EFFECT OF FEEDING HELIANTHUS TUBEROSUS (JERUSALEM ARTICHOKE) ON RATIONS NUTRITIVE VALUE AND SOME BLOOD PARAMETERS OF OSSIMI RAMS

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SUMMARY

The present study was carried out at farm and laboratory of Animal Production Department- Faculty of Agriculture, Fayoum University, Egypt. *Helianthus tuberosus* in Ossimi rams rations were evaluated through digestibility, feeding values and some blood parameters. Ossimi rams weighed 45 ± 5 kg in average were used to evaluate the rations supplemented by *Helianthus tuberosus*. The tested rations were D₁ (control) composed of 60% concentrate mixture (CM) + 40% wheat straw (WS); D₂, 52.5% CM + 7.5% *Helianthus tuberosus* (HT) +40% (WS); D₃, 45% CM +15% HT +40% WS and D₄, 30% CM +30% HT +40% WS. The results revealed that Ossimi rams fed rations of D₂ and D₃ showed higher significantly values (P<0.05) of most digestibility coefficients and feeding values compared with control ration. While, insignificant differences were found between D₁ and D₄ regarding digestibilities and feeding values. The obtained results of some blood parameters showed the same trend of rations digestibilities. Adding *Helianthus tuberosus* in the rations of D₂ and D₃ (7.5 and 15% HT) of Ossimi rams improved nutrients digestibility and feeding values and some blood parameters.

Keywords: Helianthus tuberosus, rams, digestibility coefficients, feeding values and blood parameters.

INTRODUCTION

Helianthus tuberosus (Jerusalem artichok), a plant of the Asteraceae family that grows in cool to warm climates, also cultivated widely in China for its highly adaptability and multiple tuber usability options, and its tubers can be produced world-wide, including Asia, Europe, and North America. (Slimestad *et al.*, 2010 and Ma, 2011).

Helianthus tuberosus is a valuable fodder, technical and food culture that are becoming more widely used in various industries (Ryazanova *et al.*, 1997). Also, *Helianthus tuberosus* useful components intended for use as raw materials for foods. Powder *Helianthus tuberosus* flour contains complex carbohydrate components, represented mainly polysaccharide nature inulin (up to 82%), proteins (up to 7%), fat (0.3-0.7%), vitamins (B1, B2, C), pectin (10%), fiber (7%), organic acids, macro- and trace elements (Sidorenko and Gorshkov, 2003 and Liflyandskiy, 2006)

Helianthus tuberosus contains many compounds including coumarines, unsaturated fatty acid, polyacetylenic derivatives (Matsuura *et al.*, 1993) and sesquiterpenes (Baba *et al.*, 2005). Therefore, it has various pharmacological activities, such as cholagogue, aphrodisiac, aperient, stomachic, diuretic, and tonic effects. Also, its tuber, a potential source of biomass, is used as food and folk medicine for the treatment of diabetes and rheumatism due to the presence of inulin, which can be converted into fructose. Moreover, it was found that the extracts of the aerial part were also possess antifungal, antimicrobial, and anticancer activities. (Pan *et al.*, 2009) Additionally, *Helianthus tuberosus* leaf is a natural medicine for the treatment of skin wound, bone fracture and swelling. (Baba *et al.*, 2005).

Inulin and oligofructose are present naturally in several fruits and vegetables like Jerusalem artichoke, chicory, onion, garlic, banana and others. *Helianthus tuberosus* is a plant that can serve as an alternative source of carbohydrates. Certain dietary oligosaccharides, such as inulin and oligofructose, are considered as prebiotics and they are possible substitutes for antibiotics (Best, 2000). Other benefits noted with fructooligosaccharides (FOS) or inulin supplementation include increased production of beneficial short-chain fatty acids such as butyrate, Increased absorption of calcium and magnesium, and improved elimination of toxic compounds (Tomomatsu, 1994 and Van den Heuvel *et al.*, 1999).

