

## **EFFECT OF ADDING SOME ANTIOXIDANTS TO DIET CONTAINING FATTY ACIDS ON PRODUCTIVE AND SOME PHYSIOLOGICAL PARAMETERS OF SILVER MONTAZAH CHICKENS STRAIN. 2-DURING LAYING PERIOD**

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### **SUMMARY**

**A**n experiment was conducted for a period of 12 wks to investigate the effect of some antioxidants in the diet on productive performance and some physiological and hematological parameters as well as fertility and hatchability of chickens. For this purpose, 231 Silver Montazah strain birds (210 hens and 21 cocks) 24-wks-old were used in this experiment up to 36 wks of age. All birds were individually weighed and randomly divided into 7 equal experimental groups (30 hens and 3 cocks of each) with three replicates (10 hens and 1 cock each) with almost similar initial average body weight. Replicates were randomly housed in floor pens (280 cm long x 220 cm wide). The 1<sup>st</sup> group was fed the basal diet that supplemented linoleic acid (n-6) and linolenic acid (n-3) at the ratio of 4:1, respectively and served as control. The 2<sup>nd</sup> and 3<sup>rd</sup> groups were fed the basal diet supplemented with 125 and 250 g Butylated Hydroxy Toluene/ton diet, respectively. The 4<sup>th</sup> and 5<sup>th</sup> groups were fed the basal diet supplemented with 100 and 200 g vit. E /ton diet, respectively. The 6<sup>th</sup> and 7<sup>th</sup> groups were fed the basal diet supplemented with 5 and 10 kg citric acid/ton diet, respectively. Results indicated that antioxidants supplementation significantly increased egg number, egg production percentage and egg mass and improved feed conversion ratio and egg shell thickness compared to the control group. Moreover, significant increase was obtained for blood hemoglobin, red blood cells and white blood cells, as well as plasma total protein, globulin and high density lipoproteins for groups fed antioxidants compared with the control group. Ejaculate volume, sperm motility, live sperm percentage, sperm concentration, total sperm per ejaculate and total live sperm per ejaculate, as well as fertility and hatchability percentages were increased. Conversely, significant lower heterophils/lymphocytes ratio and plasma concentration of total lipids, cholesterol, triglycerides and low-density lipoproteins were associated with supplemental antioxidants, while abnormal sperm percentage was decreased with antioxidants supplementation. In conclusion, dietary supplemental of some antioxidants to feed containing fatty acids is a practical application recommended especially by 100 and 200 g vit. E or 10 kg citric acid/ton diet to Silver Montazah chickens strain during the laying period and had beneficial effects on productive, reproductive, physiological and hematological parameters and immune status.

**Keywords:** *antioxidants, chickens, productive performance, physiological and hematological parameters, fertility and hatchability.*

### **INTRODUCTION**

In avian species, alpha linolenic acid (18:3 n-3) and linoleic acid (18:2 n-6) cannot be synthesized in the body and have to be supplied in the diet, therefore they are called essential fatty acids. Fatty acids are the major components of chicken egg yolk lipids and constitute over 4 g/egg on the average. From a nutritional standpoint of view, yolk fatty acids are the major source of energy and long-chain C<sub>20</sub> and C<sub>22</sub> polyunsaturated fatty acids (PUFA) are essential for developing chick during embryogenesis (Cherian, 2007). It is interest to note that the ratio of n-6/n-3 PUFA appears to be more important for fat and cholesterol metabolism and in modulating antibody synthesis than the absolute concentrations of n-3 PUFA in the diet (Sijben *et al.*, 2000). So, there is a need to regulate the intake of n-6 and n-3 PUFA in balanced proportions. Feeding either 4:1 or 6:1 linoleic acid (LA) to linolenic acid (LNA) containing diet improving productive

performance, enhancing fertility and hatchability, produced low triglycerides, cholesterol and rich in PUFA eggs in laying hens (Radwan *et al.*, 2012).

Synthetic antioxidants, such as butylated hydroxytoluene (BHT), can inhibit lipid oxidation in feed but they have toxic properties (Kahl and Kappus, 1993) resulting in strict regulations over their use in foods. These findings, together with increased resistance to the use of synthetic additives, have increased interest in the antioxidant properties of naturally occurring substances (Gordon, 1996).

Vitamin E (vit. E) is a metabolic nutrient that has received a lot of attention with respect to its importance to the immune response in poultry. However, chicken cannot synthesis vit. E, therefore, its requirements must be given from dietary sources (Chan and Decker, 1994). Also, vit. E has been reported as a natural antioxidant. It prevents the oxidation of unsaturated lipid materials within cells, thus protecting the cell membrane oxidative damage (Gore and Qureshi, 1997). Furthermore, vit. E serves as a physiological antioxidant through inactivation of free radicals. As well as, it improves egg production, feed intake, egg yolk and albumen solids (Kirunda *et al.*, 2001), and improved growth performance and immune response in Inshas chickens (Alm El-Dein *et al.*, 2013).

Citric acid (CA) is one of the most widely used food additives, which is commonly used as a preservative, acidulant, pH control agent, flavor enhancer, and antioxidant in many foods (Kristiansen *et al.*, 1999). Al-Harathi and Attia (2015) found that feed intake and feed conversion ratio were significantly affected by CA supplementation in laying hens, the positive effect of CA on feed intake may be due to the effect of maintaining feed freshness and thus the palatability of the feed. High levels of calcium in the laying hens diets may interfere with CA affecting pH in the gut towards alkalinity, thus reducing its effectiveness as an acidic agent (Nezhad *et al.*, 2007).

Therefore, the objectives of this experiment were to study the effect of adding some antioxidants to feed containing fatty acids on productive and reproductive performance and some physiological and hematological parameters of Silver Montazah (SM) chickens strain during the laying period (24-36 weeks of age).

## **MATERIALS AND METHODS**

This experiment was carried out at Inshas Poultry Breeding Station, Animal Production Research Institute, Agricultural Research Center, Egypt.

