



## IMPACT OF SALINITY SEED SPROUT CHARACTERIZATION OF FIVE FABA BEAN (*Vicia faba* L.) VARIETIES

[180]

Ali\* M.A.A., Abdallah M.M.F., Nashwa A. Abo El-Azam  
and Abou El-Yazeid A.

Horticulture Dept., Fac. of Agric., Ain Shams Univ., P.O. Box 68, Hadayek Shoubra,  
11241 Cairo, Egypt

\*Corresponding author: [mahmoud\\_adel489@agr.asu.edu.eg](mailto:mahmoud_adel489@agr.asu.edu.eg)

Received 25 July, 2019

Accepted 12 November, 2019

### ABSTRACT

One of the unique properties of earlier stages of plant growth is germination. Germination is morphological expression of various metabolic activities in seeds. Plants exposure to the abiotic stresses for instance salinity that influence physiological processes, anatomy, developmental systems and plant development. Agriculture production under arid and semi-arid zone is low owing to many factors for example accumulation of salts in soils and water, total of land area under salinity is about 953 million ha. The aim of this research was to evaluate the germination characters of five faba bean (*Vicia faba* L.) varieties [Nubaria (1), Giza (843), Sakha (1), Sakha (3) and, Sakha (4)] were using saline solution containing (0, 1000 and 2000 ppm NaCl), were utilized to examine their water uptake, germination percentage, germination index, relative salt injury rate, radicle length, sprout fresh and dry weight, weight losses and seed volume using saline solution. Results indicated that the higher concentrations of salt have a negative impact on water uptake, germination percentage, radical length, sprout fresh and dry weight, seed volume, and relative salt injury rate. At all salinity concentrations, all varieties showed different degrees of salt tolerance. Sakha (3), (4) and Giza (843) had a better salt tolerance index than the others. Nubaria (1) and Sakha (1). Nubaria (1), Giza (843) had the highest percentage and index of germination, while Sakha (1) had the shorter radical length and susceptible to salt injury rate. Moreover, Giza (843) and Sakha (3) genotypes can be considered as tolerant to salt stress compared to the other ones. These genotypes also could be used in the breeding program for enhanc-

ing faba bean cultivation in newly reclaimed lands. Data suggest a test of seed germination may be useful to use in developing new lines of faba bean to grow in saline soils.

**Keywords:** Faba bean seed germination, Water uptake, Relative salt injury rate, Salt tolerance index, Sprout character, Salinity stress

### INTRODUCTION

Faba bean (*Vicia faba* L.) is a remarkable crop belonged to family Fabaceae, which contain crops such as soybean, chickpeas, peas, and beans. Faba bean is prevalent legume diet with significant yield capacity and high nutritional prominence for human and animals. Egypt is ranked seventh productions of faba bean in the world after China, Ethiopia, Australia, United Kingdom, Germany, France (FAO, 2017). Faba bean is one of important and unique legume crop planted in Egypt due to the protein, carbohydrates, dietary fiber and secondary metabolites such as phenolic and antioxidant value (Erba et al 2019; Hossain and Mortuza, 2006). An extensive problem in agriculture is salinity, as a major part of the abiotic stresses in arid and semi-arid zone and it affects the agriculture world land area (Kumawat et al 2017).

Seed germination is the decisive period in the life stages of plants. Germination processes include those steps that begin with the absorption of water by dry seed and end with the emergence of radical (Copeland and McDonald, 2001). Water imbibition with a seed is tri-phasic; phase 1 rapid initial imbibed; phase 2 is lull phase and then phase 3 an increment of water imbibition, but, only

while germination happens (Ghazi, 1998; Manz, 2005 and Schopfer & Plachy, 1984). Thus, salinity tolerance through sprouting is crucial for promotion of growing plants and under salt stress (Khan et al 2000). Despite, the sprouting importance for growing in saline soils, more is known of in stages of reproductive and vegetative about salt tolerance mechanism than during the sprouting stage (Ibrahim et al 2019 and Munns & Tester, 2008). Since excess chloride and sodium ions imbalance has a deleterious effect at all stages of plant growth (Begum et al 2013; Zhang et al 2010), therefore tolerance metabolism to Na<sup>+</sup> and Cl<sup>-</sup> is important during sprouting than later stages of plant growth, owing to limited carbohydrates available in seed (Zhang et al 2010).