This study aimed to investigate the effect of partial replacement of concentrate mixture by *Helianthus tuberosus* in Ossimi rams ration on digestibility coefficient, feeding values and some blood parameters.

MATERIALS AND METHODS

The experiment was carried out at the Experimental Farm Station, Faculty of Agriculture, Fayoum University, Egypt to study the effect of supplementing *Helianthus tuberosus* replacement of concentrate mixture in ration of Ossimi rams on feed intake, nutrient digestibility, feeding values and some blood parameters.

Experimental animals

Twelve Ossimi rams weighted 45 ± 5 Kg in average were divided into 4 similar groups (3 rams each) and fed on the tested ration (Table 1). The tested rations were: the control ration (D₁) composed of 60% concentrate mixture (CM) plus 40% wheat straw (WS). Concentrate mixture were replaced by 12.5, 25 and 50% of *Helianthus tuberosus* (HT) on dry matter basis for D₂, D₃ and D₄, respectively. These replacements formed 7.5, 15 and 30% of the total rations dry matter for D₂, D₃ and D₄, respectively. Thus, four tested rations were performed.

Digestibility trial

Digestibility trial was carried out to evaluate the nutrients digestibility and nutritive value of the tested rations. The animals were individually placed in metabolic cage for 15 day as a preliminary period followed by 7 days as a collection period. The ration was offered daily and refused if found were recorded every day. Total feces were daily collected and weighted. Feces sample (10%) was taken, sprayed with 10% H_2SO_4 and dried at 60°c for 24 hours, then it was ground and kept for chemical analysis. Samples of offered feeds were composite dried separately, finally ground and kept for chemical analysis according to the methods of the A.O.A.C. (1995). Gross energy (GE) and digestible energy (DE) of feeds were calculated after Nehring and Haenlien (1973).

Blood analysis

Blood samples were taken at the end of the digestion trail before the morning feeding from the jugular vein. The samples were directly collected into vacuum tube and centrifuged at 3000 rpm for 5 min. serum was separated into polypropylene tube and stored at -18 °C until analysis for total protein and albumin according to Weichselbaum, (1946) and Doumas *et al.* (1971), respectively. Globulin value was calculated by the differences between total protein and corresponding value of albumin. The total lipids was determined following method of Boutwell (1972), while cholesterol following Allain *et al.* (1974).

Statistical analysis

Data of the digestibility trial and some blood parameters were analyzed using general linear model procedure by computer program of SPSS (1997) and the differences between means were tested using Duncans new multiple test (Duncan, 1955). The statistical model was as follows: Yij = m + Ai + eij Where Yij is dependent variable; m= overall mean; Ai= is the effect of treatment and eij= is the experimental error.

Item	Tested rations					
	D ₁ (control)	D ₂	D ₃	D_4		
Concentrate mixture*, %	60	52.5	45	30		
Wheat straw, %	40	40	40	40		
Helianthus tuberosus,%		7.5	15	30		

Table (1): Formula of the tested rations used in digestibility trial, on dry matter basis.

* Concentrate mixture consisted of 22% soybean cake, 50% yellow corn, 10% rice bran, 10% wheat straw, 3.5 % molasses, 3% lime stone, 1% common salt and 0.5% minerals mixture.

RESULTS AND DISCUSSION

Chemical composition of the tested ration

Chemical composition of concentrate mixture, wheat straw, *Helianthus tuberosus* and tested rations are presented in Table (2). *Helianthus tuberosus* was higher of nitrogen free extract (69.65%) compared with concentrate mixture which was 48.61%. Also, crude protein of HT was 9.8% which was nearly similar to yellow corn. Crude fiber of HT was very lower (4.14%) than concentrate mixture which 15.32% was. Ash of HT had the same trend of crude fiber but gross energy (GE) of HT was similar to concentrate mixture and wheat straw. Also, chemical composition of *Helianthus tuberosus* was nearly similar to yellow corn. Thus, has been replaced to concentrate mixture.