### ***Chicks and experimental design:***

A total number of 231 Silver Montazah (SM) strain birds (210 hens and 21 cocks) 24-wks-old were used in this experiment up to 36 wks of age. All birds were individually weighed and randomly divided into 7 equal experimental groups (30 hens and 3 cocks of each) with three replicates (10 hens and 1 cock each) with almost similar initial average body weight. Replicates were randomly housed in floor pens (280 cm long x 220 cm wide). The 1<sup>st</sup> group was fed the basal diet that contained linoleic acid (n-6) and linolenic acid (n-3) at the ratio of 4:1, respectively and served as control. The 2<sup>nd</sup> and 3<sup>rd</sup> groups were fed the basal diet supplemented with 125 and 250 g BHT/ton diet, respectively. The 4<sup>th</sup> and 5<sup>th</sup> groups were fed the basal diet supplemented with 100 and 200 g vit. E /ton diet, respectively. The 6<sup>th</sup> and 7<sup>th</sup> groups were fed the basal diet supplemented with 5 and 10 kg CA/ton diet, respectively.

### ***Managements and feeding:***

All birds were kept under the same managerial hygienic and environmental conditions. Birds were kept in a windowed house with light cycle regimen of 16 h light: 8 h darkness, throughout the experimental period (24-36 wks of age). Feed and water were provided for *ad libitum* consumption. The basal experimental diet was formulated to meet the nutrient requirements of SM laying hens during period from (24-36 wks of age) according to feed composition tables for animal and poultry feed stuffs used in Egypt (2001). The composition and calculated analysis of the experimental basal diet is present in (Table 1).

**Table (1): Composition and calculated analyses of the experimental basal diet.**

Ingredient	Layer diet (%)
	LA:LNA ratio 4:1
Yellow corn	54.30
Soya bean meal 44%	27.30
Wheat bran	5.20
Di Calcium Phosphate	1.57
Limestone	7.95
Vitamins & Minerals premix*	0.30
NaCl (salt)	0.30
DL-Methionine	0.08
Linseed oil	0.83
Soybean oil	2.17
Total	100
Calculated analyses**:	
Crude protein	17.02
Crude fiber	3.81
Ether extract	5.68
Calcium	3.42
Available Phosphorus	0.428
Lysine	0.968
Methionine	0.376
Methionine + Cystine	0.665
ME (kcal/kg diet)	2749.6

\*Premix added to the 1 kg of diet including vit.A 10000 I.U; vit. D<sub>3</sub> 2000 I.U; vit. E 15 mg; vit. K<sub>3</sub> 1 µg; vit B<sub>1</sub> 1mg; vit. B<sub>2</sub> 5mg; vit. B<sub>12</sub> 10 µg; vit B<sub>6</sub> 1.5 mg; Niacin 30mg; Pantothenic acid 10mg; folic acid 1mg; Biotin 50 mg; choline 300 mg; Zn, 50mg; Cu, 4mg; iodine 0.3 mg; Fe, 30mg; Se, 0.1mg; Mn, 60mg; cobalt 0.1mg and carrier CaCo<sub>3</sub> up to 1kg.

\*\* According to feed composition tables for animal and poultry feedstuffs used in Egypt, (2001).

**Laying performance traits:**

Body weights were recorded at the beginning (24 weeks of age) and the end of the experiment (36 weeks of age). Feed intake and feed conversion ratio (FCR), egg number and egg production percentage, egg weight and egg mass (number of eggs x egg weight) were recorded for each replicate at the end of each week from 24 up to 36 wks of age.

**Egg quality parameters:**

A total of 105 eggs (15 eggs from each treatment) were taken after 36 wks of age to determine the interior and exterior egg quality parameters. Eggs were weighed individually then broken and the inner contents were placed on a leveled glass surface to determine the inner egg quality. The inner egg quality parameters included shell weight % (SW), shell thickness, mm (ST) including shell membranes which was measured using a micrometer at three locations on the egg (air cell, equator and sharp end), egg length, cm (EL), egg width, cm (EWd), shape index (SI) which was estimated as the percentage of (EWd) to (EL), albumen weight % (AW), yolk weight % (YW), yolk height, mm (YH), yolk diameter, mm (YD) and yolk index (YI) which was estimated as the percentage of YH to YD.

**Blood biochemical analysis and hematological picture:**

At 36 wks of age (end of experiment), six blood samples were collected from each experimental treatment from the wing vein in two heparinized test tubes. Blood of the first tube was used to evaluate the total count of red and white blood cells, and blood smears were done and stained with Fleishman's stain for the differential counts of leucocytes. The other blood tube was centrifuged at 3000 rpm for 15 min, so plasma samples were harvested and stored at -20°C until the time of biochemical analysis. Commercial kits were used for colorimetrically determination of the following blood plasma constituents according to procedure outlined by the manufacturer. Total protein and albumin, total lipids, total cholesterol, high density lipoproteins (HDL), low density lipoproteins (LDL), and hepatic enzymes including aspartate amino

transaminase (AST) and alanine amino transaminase (ALT) were determined. Globulin was calculated by subtraction of plasma albumin from total protein, and albumin/globulin ratio was calculated.

**Semen quality:**

Semen samples were collected randomly from 21 cocks (3 cocks of each treatment) at 36 weeks of age using the massage method. Semen samples were examined for the following characteristics.

1-The ejaculate volume was determined to the nearest 0.01 ml. using 1.00 ml. tuberculin syringe.

2-Mass motility score (from 1 to 5 grades).

3-Percentage of live and abnormal sperm was determined after staining with iosine and nigrosine.

4-Sperm concentration was determined by using Thomes – Zeis haemocytometer.

5-Total sperm/ejaculate  $\times 10^9$  = (ejaculate volume  $\times$  sperm concentration).

6-Total abnormal sperm/ejaculate  $\times 10^9$  = (abnormal sperm%  $\times$  total sperm/ejaculate /100).

7-Total live sperm/ejaculate  $\times 10^9$  = (live sperm%  $\times$  total sperm/ejaculate /100).

The previous characteristics were determined according to Kalamah *et al.*, (2000).

**Fertility and hatchability:**

Eggs from each treatment were collected through one week at the end of experimental period (36 wks of age), to study the fertility and hatchability% as follows:-

$$\text{Fertility \%} = (\text{Number of fertile eggs} \times 100) / \text{Total eggs set}$$

$$\text{Hatchability\% 1} = (\text{Number of hatched chicks} \times 100) / \text{Total number of eggs}$$

$$\text{Hatchability\% 2} = (\text{Number of hatched chicks} \times 100) / \text{Total number of fertile eggs}$$

**Statistical analysis:**

Data were subjected to one-way analysis of variance using **SAS (2001)**. Differences among means were detected by using Duncan's multiple range test (**Duncan, 1955**). The percentage values were transferred to percentage angle using arcsine equation before subjected to statistical analysis, and then actual means are presented. The following model was used:

$$Y_{ij} = \mu + T_i + e_{ij}$$

Where,  $Y_{ij}$  = an observation for each dependent variable;  $\mu$  = overall mean;

$T_i$  = Treatment effects ( $i = 1,2,\dots$  and 7);  $e_{ij}$  = experimental random error.