Salinity reduces the proficiency of plants to absorption water leading to a decrease in plant growth also metabolic changes (Kader and Jutzi, 2002). Experiment done on six wheat varieties under different salinity concentrations, indicated that water uptake decreased by increasing salinity concentration (Ibrahim et al 2016). Hence, salinity may reduce crop production by uncomfortable water and nutritional balance of the plant (Begum et al 2013; Santos et al 2018). Solution imbibe by root system restricted at high osmotic potential and toxicity effect due to an accumulation of sodium and chloride (Na-Cl) ions (Al-Karaki, 1997; Munns, 2002). Salt stress affects seed germination percentage, rate and seedling growth. Study on Sugar beet (*Beta vulgaris*), cabbage (*Brassica oleracea capitata*), amaranth (*Amaranthus paniculatus*) and pak-choi (*Brassica campestris*) seed using salt solution reducing to seed germination percentage rate and percentage also reducing seedling growth (root and shoot lengths and fresh root and shoot weights) (Jamil et al 2006). Moreover, three cultivars of pea (*Pisum sativum* L.) were investigate under NaCl solutions and the result was decreasing germination and seedling growth in all cultivars (Okçu et al 2005). In spinach experiment, salinity negatively reducing absorption efficiency, seed vigor, hypocotyl and root length (Ibrahim et al 2019). However, Kuntze (*Limonium sinense*), sieb (*Glycine soja*) and Stapf (*Sorghum sudanense*) were investigated under various concentrations of salinity on their germination (percentage, energy, index and rate), relative of salt-injury rate, length of radicle and hypocotyl. *G. soja* was germinating adeptly at concentration < 200 mmol/ L, 50 mmol /L salt level is superior to *S. sudanense*. Seeds of *L. sinense* and *S. sudanense* may germinate at high concentration 400 mmol/L,

radicle hypocotyl ratio for last two plants were increment (Li, 2008). Other research, study the effects of various level of NaCl (200, 400, 600 and 800 mM) on germination of four plants belong to family Brassicaceae (*Sinapis alba* and *Brassica oleracea*) and Solanaceae (*Capsicum annum* and *Solanum lycopersicum*) found all the plants only germinated in the lowest level of NaCl (200 mM) (Bojović et al 2010). Moreover, in ryegrass mean germination time elongated with rising salinity and germination rate decreased. Higher NaCl concentration lead to a reduction notable of root and shoot length and decreased in there dry and fresh weight also reduce the leaf number (Ilker, 2011).

The enormous genetic variance was found in faba been in terms of seed size and structure in addition to tolerance to several environmental stresses (Duc et al 2010; Hendawey and Younes, 2013). Few reports have been done on seed imbibition and several characteristics of faba been varieties related to stress tolerance. This study was conducted to test the germination under salt stress can distinguish faba been varieties with good salinity tolerance during a crucial germination on five faba bean Egyptian varieties. Therefore, the purpose of the instant study was 1) Appraise the impression of salt stress on various varieties of faba bean 2) Screen out immeasurable salinity resistant faba bean 3) Study the impact of salinity on sprout production of faba bean and to assess the various morphological changes associated with the sprouts under different salinity gradient.

## MATERIALS AND METHODS

Two experiments have been done in Seed Laboratory, Horticulture Department, Faculty of Agriculture, Ain Shams Univ., Cairo, Egypt.

### Materials

Faba been (*Vicia faba* L.) of five varieties, Nubaria (1), Giza (843), Sakha (1), Sakha (3) and, Sakha (4) were obtained from Agriculture Research Center, Giza, Egypt. Na Cl was obtained from El-Gomhoria Company, Cairo, Egypt were used in this experiment.

### First experimental design

The design of the experiment was a factorial complete randomized design with two factors (five faba bean varieties and three salt concentrations) with three replicates for each treatment. The NaCl

solutions concentration were 0, 1000, 2000 ppm. 100 Seed of each varieties were soaked in a glass jar method for imbibition up to 18 h. Soaked water was removed and seeds were sprouting for 2 days using method reported by (Abdallah, 2008).

