



كلية معتمدة 2013

Assessment of bacterial contamination in cattle carcasses at Gharbia Abattoirs Saafan Elsaid,¹Reham , A. Amin and ² Eeilwa-Nesreein , Z

¹ Department of Food Hygiene and Control, Faculty of Veterinary Medicine, Benha University

² Animal Health Research Institute; Dokki

ABSTRACT

The bacteriological examinations were carried out to evaluate the degree of bacterial contamination in cattle carcasses at abattoirs of Gharbia governorate. A grand total of 100 random swab samples from cattle carcasses were collected in the year at 2018, under complete aseptic conditions just after washing and before stamping from different abattoirs and transferred without undue delay to the Lab. of Animal Health Research Institute, Tanta lab, and subjected to the bacteriological examination. The obtained results indicated that the mean values of Aerobic Plate Count (APC), at the region of fore quarter, hind quarter and abdomen were $1.09 \times 10^9 \pm 0.47 \times 10^9$, $1.61 \times 10^9 \pm 0.74 \times 10^9$ and $1.19 \times 10^9 \pm 0.78 \times 10^9$ cfu/g, respectively. While the mean values of Coliform count at the previous regions were $4.78 \times 10^5 \pm 3.82 \times 10^5$, $1.30 \times 10^5 \pm 6.57 \times 10^4$ and $1.42 \times 10^5 \pm 1.27 \times 10^6$ cfu/g, respectively, and the mean values of total Staphylococcal count were $2.39 \times 10^7 \pm 8.57 \times 10^6$, $2.76 \times 10^7 \pm 7.50 \times 10^6$ and $3.56 \times 10^7 \pm 1.54 \times 10^7$ cfu/g, respectively. All the obtained results showed non-significant difference ($P < 0.05$).

Keywords: Cattle carcasses, bacteriological examination, aerobic plate count

(<http://www.bvmj.bu.edu.eg>)

(BVMJ-36(2): 247-251, 2019)

1. INTRODUCTION

Meat considered as an important source of protein, fat, vitamins and minerals, low in carbohydrate content and with sufficient water activity, supports the growth of both spoilage and pathogenic bacteria. A great diversity of microbes inhabit fresh meat generally, but different types may become dominant depending on pH, composition, textures, storage, temperature, and transportation means of raw meat (Adu-Gyamfi *et al.*, 2012). The raw meat may harbor many important pathogenic microbes such as *Salmonella spp.*, *E. coli*, and *Staph. aureus*, making the meat a risk for human health, as without the proper handling and control of these pathogens, foodborne illnesses may occur (Nørrung *et al.*, 2009). The World Health Organization (WHO) and the Food and Agricultural Organization (FAO) of the United Nations stated that illness due to contaminated food considered the most widespread health problem and an important cause of reduced economic productivity (Käferstein, 2003). The main sources of the microbial contamination were found to be the hide and hair of the slaughtered animals, deriving mainly from the microflora of soil. The transfer of contaminating organisms from the hide to the underling tissues was found to be replaced with recorded during the stage of skinning

by means of knives, through the hands, arms, the workers clothes and accidental puncture of stomach and intestine (Gracey *et al.*, 1999). Another important source of contamination arises from unwholesome contacts of meat with excretions from skin, mouth, and nose of the abattoir workers (Omoruyi *et al.*, 2011). Aerobic plate count (APC) is the most reliable index of meat quality, sanitary processing and storage life (ICMSF, 1980). *Staphylococcus aureus* is the second most common pathogen associated with outbreaks of food poisoning, its presence in foods is usually indicative for lack of hygiene in food production (Charlier *et al.*, 2009), and can lead to food intoxication (Normanno *et al.*, 2007). Staphylococcal enterotoxins were highly resistant to heat (ICMSF, 1996). There for the present study was aimed to evaluate the degree of bacterial contamination in cattle carcasses through the total APC, Total Coliform count and Total Staphylococcal count.

2. MATERIALS AND METHODS

2.1. Collection of samples

A grand total of 100 random swab samples from cattle carcasses were collected from different abattoirs in Gharbia Governorate under complete

aseptic conditions just after washing and before stamping. The collected swab samples which are moistened in rinsing fluid solution (buffered peptone water 0.1%) and preserved in an insulated ice box under complete aseptic conditions and transferred without undue delay to the Laboratory of Animal Health Research Institute, Tanta lab, and subjected to the bacteriological examination.