## **RESULTS AND DISCUSSION**

**Laying performance:**

Results obtained from Table (2) showed a non-significant difference among antioxidant treatment groups for final body weight and body weight gain (change in body weight) at the end of the experimental period (36wk old) as compared to the control group. On the other hand, there was a significant difference ( $P \leq 0.05$ ) among antioxidant treatment groups (except CA groups) for feed intake values compared with control group. However, the hens fed diet contains BHT or vit. E groups consumed diet significantly ( $P \leq 0.05$ ) less than control group. Moreover, FCR (g feed/g egg) and laying performance parameters (egg number, egg production % and egg mass) were significantly ( $P \leq 0.05$ ) improved in the hens fed diet containing different types and levels of antioxidant compared with the control group. In this respect, antioxidant such as vit. E act during the initiation stage, slowing the generation of free radicals (Carrillo-Dominguez *et al.*, 2012). Vit. E acts using at least two different mechanisms. Firstly, it directly scavenges the free radicals through modulates the expression of genes that are required by free radical signaling, and this free radicals damage cell membranes by inducing lipid peroxidation of polyunsaturated fatty acids in the cell membrane (Packer *et al.*, 1997). Secondly, vit. E increases the activity of antioxidant enzymes such as superoxide dismutase, glutathione peroxidase, glutathione reductase, catalase and nicotinamide adenine dinuclotide phosphate, which considered the enzymatic defense system against free radicals and protecting cells against oxidative damage, also, vit. E has known to reduce  $Fe^{++}$  induced lipid peroxidation (Vertuani *et al.*, 2004).

**Table (2): Effect of adding some antioxidants to diet containing fatty acids on the performance of Silver Montazah laying hens during the period from 24 to 36 weeks of age.**

Item	Treatments/ton diet							±SEM <sup>3</sup>
	Control <sup>1</sup> (C)	C+125g BHT <sup>2</sup>	C+250g BHT	C+100 g vit. E	C+200 g vit. E	C+5 kg citric acid	C+10 kg citric acid	
Initial body weight (g)	1373.73	1376.93	1376.63	1376.30	1375.17	1376.63	1379.30	1.946
Final body weight (g)	1517.93	1531.20	1534.43	1547.97	1565.73	1543.10	1546.83	17.311
Change in body weight (g)	144.20	154.27	157.80	171.67	190.57	166.47	167.53	16.946
Total feed intake (g/hen)	9286.00 <sup>a</sup>	9061.67 <sup>c</sup>	9110.00 <sup>bc</sup>	9163.00 <sup>bc</sup>	9163.33 <sup>bc</sup>	9193.67 <sup>ab</sup>	9170.67 <sup>abc</sup>	36.741
Feed conversion (g feed/g egg)	4.783 <sup>a</sup>	3.837 <sup>b</sup>	3.873 <sup>b</sup>	3.837 <sup>b</sup>	3.833 <sup>b</sup>	3.903 <sup>b</sup>	3.890 <sup>b</sup>	0.049
Egg number	44.33 <sup>b</sup>	53.60 <sup>a</sup>	53.59 <sup>a</sup>	54.39 <sup>a</sup>	54.42 <sup>a</sup>	53.54 <sup>a</sup>	53.78 <sup>a</sup>	0.507
Egg production (%)	52.78 <sup>b</sup>	63.81 <sup>a</sup>	63.80 <sup>a</sup>	64.74 <sup>a</sup>	64.79 <sup>a</sup>	63.74 <sup>a</sup>	64.03 <sup>a</sup>	0.603
Egg weight (g)	43.83	44.07	43.88	43.90	43.95	43.98	43.84	0.141
Egg mass (g)	1943.14 <sup>b</sup>	2362.23 <sup>a</sup>	2351.67 <sup>a</sup>	2387.55 <sup>a</sup>	2391.96 <sup>a</sup>	2354.76 <sup>a</sup>	2357.98 <sup>a</sup>	22.104

<sup>a, b, c</sup>...Means within the same row with different superscripts are significantly differ ( $P \leq 0.05$ ). Control<sup>1</sup> basal diet containing linoleic acid (n-6) and linolenic acid (n-3) at the ratio of 4:1, respectively BHT<sup>2</sup> butylated hydroxyl toluene  
<sup>3</sup> Pooled SEM

Accordingly results in the present study demonstrated that the high level of vit. E (150 mg/kg diet) resulted in significantly ( $P \leq 0.05$ ) better value of FCR in laying periods compared to either low levels or control group, and this effect reflected on 1) improvement in body weight in laying period 2) earlier age of sexual maturity with significance ( $P \leq 0.05$ ) for chickens in this group compared to low levels and control group (Alm El-Dein *et al.*, 2017). Panda *et al.* (2008) found that the high level of vit. E (125 IU/kg diet) significantly improved FCR compared to the low level (25 IU/kg diet) in White Leghorn layers. Concerning body weight, Sahin *et al.* (2002) reported that supplementation of 250 mg/kg diet vit. E significantly ( $P \leq 0.05$ ) increased body weight in laying Japanese quail as compared to 125 mg/kg diet and control group.

The highest level of vit. E significantly ( $P \leq 0.05$ ) improved egg production more than other low levels and control group. This may be because vit. E playing an important role in, 1) enhancing synthesis of egg yolk precursors (vitellogenin and very low density lipoprotein) in the liver through its action as an antioxidant which protect the liver from lipid peroxidation and damage to the cell membrane and 2) facilitating the release of the previous precursors from the liver and increasing the circulating supply of them for yolk formation (Bollengier-Lee *et al.*, 1998 and 1999 and Puthpon-Gsiriporn *et al.*, 2001). Results reported herein for egg production support the previous finding of Panda *et al.* (2008) who reported that, increasing the level of vit. E from 25 to 125 IU/kg diet significantly improved egg production.