	-	% on DM basis						
Item	DM%	OM%	CP%	EE%	CF%	NFE%	Ash%	GE [*] Mcal/ kgDM
Concentrate mixture	91.44	84.09	16.54	3.62	15.32	48.61	7.35	3.84
Helianthus tuberosus	92.07	85.78	9.80	2.19	4.14	69.65	6.29	3.83
Wheat straw	91.54	79.81	3.89	1.85	33.42	40.65	11.73	3.81
D ₁ (control)	91.48	82.38	11.48	2.91	22.56	45.43	9.10	3.84
D_2	91.53	82.50	10.97	2.80	21.72	47.00	9.02	3.83
D_3	91.57	82.63	10.47	2.70	20.88	48.58	8.94	3.81
D_4	91.67	82.89	9.458	2.483	19.21	51.738	8.78	3.78

Table (2): Chemical analyses of ingredients and tested rations (on DM basis).

*GE = tested diet three (30% concentrate mixture+40% wheat straw+30 Helianthus tuberous)

Digestibility and nutritive values

The results in Table (3) showed that digestion coefficient of dry matter (DM%) was 58.83% in control diet (D₁) and was 63.04%, 62.08% and 59.31% of D₂, D₃ and D₄, respectively with significant differences (P<0.05). Organic matter (OM %) digestibility almost followed the same trend of DM digestibility. Digestion coefficient of crude protein (CP %) ranged between 58.52% and73.59% being significantly (P<0.05) higher in D₂ than those of D₄ and control one. Digestion coefficient of ether extract (EE %) was nearly similar to the trend of CP digestibility. Generally, ration D2 (7.5 % *Helianthus tuberosus) showed* the highest digestibilities of all nutrients especially when compared with D₁ (control ration) or D₄.

	Tested Rations					
Item	D ₁ (control)	D_2	D ₃	D_4	\pm SE	
Digestion coefficients %						
DM	58.83 ^c	63.04 ^a	62.08^{ab}	59.31 ^{bc}	± 0.651	
OM	62.99 ^c	67.81 ^a	65.87 ^b	63.16 ^c	± 0.685	
CP	58.52 ^b	73.59 ^a	69.58 ^{ab}	59.53 ^b	± 2.509	
EE	75.44 ^b	79.85 ^a	77.91 ^{ab}	76.17 ^{ab}	± 0.687	
CF	52.68 ^c	59.00 ^a	56.45 ^b	51.51 ^c	± 0.942	
NFE	68.99 ^b	73.93 ^a	72.67^{a}	70.38 ^b	± 0.643	
Feeding values						
TDN,%	54.88 ^b	60.67^{a}	59.10 ^a	56.17 ^b	± 0.312	
DCP, %	6.72 ^{bc}	8.07^{a}	7.28^{ab}	5.63 ^c	± 0.312	
DE (Mcal/KgDM)	237.59 ^b	263.61 ^a	248.73 ^b	261.82 ^a	± 3.499	
GE (Mcal/KgDM)	384.43	382.80	381.27	378.05	± 0.709	

Table (3): Digestion coefficients and feeding values of the tested rations (on DM basis) fed to Ossimi rams.

Average in the same row with different superscripts are different ($P \le 0.05$) for a,b and c.

These results were in agreement with those obtained by Swanson *et al.* (2002); Strickling *et al.* (2000) and Flickinger *et al.* (2003). Samal *et al.* (2017) showed that supplementation of *Jerusalem artichoke* (JA) to diets improved digestibility coefficient of CP, EE and CF when addition JA tuber powder (JA 0,2,4 and 6% pulverized) in rats and better absorption of calcium and phosphorus. Pradhan *et al.* (2015) observed a reduction in protein digestibility upon dietary supplementation with 2% of *Helianthus tuberosus* in the diet. Increased CF digestibility in *Helianthus tuberosus* -fed rams is in agreement with the findings of Zentek *et al.* (2002) and Samal *et al.* (2012) also recorded increased CF digestibility when *Helianthus tuberosus* was included in the diet.

