مستوى الكوليستيرول عالي الكثافة بالدم بعد علاج الفئران بالأوزون، وكانت النتائج ذات دلالة إحصائية بالمقارنة مع المجموعة المصابة بمرض البول السكري الغير معالجة، ولوحظ أيضا إرتفاع مستوى مضادات الأكسدة داخل نسيج عضلة القلب في هذه المجموعة وكان ذو دلالة إحصائية بالمقارنة بالمجموعة المصابة بمرض البول السكري الغير معالجة. كما لوحظ إرتفاع عدد دقات القلب وقوة القلب الانقباضية قبل وأثناء منع التغذية الدموية في هذه المجموعة، وكان ذو دلالة إحصائية بالمقارنة بالمجموعة المصابة بمرض البول السكري الغير معالجة. كذلك لوحظ أيضا إنخفاض في عدد دقات القلب مصحوب بارتفاع في قوة القلب الإنقباضية بعد إعادة التغذية الدموية في هذه المجموعة وكان ذو دلالة إحصائية بالمقارنة بالمجموعة المصابة بمرض البول السكري الغير معالجة. وقد كانت هناك إنخفاضات ذات دلالة إحصائية في إستجابة وتفاعلية شريان الأبهر للانقباض نتيجة الأدرينالين والإندوميثاسين والفازوبرسين، كما كانت هناك إرتفاعات في نسبة الإرتخاء وتفاعلية شريان الأبهر للأسيتيل كولين في هذه المجموعة، وكانت ذات دلالة إحصائية بالمقارنة بالمجموعة المصابة بمرض البول السكري الغير معالجة، كما لم تتغير نتائج إستجابة شريان الأبهر لمادة نيتروبروسيد الصوديوم عند مقارنتها بالمجموعة الضابطة.

الإستنتاج: مضاعفات مرض السكر ترجع إلى الإجهاد التأكسدي، والأوزون ينشط مضادات الأكسدة بالجسم.