Alm El-Dein *et al.* (2017) found that, adding different levels of vit. E to laying hen's diet increased hen productive performance significantly ( $P \leq 0.05$ ) in terms of egg number, egg production percentage and egg mass and without significant effects on body weight gain and egg weight compared to control group, although, total feed intake had comparable values (slight increase) among all experimental groups compared to control group. Consequently, the presence of anti-oxidants (vit.E) could partially interfere with oxidative protein denaturation and thereby improving its digestibility thus reflect on hen productivity (Ciftci *et al.*, 2005). Similar trends were reported by Metwally (2003); El-Mallah *et al.* (2011); Alm El-Dein *et al.* (2013)

and Younis (2014) who showed a significant increase in egg production for the group supplemented with 450 mg vit. E/kg diet as compared with control group. It may be attributed to the role of vit. E. Also, vit.E increased egg productivity by preventing liver cell damage which is important in egg yolk synthesis (Kirunda *et al.*, 2001).

Al-Harathi and Attia (2015) found that feed intake and FCR were significantly affected by CA supplementation in laying hens, the positive effect of CA on feed intake may be due to the effect of maintaining feed freshness and thus the palatability of the feed. High levels of calcium in the laying hens' diets may interfere with CA affecting pH in the gut towards alkalinity, thus reducing its effectiveness as an acidic agent (Nezhad *et al.*, 2007). In the literature, CA supplementation has been reported to preserve freshness and to decrease microbial growth and harmful bacteria (*E. Coli*), thereby maintaining feed quality during storage and in the gut and leading to better animal growth (Deepa *et al.*, 2011).

#### Egg quality:

Concerning external and internal egg quality traits, Table (3) showed that hens fed diets provided with antioxidant improved significantly ( $P \leq 0.05$ ) shell thickness as compared with the hens in the control group. However, the hens fed diet contains 125g BHT/ton diet recorded significantly ( $P \leq 0.05$ ) higher yolk (%) compared with control group. Moreover, yolk index was significantly ( $P \leq 0.05$ ) increased in the hens fed diet containing the highest level of vit. E (200g/ton diet) compared with hens fed diet containing the lowest level of vit. E (100g/ton diet). These results agreed with the findings of Al-Harathi and Attia (2015) who indicated that adding 0.2% CA to laying hens diets significantly improved eggshell quality compared to adding 0.1% CA. In the literature, CA supplementation at 0.15-0.2% improved eggshell quality. This is probably due to the positive impact of CA on phytate phosphorus utilization and thus on the release of minerals bound to the phytate molecule (Islam, 2012 and Kaya *et al.*, 2014). In the literature, organic acids was found to promote the growth of the epithelial cells of the intestine and thereby improve the absorption and metabolism of calcium (Langhout and Sus, 2005) and improve the digestion and absorption of nutrients (Vogt, 2001 and Kaya *et al.*, 2014). El-Mallah *et al.* (2011) showed that most egg quality parameters were not affected ( $P \leq 0.05$ ) by adding the vit. E at either level 0.20 or 0.40 mg/kg except, shape index which was significantly ( $P \leq 0.05$ ) decreased and shell thickness which significantly ( $P \leq 0.05$ ) improved compared to the control.

**Table (3): Effect of adding some antioxidants to diet containing fatty acids on external and internal egg quality of Silver Montazah laying hens at 36 weeks of age.**

Item	Treatments/ton diet							±SEM <sup>3</sup>
	Control <sup>1</sup> (C)	C+125g BHT <sup>2</sup>	C+250g BHT	C+100 g vit. E	C+200 g vit. E	C+5 kg citric acid	C+10 kg citric acid	
	External egg quality							
Egg length (mm)	53.78	52.12	52.24	51.78	51.56	51.64	52.54	0.897
Egg width (mm)	41.56	41.08	40.56	41.22	40.90	40.90	40.98	0.639
Shell thickness (mm)	0.332 <sup>b</sup>	0.376 <sup>a</sup>	0.378 <sup>a</sup>	0.372 <sup>a</sup>	0.370 <sup>a</sup>	0.374 <sup>a</sup>	0.372 <sup>a</sup>	0.004
Shell weight (%)	13.08	13.58	13.47	14.26	14.80	14.25	13.89	0.975
Egg shape index	77.30	78.91	77.66	77.67	79.32	77.92	78.05	1.015
	Internal egg quality							
Albumen height (mm)	6.43	7.08	6.70	7.61	6.74	6.87	7.22	0.955
Albumen (%)	57.28	53.40	54.37	53.14	53.32	52.95	54.27	1.518
Yolk height (mm)	16.91	17.12	16.79	16.02	17.05	16.62	15.95	0.450
Yolk diameter	39.18	38.84	39.20	39.64	37.44	38.66	39.18	0.804
Yolk (%)	29.63 <sup>b</sup>	33.01 <sup>a</sup>	32.16 <sup>ab</sup>	32.60 <sup>ab</sup>	31.88 <sup>ab</sup>	31.86 <sup>ab</sup>	31.54 <sup>ab</sup>	0.949
Yolk index	43.21 <sup>ab</sup>	44.15 <sup>ab</sup>	42.93 <sup>ab</sup>	40.52 <sup>b</sup>	45.52 <sup>a</sup>	43.00 <sup>ab</sup>	40.89 <sup>ab</sup>	1.443

<sup>a, b, c</sup> ...Means within the same row with different superscripts are significantly differ ( $P \leq 0.05$ ). Control<sup>1</sup> basal diet containing linoleic acid (n-6) and linolenic acid (n-3) at the ratio of 4:1, respectively BHT<sup>2</sup> butylated hydroxyl toluene

<sup>3</sup> Pooled SEM

These results confirmed those of El-Sheikh and Salama (2010) who reported that vit E improved shell thickness and haugh unit score as compared to the control but, did not affect significantly shell weight and albumen weight % as compared to control, Similar results were reported by Eng-elmann *et al.* (2001), Kirunda *et al.* (2001) and Abd-El-Galil and Abd El-Samad (2004). In this connection, the achieved

improvement in shell-thickness could be due to enhancement of calcium bioavailability by the action of supplemental vit. E. These facts confirmed the results of increased serum Ca concentration that has been established in the present study (Abdel-Fattah and Abdel-Azeem, 2007).