### Second experimental design

The design of the experiment was a factorial complete randomized design with three factors (five faba bean varieties and three salt concentrations as the first experiment) and the third factor is three soaking period (0 to 9 h, 9 to 18 h and 0 to 18 h) with three replicates for each treatment.

### Water uptake

Imbided water was measuring by using three methods. First, was get an initial volume (400 ml) of water in each jar before soaking then after 9-hours when changing the soaked solution take up the water only and measuring the residues of soaked water and deduct from initial volume as well as at 18hour similar process. Second water uptake based on weight, seeds in each treatment jar were weighted before soaking (initial weight) and after 18-hour soaking (final imbibition seed weight). Thirdly water uptake based on volume, seeds in each treatment jar were volume measured before soaking (initial volume) and after 9 and 18-hour soaking (final imbibition seed volume). For each measurement of seed imbided water, seeds were accurately removed, drained, quickly dry up by absorbent paper, weighted, and back again into the jar. Imbided water determined before (Baskin et al 2006; Orozco-Segovia et al 2007; Silva et al 2018) was calculated by following equation:

$$\text{Water uptake by weight(\%)} = \frac{\text{Final weight} - \text{Initial weight}}{\text{Initial weight}} \times 100$$

$$\text{Water uptake by volume(\%)} = \frac{\text{Final volume} - \text{Initial volume}}{\text{Initial volume}} \times 100$$

### Germination Percentage

Germination percentage was registered for a total of 2 days, and was calculated by the next formulation:

$$\text{Germination percentage(\%)} = \frac{\text{Number of germinated seeds}}{\text{Number of total seed}} \times 100$$

### Germination index

Germination index was calculated as ratio germination percentage of each treatment to germination percentage of control and calculated by the following equation according to (Kandil et al 2012):

$$\text{Germination Index (\%)} = \frac{\text{Germination \% of each treatment}}{\text{Germination \% of control}} \times 100$$

### Salt Tolerance Index (STI)

Salt tolerance index was calculated as the ratio of the dry plant weight to specific NaCl concentrations to the dry plant weight of the control and calculated by the next formula according to (Carpici et al 2009; Ibrahim et al 2016; Khatun et al 2013):

$$\text{Salt Tolerance Index (\%)} = \frac{\text{Total dry weight under NaCl concentration}}{\text{Total dry weight under control}} \times 100$$

### Relative salt-injury rate

Relative salt-injury rate was calculated as subtract between germination percentage of control and germination percentage in NaCl concentration of each treatment to germination percentage of control and calculated by the next equation (Li, 2008):

$$\text{Relative salt-injury rate} = \frac{\text{Germination \% of control} - \text{Germination \% in NaCl}}{\text{Germination \% of control}}$$

### Morphological characteristics

Seed volume: was measured by water displacement method by following steps; first, fill a beaker with water then registered the baseline initial measurement. Second, submerge seeds inside beaker and registered final measurement. Then, calculated as deduct the initial volume from the final volume (Siswantoro et al 2013) 100 seed.

Sample of 10 sprouts from each replicate were collected for measuring the following character:

Radical length: the radical length measured from the seed to the tip of the radical by centimeters (cm).

Sprout fresh weight: germinated seeds were measured by gram (g).

Sprout dry weight: germinated seeds recorded by gram (g) after oven drying 60 ° C for 72 hours.

### Statistical analysis

The data were analyzed by analysis of variance (ANOVA) using completely randomized design (Two way analysis of variance for the first experiment and three way analysis for the second experiment). The least significant difference (LSD) at 0.05 levels according to the method described by **Snedecor and Cochran, (1980)**.