Pawar (2007) and Kore *et al.* (2012) who used MOS as a prebiotic. The digestibility of CF was improved by dietary supplementation with *Helianthus tuberosus*. The increase in *Lactobacillus* spp. and *Bifidobacterium* spp. populations by *Helianthus tuberosus* addition represents a clear improvement in the status of eubiosis. This may be the result of selective fermentation of inulin and fructooligosaccharides (FOS) by these beneficial bacteria in the hindgut.

Inulin-type fructans are either synthesized from sucrose or prepared commercially from inulin-rich plants such as chicory (*Cichorium intybusL.*) root and *Helianthus tuberosus* tuber. The yield potential, both for biomass and sugars, is higher in *Helianthus tuberosus* than chicory. The *Helianthus tuberosus* plant shows good frost and drought tolerance, is resistant to diseases and can achieve a high yield of biomass (Slimestad *et al.*, 2010). The inulin content in *Helianthus tuberosus* tuber ranges from 7 to 30% of fresh weight (Kays and Nottingham, 2007) or from 60 to 85.5% of dry weight (Aduldecha *et al.*, 2016). Tubers of *Helianthus tuberosus* have high metabolizable energy content of 15 MJ \cdot kg–1 dry matter (Kleessen *et al.*, 2007)

Some blood Parameters

The results of some blood constituents are shown in Table (4). The obtained values were in normal range. Concentration of total protein and albumin were significantly (P<0.05) higher in D_2 which consisted 7.5% *Helianthus tuberosus* than D_1 and D_3 . No significant differences were found between D_2 , D_3 and D_4 regarding globulin concentration. Meanwhile, D_2 and D_3 differed significantly (P<0.05) compared to D_1 (control ration). Concentration of cholesterol and creatinine were increased (P<0.05) significantly in D_1 (control diet) than the other diets. AST and ALT were not significantly different (P<0.05) among treatment. These results indicated that animals were generally in a good nutritional status and livers were in normal physiological conditions. Tiengtam *et al.* (2017) found that supplementation with inulin at 5g/kg or *Jerusalem artichoke*. While Nile tilapia diet it did not affect any significant changes in several blood metabolites, including glucose, cholesterol, triglyceride or albumin. A previous study in juvenile tilapia

both increased glucose, plasma protein and albumin (Tiengtam *et al.*, 2015). Such differences may reflect the effect of animal used or the level of *Helianthus tuberosus* in the ration.

Item		\pm SE			
	D_1 (control)	D ₂	D ₃	D_4	
Total protein(g/dl)	5.72 ^b	6.91 ^a	5.57 ^b	6.48 ^{ab}	0.201
Albumin(g/dl)	2.93°	3.49 ^a	3.17 ^b	3.20 ^b	0.063
Globulin(g/dl)	3.19 ^b	3.55 ^a	3.60^{a}	3.27 ^{ab}	0.067
Cholesterol(mg/dl)	98.40^{a}	94.57 ^{bc}	93.48 ^c	96.91 ^{ab}	0.683
Triglycerides(mg/dl)	51.07 ^a	50.14 ^{bc}	49.83 ^c	50.71 ^{ab}	0.172
Creatinine (mg/dl)	1.20 ^a	1.13 ^{ab}	1.04 ^b	1.11 ^{ab}	0.023
ALT (u/ml)	28	27	27	28	0.328
AST(u/ml)	43	40	41	42	0.835

Table (4): Some blood parameters of tested diets (on DM basis) fed to Ossimi rams.

Average in the same row with different superscripts are different ($P \le 0.05$) for a,b and c.

The *Helianthus tuberosus* addition into diets significantly ($P \le 0.05$) increased total short-chain fatty acids concentrations as the level of Jerusalem artichoke in the diet increased. Also, lactic acid content was increased and ammonia concentration was reduced in feces due to *Helianthus tuberosus* supplementation (Samal *et al.*, 2017). These findings may suggest the obtained results of total protein and triglycerides.