**Hemato-biochemical parameters:**

The results in Table (4) showed that vit. E treated laying hens had significantly ( $P \leq 0.05$ ) higher RBC's and WBC's number compared with control group. On the other hand, heterophils/ lymphocytes ratios were significantly ( $P \leq 0.05$ ) lower due to adding the highest level of CA (10 kg /ton) to laying hens diet compared with both the lowest level of CA (5 kg /ton) and control groups. While, adding different types and levels of antioxidants to laying hens diet had significantly ( $P \leq 0.05$ ) higher hemoglobin compared with control group. Belong to cell mediated immunity, vit. E has been reported to protect the cells involved in the immune response (lymphocytes, macrophages and plasma cells) against oxidative damage and enhance the function and proliferation of these cells through maintain the macrophage membrane integrity which are needed for phagocytosis (Gore and Qureshi, 1997). Thus, Latshaw (1991) indicated that high levels of vit. E greater than 10 times the required level (recommend 5 to 25 IU of vit-E/kg of diet according to (NRC, 1994) were immunostimulatory in chicks. Younis (2014) revealed a significant increase in total RBC count, Hb concentration, PCV%, heart glycogen and a significant improving in glucose, triglycerides, ALT, AST and liver glycogen concentration due to supplementation of broiler breeders diet with vit. E.

**Table (4): Effect of adding some antioxidants to diet containing fatty acids on hematological picture of Silver Montazah laying hens at 36 weeks of age.**

Item	Treatments/ton diet							±SEM <sup>3</sup>
	Control <sup>1</sup> (C)	C+125g BHT <sup>2</sup>	C+250g BHT	C+100 g vit. E	C+ 200g vit. E	C+5 kg citric acid	C+ 10kg citric acid	
Red blood cells (x 10 <sup>6</sup> /µl)	2.067 <sup>b</sup>	2.367 <sup>ab</sup>	2.407 <sup>ab</sup>	2.607 <sup>a</sup>	2.581 <sup>a</sup>	2.473 <sup>ab</sup>	2.300 <sup>ab</sup>	0.131
Hemoglobin	10.800 <sup>b</sup>	13.333 <sup>a</sup>	13.167 <sup>a</sup>	14.033 <sup>a</sup>	13.920 <sup>a</sup>	13.367 <sup>a</sup>	13.00 <sup>a</sup>	0.683
Hematocrit	35.767	35.000	39.733	36.200	35.090	39.833	33.100	2.820
White blood cells (x 10 <sup>3</sup> /µl)	5.600 <sup>b</sup>	6.767 <sup>ab</sup>	7.100 <sup>ab</sup>	8.067 <sup>a</sup>	7.940 <sup>a</sup>	6.567 <sup>ab</sup>	6.900 <sup>ab</sup>	0.528
Heterophil (%)	29.400	28.133	28.200	27.967	27.090	28.133	27.800	0.585
Lymphocyte (%)	59.467	59.733	59.833	58.967	58.310	60.533	60.900	0.782
Heterophil/ Lymphocyte ratio	0.495 <sup>a</sup>	0.471 <sup>ab</sup>	0.471 <sup>ab</sup>	0.474 <sup>ab</sup>	0.460 <sup>ab</sup>	0.495 <sup>a</sup>	0.456 <sup>b</sup>	0.010

<sup>a, b, c</sup>...Means within the same row with different superscripts are significantly differ ( $P \leq 0.05$ ). Control<sup>1</sup> basal diet containing linoleic acid (n-6) and linolenic acid (n-3) at the ratio of 4:1, respectively BHT<sup>2</sup> butylated hydroxyl toluene  
<sup>3</sup> Pooled SEM

Concerning biochemical parameters, data of some plasma blood constituent of Silver Montazah laying hens as affected by feeding diet containing varying types and levels of antioxidant are presented in (Table 5). It is clear that the effect of antioxidant supplementation was more pronounced in laying hens on plasma lipids and proteins, thus plasma total lipids, cholesterol and triglycerides concentrations were decreased significantly. On the contrary, plasma total protein and globulin were increased significantly. However, plasma level of LDL was significantly the lowest for the vit. E supplemented groups at different levels and CA supplemented group at the highest level (10 kg/ton diet), but plasma level of HDL was significantly higher for the vit. E supplemented group at higher level (200 g/ton diet) comparable to the control group. It is clearly observed that, dietary addition of antioxidants, could insignificantly affect the plasma albumin, albumin/globulin ratio and hepatic enzymes activities (AST and ALT enzymes) comparable to control group. In this respect, El-Mallah *et al.* (2011) showed that hens fed diet added with vit. E up to 0.50 mg/kg had significantly ( $P \leq 0.05$ ) increased blood serum total protein, albumin and globulin, whereas no effect ( $P \leq 0.05$ ) on serum AST, ALT, triglycerides and glutathione peroxidase was detected comparing to the control group. These results supported those of Gursu *et al.* (2003) who found that serum activities of AST and ALT were not influenced by dietary vit. E supplementation. But through its known properties as an intra-membrane antioxidant, vit. E may protect tissue membrane from lipid peroxidation caused by free radicals attack. It could therefore reduce the associated loss of integrity of function of cell membranes and associated

increased cellular permeability and play a role in alleviating the effect of heat stress in laying hens. Younis (2014) and Mobaraki *et al.* (2013) reported that the addition of vit. E affected triglyceride level in serum and the findings of (Ozcan *et al.*, 2001) showed that dietary supplementary of vit. E affect the triglycerides level of chicks.

**Table (5): Effect of adding some antioxidants to diet containing fatty acids on blood plasma proteins and lipids concentrations and liver functions of Silver Montazah laying hens at 36 weeks of age.**