## RESULTS

### Germination Percentage

At three NaCl concentrations, variety Nubaria (1) recorded the highest germination percentage (75.11%) > Giza (843) (73.78%), whereas Sakha (1) had the lowest percentage (63.56%). Nubaria (1) and Giza (843) showed higher tolerance to salinity stress more than Sakha genotypes in terms percentage of germination (**Table 1**). With the increase of salt levels, germination percentage was remarkably decreased. Concerning salinity x variety interaction Nubaria (1) reached the higher germination percentage (80%) at NaCl 1000ppm while, Giza (843) had the highest germination percentage of (68%) at the 2000 ppm NaCl concentration (**Table 1**).

### Germination Index %

At different salinity levels, Nubaria (1) variety had the higher germination index (89.42%) > Giza (843) (86.53%), whereas Sakha (1) had the lowest index (79.49%). Nubaria (1) had higher salt stress tolerance than other varieties using germination index. With the increase concentration of NaCl, germination index was notably decreased. The germination index % recorded (100%) in all varieties using distilled water while, Nubaria (1) recorded the higher germination index %when using NaCl at 1000 ppm (95.24%). At the 2000 ppm NaCl concentration, Giza (843) had the highest germination index of (79.8%). However, Sakha (1) variety recorded the lowest germination index (65.84%) when using saline water at 2000 ppm NaCl for sprouting (**Table 1**).

### Sprout growth Characteristics

Data in **Table (1)** showed also that the differences among varieties x salinity concentrations were significant where increasing salinity concentrations, the length of radical was decreased from 1.71cm when using distilled water to 1.14 cm using 2000 ppm NaCl concentration. Moreover, the variety Sakha (1) recorded the lowest radical length. The interaction showed.

### Sprout fresh and dry weight

Sprout fresh weight was decreased with increasing salinity concentration. While, sprout dry weight did not affected significantly with different salinity concentrations. Nubaria (1) recorded the highest sprout fresh and dry weight while, Giza (843) recorded the lowest sprout fresh and dry weight. Regarding to variety x salinity interaction, data in (**Table 1**) recorded the highest sprout fresh and dry weight of Nubaria (1) using distilled water while, the lowest sprout fresh and dry weight were recorded using Giza (843) combined with 2000 ppm NaCl concentration.

### Sprout Weight losses %

The weight losses indicated significant differences among varieties. Giza (843) had the highest losses (20.66%) followed by Nubaria (1) (16.59%) whereas Sakha (1) had the lowest losses of (5.29%). Salinity concentrations showed significant increase of sprout weight losses% by increasing the salinity concentrations. The interaction effect showed that sprout weight losses increased with increasing NaCl concentration, combined with Giza (843) and Nubaria (1) varieties (**Table 1**).

### Imbibed seed weight

Seed soaking weight for 18 h increased 100 seed weight from 96.86 to 201.58 (more than two-fold). The imbibed seeds weight showed significant differences among varieties, salinity concentrations, soaking time and their interactions (**Table 2**). Data in (**Table 2**) recorded the heaviest imbibed seeds weight of Nubaria (1) followed by Sakha (4) & (3) while the lower weight recorded by Giza (843) and Sakha (1). Using saline water for soaking seeds reduced imbibed seed weight comparing with soaking in distilled water.

**Table 1.** Effect of NaCl concentration on 2 days old five faba bean sprout characters that, the longest radical length was 1.98 cm recorded by Saka 3 using distilled water while the shortest was 0.65 cm recorded by Sakha (1) combined with 2000 ppm NaCl concentration (Table 1)