CONCLUSION

Supplementing *Helianthus tuberosus* to the ration of Ossimi rams as replacement with 7.5% and 15% improvement nutrient digestibility, feeding values and some blood parameters. Also, more researches were needed to examine different levels of *Helianthus tuberosus* in the diets with different animal types.

REFERENCES

- A.O.A.C. (1995). Association of Official Analytical Chemists. Official Methods of Analysis. 16th ed. Washington, D.C., USA.
- Aduldecha, C.; W. Kaewpradit; N. Vorasoot; D. Puangbut; S. Jogloy and A. Patanothai (2016). Effects of water regimes on inulin content and inulin yield of Jerusalem artichoke genotypes with different levels of drought tolerance. Turk. J. Agric. For. 40, 335–343, <u>https://doi.org/10.3906/tar-1506-39</u>
- Allain, C. C.; L. S. Poon; C. S. Chan; W. Richmond and P. C. Fu (1974). Enzymatic determination of total serum cholesterol. Clin. Chem., 20:470-475.
- Baba, H.; Y. Yaoita and M. Kikuchi (2005). Sesquiterpenoids from the leaves of *Helianthus tuberosus L*. J Tohoku Pharm Univ. 52:21-5.
- Best, P. (2000): Starter pig feeds: Oligosaccharides. Do these feed sugars assist the right bacteria? Feed Int., 2, 24.

Boutwell, J.H.M. (1972). USD.H.E.W. pamphlet.

Doumas, B.; W. Wabson and H. Biggs (1971). Albumin standards and measurement of serum with bromocresol green. Clinical Chemistry 31:87.

Duncan, D. B. (1955). Multiple range and multiple F test. Biometrics. 11:1-42.