Item	Treatments/ton diet							±SEM <sup>3</sup>
	Control <sup>1</sup> (C)	C+125g BHT <sup>2</sup>	C+250g BHT	C+100 g vit. E	C+ 200g vit. E	C+5 kg citric acid	C+ 10kg citric acid	
Plasma proteins (g/dl):								
Total protein	4.96 <sup>b</sup>	5.30 <sup>ab</sup>	5.48 <sup>ab</sup>	5.74 <sup>a</sup>	5.85 <sup>a</sup>	5.44 <sup>ab</sup>	5.88 <sup>a</sup>	0.215
Albumin (Al)	2.87	3.02	2.97	3.13	3.18	3.04	3.18	0.099
Globulin (Gl)	2.09 <sup>b</sup>	2.28 <sup>ab</sup>	2.51 <sup>ab</sup>	2.61 <sup>ab</sup>	2.67 <sup>a</sup>	2.40 <sup>ab</sup>	2.70 <sup>a</sup>	0.166
Al/GI Ratio	1.380	1.320	1.200	1.238	1.214	1.270	1.198	0.080
Plasma lipids (mg/dl):								
Total lipids	405.85 <sup>a</sup>	405.76 <sup>a</sup>	383.51 <sup>ab</sup>	355.92 <sup>ab</sup>	347.14 <sup>b</sup>	375.71 <sup>ab</sup>	342.12 <sup>b</sup>	16.337
Total cholesterol	173.41 <sup>a</sup>	164.30 <sup>ab</sup>	150.67 <sup>ab</sup>	118.49 <sup>b</sup>	123.69 <sup>b</sup>	144.85 <sup>ab</sup>	132.72 <sup>ab</sup>	14.560
Triglycerides	91.46 <sup>a</sup>	87.54 <sup>a</sup>	79.79 <sup>ab</sup>	68.81 <sup>b</sup>	66.97 <sup>b</sup>	77.39 <sup>ab</sup>	64.22 <sup>b</sup>	5.715
Height density lipoproteins cholesterol	69.05 <sup>b</sup>	70.96 <sup>ab</sup>	69.42 <sup>ab</sup>	82.06 <sup>ab</sup>	83.39 <sup>a</sup>	67.35 <sup>ab</sup>	80.55 <sup>ab</sup>	4.318
Low density lipoproteins cholesterol	54.68 <sup>a</sup>	48.97 <sup>ab</sup>	48.85 <sup>ab</sup>	45.51 <sup>b</sup>	44.09 <sup>b</sup>	47.64 <sup>ab</sup>	43.11 <sup>b</sup>	2.566
Liver functions (U/L):								
Aspartate amino transaminase	52.26	51.61	49.90	49.99	50.62	48.90	50.06	1.182
Alanine amino transaminase	24.76	24.21	23.80	23.97	23.00	23.14	22.77	1.778

<sup>a, b, c</sup>...Means within the same row with different superscripts are significantly differ ( $P \leq 0.05$ ). Control<sup>1</sup> basal diet containing linoleic acid (n-6) and linolenic acid (n-3) at the ratio of 4:1, respectively BHT<sup>2</sup> butylated hydroxyl toluene

<sup>3</sup> Pooled SEM

However, Abdel-Fattah *et al.* (2008) showed that the addition of 1.5 or 3% CA increased the calcium and phosphors concentrations in the blood serum. However, CA did not affect the osteocalcin, or the blood serum content of 1,25 (OH)<sub>2</sub> vit. D or Ca of chickens fed a diet containing 0, 0.25, 0.75 and 1.25% CA (Islam *et al.*, 2010). In addition, CA affected mineral utilization, but did not affect the plasma mineral contents (Nourmohammadi *et al.*, 2010). These differences in the response to CA may be due to strain differences, dietary composition, hygienic conditions and the hormonal balance responsible for maintaining Ca and P homeostasis (Fraser, 1988).

There were increase in plasma total protein and globulin concentrations due to adding 0.2% CA to laying hens diet (Chowdhury *et al.*, 2009; Das *et al.*, 2011 and Islam, 2012). Chicks fed acidifiers have a better immune response, as evidenced by increased serum globulin and an allied increase in lymphoid organ weights when compared to a control group (Abdel-Fattah *et al.*, 2008). Contradictory, El-Afifi *et al.* (2001) found a decrease in blood plasma protein with supplementation of different CA concentrations (0.0%, 0.2%, 0.4%, 0.6%, and 0.8%) to diet for broilers. In a subsequent study, El-Afifi (2003) cited that total plasma protein, albumin and globulin were not affected by CA. This contradiction in response to CA supplementation could be attributed to the CA concentration, dietary composition and hygienic conditions (Islam, 2012). However, Al-Harhi and Attia (2015) found that CA at 0.1% led to a significant increase in plasma albumin compared to 0.2% CA, a non-specific immune index compared to the other concentrations of CA. Similarly, Capcarova *et al.* (2014) found that CA did not negatively affect serum total protein but significantly increased serum albumin and decreased the albumin to globulin ratio.

#### Semen quality:

Data of the semen physical characteristics are shown in Table (6). Supplementation of some antioxidant to cock's diet at different types and levels caused a significant ( $P \leq 0.05$ ) effect on all semen physical

characteristics studied except semen pH and total abnormal sperm/ejaculate. However, adding some antioxidant to cock's diet especially BHT at the highest level (250 g/ton diet), vit. E at different levels (100 or 200 g/ton diet) and CA at the lowest level (5 kg/ton diet) recorded higher values of ejaculate volume, sperm motility, live sperm percentage, sperm concentration, total sperm per ejaculate and total live sperm

**Table (6): Effect of adding some antioxidants to diet containing fatty acids on semen quality traits of Silver Montazah cocks at 36 weeks of age.**

Item	Treatments/ton diet							±SEM <sup>3</sup>
	Control <sup>1</sup> (C)	C+125g BHT <sup>2</sup>	C+250g BHT	C+100g vit.E	C+200g vit.E	C+5 kg citric acid	C+10kg citric acid	
Ejaculate volume (ml)	0.324 <sup>b</sup>	0.342 <sup>b</sup>	0.374 <sup>ab</sup>	0.434 <sup>a</sup>	0.472 <sup>a</sup>	0.400 <sup>a</sup>	0.384 <sup>ab</sup>	0.039
Semen pH	7.40	7.38	7.81	7.62	7.80	7.43	7.57	0.233
Sperm motility (1-5)	3.00 <sup>b</sup>	3.20 <sup>ab</sup>	4.20 <sup>a</sup>	4.20 <sup>a</sup>	3.60 <sup>ab</sup>	3.80 <sup>ab</sup>	3.20 <sup>ab</sup>	0.309
Live sperm %	83.60 <sup>b</sup>	80.00 <sup>b</sup>	91.20 <sup>a</sup>	88.00 <sup>a</sup>	89.00 <sup>a</sup>	83.60 <sup>b</sup>	83.40 <sup>b</sup>	2.110
Abnormal sperm %	17.60 <sup>ab</sup>	16.80 <sup>ab</sup>	14.00 <sup>ab</sup>	14.60 <sup>ab</sup>	12.60 <sup>b</sup>	16.60 <sup>ab</sup>	18.00 <sup>a</sup>	1.539
Sperm concentration	3.280 <sup>ab</sup>	3.282 <sup>ab</sup>	3.782 <sup>a</sup>	3.808 <sup>a</sup>	3.646 <sup>a</sup>	3.582 <sup>ab</sup>	3.210 <sup>b</sup>	0.176
Total sperm/ejaculate	1.054 <sup>b</sup>	1.134 <sup>ab</sup>	1.406 <sup>ab</sup>	1.620 <sup>ab</sup>	1.724 <sup>a</sup>	1.462 <sup>ab</sup>	1.240 <sup>ab</sup>	0.196
Total live sperm/ejaculate	0.892 <sup>b</sup>	0.928 <sup>b</sup>	1.286 <sup>ab</sup>	1.410 <sup>ab</sup>	1.534 <sup>a</sup>	1.248 <sup>ab</sup>	1.034 <sup>ab</sup>	0.176
Total abnormal sperm/ejaculate	0.184	0.182	0.202	0.248	0.216	0.248	0.220	0.041