2.2. Preparation of sample (APHA, 2001)

Swabs from cattle carcasses surfaces were taken by using sterile cotton swabs moisturized in rinsing fluid solution (buffered peptone water 0.1%), and templates. The sterilized template was placed firmly against the surface of examined area. The sterile cotton swab was drawn from screw capped plastic tube and then rolled over the limited area inside the template in one direction and perpendicular to this direction to represent all the examined area. Finally, the cotton swabs were retained into the screw capped tubes containing 10ml of sterile buffered peptone water (0.1%) and transferred immediately as possible in an ice-box to the laboratory and ten-fold serial dilutions up to 10^7 were prepared.

2.3. Bacteriological examination

2.3.1. Aerobic plate count (ISO, 2013)

One ml from each of the previously prepared serial dilutions was poured into two separate sterile Petri dishes, using pour plate method, to which approximately 15 ml of sterile melted and tempered plate count agar (45°C) were poured. After thorough mixing, the inoculated and control plates were allowed to solidify at room temperature before being incubated in an inverted position at 37°C for 24 hours. Total aerobic plate count (cfu/g) per gram was calculated on plates containing 30-300 colonies and recorded

2.3.2. Coliform count (ISO, 2006)

All dark red colonies on Violet Red Bile agar plates were enumerated and the average number of coliforms per gm of the sample was recorded

2.3.3. Staphylococcal count (ISO, 2007)

All yellow colonies surrounded by halo zone on Baird parker medium plates were enumerated and total Staphylococci count (cfu/g) were calculated and recorded

3. RESULTS

It is evident from the results recorded in Table (1) that the mean values of APC(/gm) in the examined swab samples of fresh meat of cattle carcasses at the region of fore quarter, hind quarter and abdomen were $1.09 \times 10^9 \pm 0.47 \times 10^9$, $1.61 \times 10^9 \pm 0.74 \times 10^9$ and $1.19 \times 10^9 \pm 0.78 \times 10^9$ cfu/g, respectively, with no significant difference ($P < 0.05$) between the sites of examined surface. From the results achieved in Table (2), it is revealed that the mean values of total coliform counts in the examined swab samples of fresh meat of cattle carcasses at the region of fore quarter, hind quarter and abdomen were $4.78 \times 10^5 \pm 3.82 \times 10^5$, $1.30 \times 10^5 \pm 6.57 \times 10^4$ and $1.42 \times 10^5 \pm 1.27 \times 10^6$ cfu/g, respectively, with no significant difference ($P < 0.05$) between the sites of examined surface. Furthermore the results achieved in Table (3), revealed that the mean values of total Staphylococcal counts in the examined swab samples of fresh meat of cattle carcasses at the region of fore quarter, hind quarter and abdomen were $2.39 \times 10^7 \pm 8.57 \times 10^6$, $2.76 \times 10^7 \pm 7.50 \times 10^6$ and $3.56 \times 10^7 \pm 1.54 \times 10^7$ cfu/g, respectively, with no significant difference ($P < 0.05$) between the sites of examined surface

Table (1) Statistical analytical results of APC (cfu/g) of the examined samples of cattle forequarter, hindquarter and abdomen

Type of Sample	Min.	Max	Mean \pm S.E
Fore quarter (n=40)	<10	1.26×10^{10}	$1.09 \times 10^9 \pm 0.47 \times 10^{9a}$
Hind quarter (n=40)	<10	2.50×10^{10}	$1.61 \times 10^9 \pm 0.74 \times 10^{9a}$
Abdomen (n=20)	<10	1.16×10^{10}	$1.19 \times 10^9 \pm 0.78 \times 10^{9a}$

Means within a column followed by same letters showed non-significant difference ($P < 0.05$)

Table (2) Statistical analytical results of coliform count (cfu/g) of the examined samples of cattle forequarter, hindquarter and abdomen

Type of Sample	Min.	Max	Mean ± S.E
Fore quarter (n=40)	<10	1.53x10 ⁷	4.78x10 ⁵ ± 3.82x10 ^{5a}
Hind quarter (n=40)	<10	2.58x10 ⁶	1.30x10 ⁵ ± 6.57x10 ^{4a}
Abdomen (n=20)	<10	2.54x10 ⁷	1.42x10 ⁵ ± 1.27x10 ^{6a}

Means within a column followed by same letters showed non-significant difference (P < 0.05)