Salinity (NaCl ppm)	Variety	Germination %	Germination Index %	Radical length (cm)	Sprout fresh weight g 48h	Sprout dry weight g 48h	Weight losses%
Distilled water	Nubaria (1)	84 a	100 a	1.81 a	3.31 a	1.26 a	7.35 j
	Giza (843)	85.33 a	100 a	1.74 ab	1.89 fg	0.71 g	19.27 e
	Sakha (1)	80 ab	100 a	1.38 bcd	2.02 efg	0.81 fg	0.89 j
	Sakha (3)	82.67 a	100 a	1.98 a	2.3 cd	0.88 cd	7.88 i
	Sakha (4)	84 a	100 a	1.65 abc	2.35 cd	0.91 c	9.83 h
	<b>Mean</b>	<b>83.2 A</b>	<b>100 A</b>	<b>1.71 A</b>	<b>2.37 A</b>	<b>0.92 A</b>	<b>9.04 B</b>
1000 ppm	Nubaria (1)	80 ab	95.24 a	1.68 abc	2.83 g	1.08 b	20.79 c
	Giza (843)	68 abcde	79.8 bc	1.64 abc	1.84 g	0.7 g	20 d
	Sakha (1)	65.33 cde	81.64 bc	0.95 ef	2.05 dfg	0.78 efg	5.34 l
	Sakha (3)	77.33 abc	93.56 ab	1.59 abc	2.24 cde	0.86 cde	10.81 g
	Sakha (4)	74.67 abcd	89.2 ab	1.18 de	2.29 cd	0.91 c	9.76 h
	<b>Mean</b>	<b>73.07 B</b>	<b>87.89 B</b>	<b>1.41 B</b>	<b>2.25 B</b>	<b>0.87 A</b>	<b>13.34 A</b>
2000 ppm	Nubaria (1)	61.33 de	73.01 cd	1.3 cde	2.78 b	1.07 b	21.63 b
	Giza (843)	68 bcde	79.8 bc	1.67 abc	1.81 g	0.68 g	22.7 a
	Sakha (1)	45.33 f	56.84 e	0.65 f	2.02 efg	0.74 def	9.65 m
	Sakha (3)	54.67 ef	65.74 de	1.14 de	2.39 c	0.9 cd	5.82 k
	Sakha (4)	58.67 e	69.42 cde	0.91 ef	2.15 cdef	0.86 cde	15.23 f
	<b>Mean</b>	<b>57.6 C</b>	<b>68.96 C</b>	<b>1.14 C</b>	<b>2.23 B</b>	<b>0.85 A</b>	<b>15 A</b>
Average	Nubaria (1)	75.11 A	89.42 A	1.6 A	2.97 A	1.13 A	16.59 A
	Giza (843)	73.78 A	86.53 AB	1.69 A	1.85 D	0.7 D	20.66 A
	Sakha (1)	63.56 B	79.49 B	0.99 C	2.03 C	0.78 C	5.29 C
	Sakha (3)	71.56 A	86.43 AB	1.57 A	2.31 B	0.88 B	8.17 B
	Sakha (4)	72.44 A	86.21 AB	1.25 B	2.26 B	0.9 B	11.6 B

Regarding the varieties and salinity interaction, data showed that the highest imbibed seeds weight recorded by Nubaria (1) combined with distilled water whereas, the lowest imbibed seeds weight was recorded by Giza (843) with the highest salinity concentration (2000 ppm NaCl) (**Table 2**).

#### Water Uptake%

Water uptake based on weight (g/100seed) or on volume (ml/100 seed) was decreased by increasing the salinity concentration. While, varieties showed no significant differences regarding water uptake % based on weight (g/100 seed) while Sakha (4) recorded the highest water uptake based on volume (ml/100 seed). Regarding the interaction between varieties and salinity, data recorded the highest water uptake by weight recorded by Giza (843) combined with 1000 ppm NaCl concentration. The lowest water uptake by weight was recorded by Sakha (1) using 2000 ppm NaCl concentration. Also, water uptake% based on 100 seed volume interaction between varieties and salt concentrations at 18 h, Sakha (4) recorded the highest water uptake% (140.43%) using distilled water while, in Sakha (3) using 1000 ppm NaCl concentration recorded the lowest water uptake by weight (98.33%) (**Table 2**).

#### Relative salt injury rate

Data of salt relative injury rate presented in **Table (2)** Data illustrated significant differences among varieties, salinity and their interaction. The salt injury rate increased by increasing of NaCl concentration. Nubaria (1) had better tolerance to salt stress than Sakha (1) variety in terms of salt injury. The Relative salt injury rate recorded no damage in all varieties using distilled water while, Sakha varieties recorded the highest injury when using 2000 ppm NaCl concentration data was more pronounced with Sakha (1) (**Table 2**).