- Flickinger, E. A.; E. M. W. C. Schreijen; A. R. Patil; H. S. Hussein; C. M. Grieshop; N. R. Merchen and Jr. G. C. Fahey (2003). Nutrient digestibilities, microbial populations and protein catabolites as affected by fructan supplementation of dog diets. J. of Anim. Sci., 81: 2008–18.
- Kays, S.J. and S.F. Nottingham (2007). Chemical composition and inulin chemistry. In: Biology and Chemistry of Jerusalem Artichoke: *Helianthus tuberosusL*. CRC Press, Boca Raton, FL (USA), pp. 53– 96, <u>https://doi.org/10.1201/9781420044966.ch5</u>
- Kleessen, B.; S. Schwarz; A. Boehm; H. Fuhrmann; A. Richter; T. Henleand and M. Krueger (2007). Jerusalem artichoke and chicory inulin in bakery products affect faecal microbiota of healthy volunteers. Br. J. Nutr. 98, 540–549, https://doi.org/10.1017/ S0007114507730751.
- Kore, K.B.; A.K. Pattanaik; A. Dasand and K. Sharma (2012). Evaluation of mannanoligosaccharide as prebiotic functional food for dogs: effect on nutrient digestibility, hind gut health and plasma metabolic profile. Indian J. Anim. Sci. 82, 81–86.
- Liflyandskiy, V.G. (2006).Vitamins and minerals. From A to Ya. -Sankt Petersburg: Publishing House "Neva".
- Ma, X.Y; L.H. Zhang; H.B. Shao; G. Xu; F. Zhang and F.T. Ni. (2011). Jerusalem artichoke (*Helianthus tuberosusL.*), a medicinal salt-resistant plant has high adaptability and multiple-use values. J. Med. Plant Res. 5(8):1275-82.
- Matsuura, H.; T. Yoshihara and A. Ichihara (1993). Four New Polyacetylenic Glucosides, Methyl β-D-Glucopyranosyl Helianthenate C-F, from Jerusalem artichoke (*Helianthus tuberosus* L.) Biosci Biotechnol Biochem; 57(9):1492-8.
- Nehring, K. and G.F.W. Haenlien (1973). Feed evaluation and ration calculation based on net energy. J. Anim. Sci., 36: 949.
- Pan, L.; M. R. Sinden; A.H. Kennedy; H. Chai; L.E. Watson and T. L. Graham (2009). Bioactive constituents of *Helianthus tuberosus* L. (Jerusalem artichoke). Phytochem Lett; 2(1):15-8.
- Pawar, M.M. (2007). Optimization of homemade diet through supplementation and use of prebiotics in pet dog .M.V.Sc. Thesis Submitted to IVRI, Deemed University, Izatnagar (India).
- Pradhan, S.K.;A. Das; S.S. Kullu; M. Saini; A.K. Pattanaik; N. Dutta and A.K. Sharma (2015). Effect of feeding Jerusalem artichoke (*Helianthus tuberosus*) root as prebiotic on nutrient utilization, fecal characteristics and serum metabolite profile of captive Indian leopard (Pantherapardusfusca) fed a meaton-bone diet. Zoo Biol. 34, 153–162, https://doi.org/10.1002/zoo.21187.
- Ryazanova, T.V.; L.A. Dorofeeva and A.V. Bogdanova (1997). Chemical composition of the vegetative part of Jerusalem artichoke and its use. Forest magazine.
- Samal, L.; V.B. Chaturvedi and A.K. Pattanaik (2017). Effects of dietary supplementation with Jerusalem artichoke (*Helianthus tuberosus* L.) tubers on growth performance, nutrient digestibility, activity and composition of large intestinal microbiota in rats. J. of Anim. and Feed Sci. 26, 2017, 50– 58,https://doi.org/10.22358/jafs/68779/2017.
- Samal, L.; V.B. Chaturvedi; S. Baliyan; M. Saxena and A.K. Pattanaik (2012). Jerusalem artichoke as a potential prebiotic: influence on nutrient utilization, hindgut fermentation and immune response of Labrador dogs. Anim. Nutr. Feed Technol. 12, 343–352
- Sidorenko, S.M. and Gorshkov (2003). A method for producing dried tubers of Jerusalem artichoke M.F.
- Slimestad, R.; R. Seljasen; K. Meijer and S.L. Skar (2010). Norwegian-grown Jerusalem artichoke (*Helianthus tuberosus L.*): Morphology and content of sugars and fructo-oligosaccharides in stems and tubers. J. Sci. Food Agric., 90 (6):956-64.
- SPSS (1997). Statistical Package for Social Science release 8.0 copyright (c), SPSS INC., Chicago, USA.
- Strickling, J. A.; D. L. Harmon; K. A. Dawson and K. L. Gross (2000). Evaluation of oligosaccharide addition to dog diets: Influences on nutrient digestion and microbial population. Anim. Feed Sci. and Tech., 86: 205–19.

- Swanson, K. S.; C. M. Grieshop; E. A. Flickinger; L. L. Bauer; H. P. Healy; K. A. Dawson; N. R. Merchen and G. C. Jr. Fahey (2002). Supplemental fructooligosaccharides and mannanoligosacchrides influence immune function, ileal and total tract nutrient digestibilities, microbial population and concentrations of protein catabolites in the large bowel of dogs. J. of Nutr. 132: 980–89.
- Tiengtam, N.; P. Paengkoum; S. Sirivoharn; K. Phonsiri and S. Boonanuntanasarn (2017). The effects of dietary inulin and Jerusalem artichoke (*Helianthus tuberosus*) tuber on the growth performance,haematological, blood chemical and immune parameters of Nile tilapia (*Oreochromis niloticus*) fingerlings. Aquaculture Research, 00:1–9.
- Tiengtam, N.; S. Khempaka; P. Paengkoum and S. Boonanuntanasarn (2015). Effects of inulin and Jerusalem artichoke (*Helianthus tuberosus*) as prebiotic ingredients in the diet of juvenile Nile tilapia (*Oreochromis niloticus*). Anim. Feed Sci. and Tech., 207, 120–129.
- Tomomatsu, H. (1994): Health effects of oligosaccharides. A review. Food Technol., 61, 5. University of Maryland Medical Center. Traditional Chinese Medicine. [Updated 24 August 2009; 19 April 2014]; Available from: <u>www.umm.edu/altmed/articles/traditional-chinese-000363.htm</u>.
- Van den Heuvel, E.G; T. Muys; W. van Dokkum and G. Schaafsma (1999). Oligofructose stimulates calcium absorption in adolescents. Am. J. Clin. Nutr., 69, 544–548.
- Weichselbaum, T. E. (1946). Quantitative colorimetric determination of total protein in serum. American J. Clinical Pathology. 7:40-45.
- Zentek, J.; B. Marquart and T. Pietrzak (2002). Intestinal effects of mannan-oligosaccharides, trans galactooligosaccharides, lactose and lactulose in dogs. J. Nutr. 132, 1682–1684.