Table (3) Statistical analytical results of staphylococci count (cfu/g) of the examined samples of cattle forequarter, hindquarter and abdomen

Type of Sample	Min.	Max	Mean ± S.E
Fore quarter (n=40)	<10	2.80x10 ⁸	2.39x10 ⁷ ± 8.57x10 ^{6a}
Hind quarter (n=40)	<10	2.20x10 ⁸	2.76x10 ⁷ ± 7.50x10 ^{6a}
Abdomen (n=20)	<10	2.40x10 ⁸	3.56x10 ⁷ ± 1.54x10 ^{7a}

Means within a column followed by same letters showed non-significant difference (P < 0.05)

4. DISCUSSION

The bacterial contamination of the carcass occurs mainly during processing and manipulation, such as skinning, evisceration, storage and transportation. The carcass dressing and evisceration processes constitute critical points in the microbial contamination of muscle. Moreover, fecal matter was a major source of contamination could reach the carcass through direct deposition, as well as by indirect contact through contamination with equipment, workers, installations and air (Abdallah et al., 2009). Table (1) the current results were nearly similar to those reported by Mukhopadhyay et al. (2009), above 10⁷, Nafisa et al. (2010), between 10⁸ –10¹⁰, Tafesse (2014), 1.2 x10⁸, Bogere and Baluka (2014), 1.64x10⁹ and Nahla-Boshra (2017),1.4x10⁶. While, higher results were obtained by Ali et al. (2010), 1.58 x 10¹⁰. Furthermore, lower APC obtained by El-Dally (1994), 4.7 x 10³, Rahkio and Korkeala (1996), 4.5 × 10², Abdallah et al. (2009), 6.2 × 10², Antwi-Agyei and Maalekuu (2014), 2.13x10⁵, Hassan et al. (2016), 2.26× 10⁴ and Salem-Amani et al. (2017), 1.5 × 10⁴. Although the APC of any food articles are not a sure indicative of their safety for consumption, yet it is of supreme importance in judging the hygienic condition under which food has been produced, handled and stored (Levine, 1987). Accordingly, the level of the APC is generally accepted as a criterion for microbial contamination of carcasses and a useful indicator of hygiene (Zweifel and Stephan, 2003). The higher bacterial contamination in fresh meat obtained in this study might be attributed to unhygienic and improper handling of animals during slaughtering, dressing and evisceration, in addition to that there is no doubt that wiping clothes used by slaughter personnel for cleaning up the carcasses could be an important source of

contamination of carcasses. Moreover, the wiping clothes used were not sterile and one wiping cloth

was used for a number of continuous carcasses (Akafete and Haileleul,2011). Table (2) the obtained results were almost agree with that reported by Mukhopadhyay et al. (2009), 6.9 × 10⁵, Samaha (2011), 1.1 x10⁴, Gebeyehu et al. (2013), 53 x10⁵ and Magdy, (2014), 4.4 x10⁴. While higher results were obtained by Nnachi and Ukaegbu (2014), 7.8 x10¹³ and Victoria et al. (2014), 2.7 x10⁶, furthermore, lower coliform count obtained by Bogere and Baluka (2014), 5 x10², Singh et al. (2014), 1.1 x10³, Hassan et al (2016), 5.54 x10² and Nahla-Boshra (2017), 1.3 x10³. Meat contamination with coliforms indicates poor hygienic conditions of carcass processing (Kornacki, 2011). High contamination level of Coliforms in fresh meat may indicate unsanitary conditions. They are indicators of fecal pollution which begin from skinning and direct contact with knives and workers hands. Also, during evisceration and washing, contamination may come from intestinal contents as well as from water during rinsing and washing of carcasses. Table (3) the achieved results nearly similar to results obtained by Nnachi and Ukaegbu (2014), 7.6 x10⁷ and Tafesse et al. (2014), 5.5 x10⁶. While lower result was obtained by Darweesh (2004), 1.6 x10⁴, Salama (2013), 7.3 x10³, Ismail-Eman (2015), 1.56 x10³, Hassan et al. (2016), 1.28 x10³ and Nahla-Boshra (2017), 6.6 x10². The total Staphylococci count can be taken as index of sanitary conditions under which meat and its products are manufactured and handled. Staphylococci can be carried on hands, nasal passage or throats. Most food borne illness outbreak is originated as a result of contamination from meat handlers and production of heat stable toxins in meat (Potter, 2001).