<sup>a, b, c</sup>...Means within the same row with different superscripts are significantly differ ( $P \leq 0.05$ ). Control<sup>1</sup> basal diet containing linoleic acid (n-6) and linolenic acid (n-3) at the ratio of 4:1, respectively BHT<sup>2</sup> butylated hydroxyl toluene  
<sup>3</sup> Pooled SEM

per ejaculate compared to cocks of the other groups. On the opposite, adding CA at the highest level (10 kg/ton diet) recorded the highest values of abnormal sperm percentage compared to cocks fed diet containing vit. E at the highest level (200 g/ton diet). In this respect, Abdallah *et al.* (2017) and Alm El-Dein *et al.* (2017) showed that supplementation of vit. E to cock's diet caused significant ( $P \leq 0.05$ ) effect on all semen physical characteristics studied except semen pH at 60 wks of age. Increasing sperm motility by dietary supplementation of vit. E may be due to vit. E decreases semen homocysteine which is a defective amino acid formed from demethylation of methionine and increasing homocysteine blocks intra-cellular protein-carboxyl methylation reactions results in inhibition of sperm motility (Sonmez *et al.*, 2007). Results reported herein are similar to the conclusions drawn by Lin *et al.* (2005) who found that increasing level of vit.E from 40 to 160 mg/kg diet for Taiwan native cocks at 39 week of age significantly improved semen traits, particularly sperm livability and motility. Significant ( $P \leq 0.05$ ) decrease of abnormal sperm percentage and increase sperm concentration by dietary supplementation of vit. E compared to control group may be due to the ability of vit. E as an antioxidant to resist the oxidative DNA damage and genetic alterations in the spermatozoa thus decrease abnormal sperm percentage (Bagachi *et al.*, 1997). Also, Alm El-Dein *et al.* (2013) found that the highest level of vit. E (150 mg /kg diet) improved the semen physical characteristics compared to the other low levels, according to the role of vit. E which, 1) reduces lipid peroxidation in seminal plasma and maintains an adequate viability of sperm to complete the fertilization process (Eid *et al.*, 2006), 2) decreases semen homocysteine which is a defective amino acid formed from demethylation of methionine. The increased concentration of homocysteine blocks intra-cellular protein-carboxyl methylation reactions results in inhibition of sperm motility (Wallock *et al.*, 2001 and Sonmez *et al.*, 2007). Results obtained agreed with Lin *et al.* (2005) who demonstrated in 39 week old Taiwan native cockerels that, elevating the addition level of vit. E from 40 to 160 mg/kg diet plays a very significant role in improving semen traits, particularly sperm livability and motility,

On the other hand, Triques *et al.* (2016) showed that no effects of diet supplementation with the antioxidant blend were observed on semen volume and sperm motility, as well as vigor and concentration of roosters during the post-peak production phase. The observed percentages of normal sperm for both the control and antioxidant supplementation treatments were close to those reported by Łukaszewicz *et al.* (2008).

**Fertility and hatchability:**

Significant effects of antioxidant treatments on fertility and hatchability percentages are shown in Table (7). Fertility percentage was significantly ( $P \leq 0.05$ ) higher in vit. E treated group at the highest level (200 g/ton diet) than the control group. An increase of fertile eggs in antioxidant treated groups could be due to increase of ejaculate volume, sperm motility, live sperm%, sperm concentration, total sperm/ejaculate and total live sperm/ejaculate compared with those in the control group. Also, hatchability percentage (hatchability of total eggs) was significantly ( $P \leq 0.05$ ) higher in vit. E at the different levels (100 or 200 g/ton diet) groups compared to the control group. Moreover, hatchability percentage (hatchability of fertile eggs) was significantly ( $P \leq 0.05$ ) higher in vit. E at the different levels (100 or 200 g/ton diet) and CA at the

**Table (7): Effect of adding some antioxidants to diet containing fatty acids on fertility and hatchability percentages of Silver Montazah laying hens at 36 weeks of age.**

Item	Treatments/ton diet							±SEM <sup>3</sup>
	Control <sup>1</sup> (C)	C+125g BHT <sup>2</sup>	C+250g BHT	C+100 g vit. E	C+200g vit E	C+5 kg citric acid	C+10kg citric acid	
Fertility (%)	84.55 <sup>b</sup>	86.13 <sup>ab</sup>	86.24 <sup>ab</sup>	89.56 <sup>ab</sup>	91.50 <sup>a</sup>	87.74 <sup>ab</sup>	88.39 <sup>ab</sup>	1.789
Hatchability of all eggs (%)	77.99 <sup>b</sup>	78.08 <sup>b</sup>	80.66 <sup>ab</sup>	81.83 <sup>a</sup>	82.53 <sup>a</sup>	80.87 <sup>ab</sup>	81.67 <sup>ab</sup>	2.217
Hatchability of fertile eggs (%)	83.37 <sup>b</sup>	83.33 <sup>b</sup>	87.00 <sup>ab</sup>	90.33 <sup>a</sup>	90.67 <sup>a</sup>	88.00 <sup>ab</sup>	90.00 <sup>a</sup>	2.548

<sup>a, b, c</sup>...Means within the same row with different superscripts are significantly differ ( $P \leq 0.05$ ). Control<sup>1</sup> basal diet containing

linoleic acid (n-6) and linolenic acid (n-3) at the ratio of 4:1, respectively BHT<sup>2</sup> butylated hydroxyl toluene <sup>3</sup> Pooled SEM

highest level (10 kg/ton diet) groups compared to the control group. Such increase may depend on egg shell thickness improvement in most antioxidant treated groups compared with the control group. In this respect, chicken spermatozoa are unique in their structure and chemical composition. The most important feature of lipid composition of the avian semen is the extremely high proportions of long-chain polyunsaturated fatty acids (PUFAs) in the phospholipid fraction of spermatozoa. On the one hand, the high PUFAs proportion is a necessity in order to maintain specific membrane properties (fluidity, flexibility, etc). On the other hand, spermatozoa became very susceptible to lipid peroxidation and, therefore, the antioxidant defence is considered to be a key element in maintaining semen quality. Vitamin E is located in spermatozoa and provides an antioxidant protection, especially in stress conditions of in vitro semen manipulation, including dilution, storage and deep freezing of spermatozoa (Surai, 2002). Furthermore, it was shown that vit. E provides additional protection in the case of fatty acid manipulation of the semen (Surai *et al.*, 2000; Zanini *et al.*, 2003 and Cerolini *et al.*, 2005). However, in some studies the vit. E dose-response in cockerels was shown to be non-linear (Lin *et al.*, 2005).