تأثير التغذية بدرنات نبات الطرطوفة على القيمة الغذائية وبعض مقاييس الدم للأغنام الأوسيمي

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أجريت هذه الدراسة بمزرعة ومعمل قسم الإنتاج الحيواني - كلية الزراعة - جامعة الفيوم - مصر. وتم فيها تقييم درنات نبات الطرطوفة المضافة إلى علائق الكباش الأوسيمي من خلال معاملات الهضم والقيم الغذائية وبعض مقابيس الدم

إستخدام في هذه الدراسة 12 كبش يتراوح وزنها 45 ± 5 كجم في المتوسط، تم استخدامها لتقييم العلائق المحتوية على درنات نبات الطرطوفة وكانت العلائق المختبرة كالتالي: غذيت المجموعة الأولي D1 (المقارنة) على عليقة مكونة من60% خليط مركز +40% تبن قمح، وغذيت المجموعة الثانية D2 علي 52.5% خليط مركز+ 5.5% درنات نبات الطرطوفة +40% تبن قمح، أما المجموعة الثالثة D3 فغذيت على 45% خليط مركز +15% درنات نبات الطرطوفة +40% تبن قمح، بينما غذيت المجموعة الرابعة D4 على 30% خليط مركز +30% درنات نبات الطرطوفة+40% تبن قمح وقد أظهرت النتائج ما يلي:

معاملات هضم المادة الجافة، المادة العضوية، البروتين الخام، الألياف الخام، الدهن الخام، الكربوهيدرات أظهرت قيمًا معنوية (P<0.05) أعلى في العلائق المحتوية علي 7.5% ،15% درنات نبات الطرطوفة مقارنة بالعليقة المقارنة والعليقة D₄ التي تحتوي علي 30% درنات نبات الطرطوفة.

اضافة درنات نبات الطرطوفة أدي إلي تحسين المركبات المهضومة الكلية والبروتين الخام المهضوم تحسين معنوي (0.05 P) في العليقة D₃ ،D₂ والمحتوية علي 7.5% ،15% درنات نبات الطرطوفه مقارنة بالعليقة المقارنة و العليقة الرابعة L₄. النتائج التي تم الحصول عليها من بعض مقاييس الدم أظهرت زياده معنوية ايضا في البروتين الكلي، الالبيومين ، الجلسريدات الثلاثية في العليقة الثانية J₂ والمحتوية على 7.5 % درنات نبات الطرطوفة بالمقارنة بالعلائق الأخرى ، أما بالنسبة للكوليسترول والكرياتينين كانت العليقة الأولي (المقارنة) D أكثر العلائق معنوية عن باقي العلائق الأخرى. اما انزيمات غير معنوية.

نستخلص من هذه الدراسة أن إضافة درنات نبات الطرطوفة إلى علائق الأغنام الأوسيمي بنسبة 7.5% ، 15% من العليقة أدت إلي زياده معدلات هضم المركبات الغذائية وزياده معدل الاستفادة من بروتين العليقة ولم تؤثر على صحة و نشاط الحيوان.