5. REFERENCES

Abdallah, M.A., Suliman, S.E., Ahmed, D.E. and Bakhiet, A.O. 2009. Estimation of bacterial contamination of indigenous bovine

- carcasses in Khartoum (Sudan). *Afri. J. Microbio. Res.*, 3(12):882-886.
- Adu-Gyamfi, A., Torgby-Tetteh, W. and Appiah, V. 2012. Microbiological quality of chicken sold in Accra and determination of D10-value of *E.coli*. *Food Nutr. Sci.* 3 (5): 693-698.
- Akafete, T. and Haileleut, N. 2011. Assessment of risk factors and prevalence of salmonella in slaughtered small ruminants and environment in an export abattoir, Modjo, Ethiopia. *American-Eurasian J. Agric. Environ. Sci.*, 10 (6):992-999.
- Ali, N.H., Farooqui, A., Khan, A., Khan, A.Y. and Kazmi, S.U. 2010. Microbial contamination of raw meat and its environment in retail shops in Karachi, Pakistan. *J. Infect. Dev. Ctries.*, 4(6):382-388.
- Antwi-Agyei, P. and Maalekuu, B.K. 2014. Determination of microbial contamination in meat and fish products sold in the Kumasi metropolis (A Case Study of Kumasi central market and the Bantama market). *Journal of Agricultural Science and Soil Sciences.* 2(3):38-46.
- APHA (American Public Health Association). 2001. 1st NW Washington DC2000, 1-3710 Compendiums of methods for microbiological examination of food. 4th ED. 365-366.
- Bogere, P. and Baluka, S.A. 2014. Microbiological Quality of Meat at the Abattoir and Butchery Levels in Kampala City, Uganda. *Internet Journal of Food Safety.* 16: 29-35.
- Charlier, C., Cretenet, M., Even, S. and Loir, Y.L. 2009. Interactions between *Staphylococcus aureus* and lactic acid bacteria: an old story with new perspectives. *Int. J. Food Microbiol.*, 131(1):30-39.
- Darweesh, M.A.F. 2004. Microbiological picture of retailed meat in menofiya Governorate. M.V.Sc., Fac. Vet. Med. Alexandria Univ., Egypt.
- El-Dally, K.M.A. 1994. Correlation between parasitism and microbial load and meat quality of the Egyptian food animals. Ph. D. (Meat Hygiene), Fac. Vet. Med. Moshtohor, Zagzige Univ.
- Gebeyehu, A., Yousuf, M. and Sebsibe, A. 2013. Evaluation of Microbial Load of Beef of Arsi Cattle in Adama Town, Oromia, Ethiopia. *J Food Technol.* 4: 230-234.
- Gracey, J. F., Collins, D. S. and Huey, R. J. 1999. *Meat hygiene*. 10th Ed. W. B. Saunders Co. Ltd. London.
- Hassan, M.A., Heikal, G.I. and Barhoma, R.M. 2016. Bacteriological Evaluation of Cattle Carcasses at Qaliubiya Abattoirs. *Benha Journal of Applied Sciences (BJAS)*. Vol.1,(2): 39-44.
- International commission on microbiological specifications for food "ICMSF" .1980. *Microbial ecology of foods*. Vol. 1, Academic Press, New York, Toronto.
- International commission on microbiological specifications for food "ICMSF". 1996. *Staphylococcus aureus*. Ch.17 *Microorganisms in food 5: Microbiological specification of food pathogens*. Blackie Academic and Professional, London.
- International Specifications Organization "ISO". 2006. *Microbiology of Food and Animal Feeding Stuffs- Horizontal method for the enumeration of Coliforms—Colony count technique*.
- International Specifications Organization "ISO". 2007. *Microbiology of Food and Animal Feeding Stuffs – Guidelines on Preparation and Production of Culture Media- Part 2: Practical Guidelines on performance Testing of Culture Media*. ISO, Geneva.
- International Specifications Organization "ISO", 2013. *Microbiology of food chain – Horizontal method for the enumeration of microorganisms- Part 1: Colony count at 30 degrees C by the pour plate technique*.
- Käferstein, F. K. 2003. Actions to reverse the upward curve of foodborne illness. *Food Control.* 14 (2): 101-109.
- Kornacki, J.L. 2011. *Practical Sampling Plans, Indicator Microorganisms, and Interpretation of Test Results from Troubleshooting*, Chapter 25. In: Hoorfar, J. (ed), *Rapid Detection, Characterization and Enumeration of Foodborne Pathogens*. ASM Press, Herndon, VA. (manuscript in press).
- Levine, M.M. 1987. *Escherichia coli* that cause diarrhea: enterotoxigenic, enteropathogenic, enteroinvasive, enterohemorrhagic, and enteroadherent. *Journal of Infectious Diseases* vol.155, pp.377-389.
- Magdy, S. M. E. 2014. Food safety of cattle meat and offal at abattoir level. M.V.Sc. Thesis (Meat Hygiene) Fac. Vet. Med. Benha Uni.
- Mukhopadhyay, H.K., Pillai, R.M., Pal, U.K. and Kumar, V.J.A.J. 2009. Microbial quality of fresh chevon and beef in retail outlets of Pondicherry. *Tamilnadu J. Vet. Ani. Sci.*, 5 (1):33-36.