Alm El-Dein *et al.* (2017) found that fertility and hatchability percentage of both total and fertile eggs were increased by dietary vit. E compared to control group may be due to vit. E increase sperm motility as mentioned above in Table (6) and the motility is essential for sperm to traverse the vagina and reach the sperm storage tubules which is important for increasing fertility moreover, reduce lipid peroxidation in seminal plasma through reducing dexamethasone thus maintains an adequate viability of sperm which helping to complete the fertilization process (Eid *et al.*, 2006). Also, vit. E through improving antioxidant status resulted in improving hatchability because oxidative metabolism increases during embryo development at hatchability process especially in the last few days before hatch, due to normal respiration related to embryo growth results in increasing the production of free radicals that doing lipid peroxidation leading to tissue damage and hatchability decline (Freeman and Vince, 1974). This concept is confirmed by Benzie (2003) who said that although oxygen is vital for most organisms but, simultaneously, considered damage key for biological sites and this damage is met by antioxidants substances. Also, Abdallah *et al.* (2017) found that dietary vit. E significantly ( $P \leq 0.05$ ) increased the fertility and hatchability percentages from total eggs and/or hatchability from fertile eggs compared with those of the control, the concentration of vit. E (200 mg/kg diet) realized the best significant result of hatchability from total eggs and/or hatchability from fertile eggs compared with other groups. Also, Cerolini *et al.* (2005) reported that dietary supplementation of vit. E was associated with increased fertilizing ability of cockerels.

The hatching process is considered to be a time of oxidative stress. Therefore, improved antioxidant defense during embryonic development potentially could increase hatchability. It was shown that vit. E can be transferred from the diet to the egg and consequently to the developing embryo (Surai, 2002). Increased vit. E concentration in the chicken embryonic tissues was associated with decreased tissue susceptibility to lipid peroxidation (Surai *et al.*, 1999).

## CONCLUSION

Dietary supplemental of some antioxidants to diet containing fatty acids is a practical application especially by 100 and 200 g vit. E or 10 kg CA /ton diet to Silver Montazah chickens strain during the laying period and had beneficial effects on productive, reproductive, physiological and hematological parameters and immune status.

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## تأثير إضافة بعض مضادات الأكسدة لعليقة تحتوى على الأحماض الدهنية على الأداء الإنتاجي وبعض القياسات الفسيولوجية لدجاج سلالة المنتزه الفضي. 2- خلال مرحلة إنتاج البيض

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أجري هذا البحث لمدة 12 أسبوع لدراسة تأثير إضافة بعض مضادات الاكسدة لعلائق دجاج سلالة المنتزه الفضي المحلية المحتوية على الأحماض الدهنية على الأداء الإنتاجي والتناسلي وبعض الصفات الفسيولوجية. أستخدم في هذه الدراسة عدد 231 طائر (210 دجاجة + 21 ديك) عند عمر 24 أسبوع وحتى 36 أسبوع من العمر من سلالة المنتزه الفضي. حيث قسمت الطيور إلى 7 مجموعات متساوية (30 دجاجة + 3 ديوك/مجموعة) وكل مجموعة 3 مكررات (10 دجاجات + 1 ديك/مكرر)، حيث كان متوسط الوزن متماثل تقريبا في كل المجموعات. غذيت المجموعة الاولى على عليقة اساسية دون أى إضافات (كنترول) وتحتوى على كل من الأوميغا 6، الأوميغا 3 بنسبة 4:1. غذيت المجموعتين الثانية والثالثة على عليقة اساسية مضاف اليها 125 و 250 جم (BHT)/طن علف. المجموعتين الرابعة والخامسة غذيت على عليقة اساسية مضاف اليها 100 و 200 جم فيتامين E/طن علف. المجموعتين السادسة والسابعة غذيت على عليقة اساسية مضاف اليها 5 و 10 كجم حمض الستريك /طن علف، بالترتيب.

تشير النتائج الى انه بإضافة مضادات الاكسدة زاد معنويا كلا من عدد ووزن وكثلة البيض كما تحسن معنويا كلا من معدل التحويل الغذائي وسمك قشرة البيض مقارنة بمجموعة الكنترول. علاوة على ذلك حدثت زيادة في تركيز الهيموجلوبين، أعداد كل من كرات الدم الحمراء والبيضاء وكذلك تركيز البروتين الكلى، الجلوبيولين والكوليستيرول مرتفع الكثافة كما ازداد معنويا كل من حجم الفذفة، حركة الحيوانات المنوية، النسبة المئوية للحيوانات المنوية الحية، تركيز الحيوانات المنوية، العدد الكلى لكلا من الحيوانات المنوية والحيوانات المنوية الحية في الفذفة الواحدة وكذلك النسبة المئوية لكلا من الخصوبة والفقس للمجموعات المغزاة على عليقة تحتوى على مضادات الاكسدة مقارنة بمجموعة الكنترول. على العكس من ذلك إنخفضت معنويا كل من نسبة الهيتروفيل الى الليمفوسيتيس وتركيز الدهون الكلية، الكوليستيرول الكلى، الجليسيريدات الثلاثية والكوليستيرول منخفض الكثافة بإضافة مضادات الاكسدة. بينما انخفضت النسبة المئوية للحيوانات المنوية الشاذة لجميع المعاملات مقارنة بالكنترول.

نستخلص من هذه الدراسة أن إضافة فيتامين E بمعدل 100، 200جم/طن علف أو حامض الستريك بمعدل 10 كجم/طن علف كمضادات أكسدة الى عليقة دجاج سلالة المنتزه الفضي البيضاء المحتوية على الأحماض الدهنية هو تطبيق عملي يوصي به حيث أدى إلى تحسين الأداء الإنتاجي والتناسلي كما كان لها تأثيرات مفيدة على بعض المقاييس الفسيولوجية والهيماتولوجية والحالة المناعية.