- Nafisa, H.A., Farooqui, A., Khan, A., Ameera, Y. K. and Shahana U. K. 2010. Microbial contamination of raw meat and its environment in retail shops in Karachi, Pakistan. *J Infect Dev Ctries* 4(6):382-388.
- Nahla-Boshra, M. 2017. Assessment of bacterial load of raw meat at abattoir and retail outlets. M. V. Sc., Thesis (Meat Hygiene), Fac. Vet. Med. Zagazig Univ.
- Nnachi, A. U. and Ukaegbu, C. O. 2014. Microbial Quality of Raw Meat Sold in Onitsha, Anambra State, Nigeria. *International Journal of Science and Research (IJSR)*. 3(2):214-218.
- Normanno, G., Salandra, G.L., Dambrosio, A., Quaglia, N.C., Corrente, M., Parisi, A., Santagada, G. Firinu, A., Crisetti, E., Celano, G.V. 2007. Occurrence, characterization and antimicrobial resistance of enterotoxigenic *Staphylococcus aureus* isolated from meat and dairy products. *Int J Food Microbiol.*, 115(3):290-29.
- Nørrung, B. Andersen, J. K. and Buncic, S. 2009. Main concerns of pathogenic microorganisms in meat safety of meat and processed meat. F. Toldrá, ed. (Springer New York).3-29.
- Potter, N.N. 2001. Food Science, the AVI Publishing Co. Inc. New York, 3rd Ed., 2001.
- Rahkio, M. and Korkeala, H. 1996. Microbiological examination of carcasses related to hygienic practice and facilities in slaughtering lines. *Acta. Vet. Scand.*, 37:219-228.
- Salama, E. A. 2013. Sanitary conditions of cattle and camel carcasses at Menofia Abattoirs. Ph.D., Thesis (Meat Hygiene), Fac. Vet. Med. Benha Univ.
- Salem-Amani, M., Zakaria, E.M. and Abd El-Raheem, K.A. 2017. Environmental contamination with methicillin Resistant Staphylococci in food animal carcasses. *BVMJ*, Vol.32, (1): 170-176.
- Samaha, I.A., Ibrahim, H.A. and Aboukahf, H.A. 2011. Enterobacteriaceae in retailed meat. *Alexandria Journal of Veterinary Sciences*. 34(1): 1-9.
- Singh, V.K., Jain, U., Yadav, J.K. and Bist, B. 2014. Assessment of bacterial quality of raw meat samples (carabeef, chevon, pork and poultry) from retail meat outlets and local slaughter houses of Agra Region, India. *Journal of Foodborne and Zoonotic Diseases*, 2 (1):15-18.
- Tafesse, F., Desse, G., Bacha, K. and Alemayehu, H. 2014. Microbiological quality and safety of street vended raw meat in Jijiga town of Somali Regional State, southeast Ethiopia. *African Journal of Microbiology Research*. 8(48): 3867-3874.
- Victoria, O. A., Hezekiah, K. A., Charity, A. A. and Tajudeen, O. I. 2014. Bacterial Load and Antimicrobial Profile of *Escherichia coli* and *Listeria* spp. Isolates from Muscle Tissues of Slaughtered Cattle at a Major Abattoir in Ibadan, South-Western Nigeria. *Journal of Basic and Applied Sciences*, 10, 299-305.
- Zweifel, C. and Stephan, R. 2003. Microbiological Monitoring of sheep carcass contamination in three Swiss abattoirs. *Journal of Food Protection*. vol.66, pp.946- 952.