#### Salt Tolerance Index (STI).

Concerning salt tolerance index at germination stage, data indicated significant differences among varieties. Sakha (3) recorded the highest salt tolerance index while Sakha (1) recorded the lowest. On the other hand, data showed no significant differences regarding salinity concentrations. The index % in soaking x variety recorded (100%) in all varieties using distilled water while, Sakha (4) pre-

sented better salt stress tolerance than other varieties at 1000 ppm NaCl and Sakha (3) at 2000 ppm NaCl concentration (**Table 2**).

#### Seed imbibed water

Water imbibition during soaking period was significantly affected by variety, salinity and soaking period and their interactions (**Table 3**). Nubaria (1) recorded the highest water imbibition amount of water (114.07 ml) while, the lowest water imbibition was recorded by Sakha (1). Water absorption of seeds by decreased by increasing NaCl concentration. Concerning soaking period, data showed that the highest water imbibition recorded within 18hour soaking, moreover the absorption of water in the first 9-hour soaking (92.53 ml) was doubled than the absorption in the second 9-hour soaking (46.68 ml). Concerning varieties and soaking period interaction, data showed that the highest water imbibition recorded by Nubaria (1) (171.1 ml) within 18 hours soaking whereas, the lowest water imbibition was recorded by Sakha (1) (122.14 ml) and Giza (843) (123.45 ml). However, Nubaria (1) recorded about 61.6% of water absorption within the beginning 9 hour and the other38.4% was absorbed in the last 9 hours of soaking. On the other hand, the lowest water imbibition recorded by Sakha (1) variety (74.2% of water absorption within the first 9 hour of soaking) and the rest 25.8% of absorbed water was recorded in the second 9 hour of soaking. Concerning varieties, salinity and soaking period interaction, data in (**Table 3**) showed that the highest water imbibition recorded by Nubaria (1) (189.91 ml) in total 18 hour soaking and the lowest water imbibition recorded by Sakha (1) (28 ml)within second 9 hour soaking using distilled water for soaking.

#### Seed volume (cm<sup>3</sup>)

The seed volume (cm<sup>3</sup>) of 100 seeds decreased by increasing salinity concentration. The seed volume decreased from (145.67 cm<sup>3</sup>) to (132.26 cm<sup>3</sup>). Regarding the effect of varieties, data showed that there were significant differences among varieties concerning seed volume (cm<sup>3</sup>) of 100 seeds. Nubaria (1) recorded the highest volume (181.73 cm<sup>3</sup>) while, the lowest volume recorded (112.97 cm<sup>3</sup>) by Giza (843). Regarding soaking period, data showed that the seed volume (cm<sup>3</sup>) of 100 seeds increased by soaking seeds for 9 hour by about 81.2% while soaking for 18 hours increased seed volume by about 19.6% than the volume of seeds soaked for 9 hours.

**Table 2.** Effect of 100 seed soaking for 18h in saline water on weight, water uptake%, salt injury and tolerance of five faba bean varieties

Salinity (NaCl ppm)	Variety	100 seed Weight (g)			Water uptake% by weight (g/100 seed)	Water uptake% by volume (ml/ 100 seed)	Relative salt injury rate	Salt tolerance index %
		Zero time (row seed)	Total 18 hour (imbibed seed)	Mean				
Distilled water	Nubaria (1)	134.93 g	290.96 a	212.95 A	115.67 a	119.84 abcde	0.0 f	100 ab
	Giza (843)	80.27 <sup>hi</sup>	167.63 ef	123.95 DE	109.17 abc	102.8 de	0.0 f	100 ab
	Sakha (1)	82.93 hi	178.13 ef	130.53 DE	114.86 ab	133.36 abcde	0.0 f	100 ab
	Sakha (3)	96.8 hi	202.77 c	149.79 C	109.64 abc	125.82 abcd	0.0 f	100 ab
	Sakha (4)	100.67 h	206.13 c	153.4 C	104.72 abc	140.43 a	0.0 f	100 ab
	<b>Mean</b>	<b>99.12 C</b>	<b>209.13 A</b>	<b>154.12 A</b>	<b>110.81 A</b>	<b>124.45 A</b>	<b>0.0 C</b>	<b>100 A</b>
1000 ppm	Nubaria (1)	122.13 g	249.94 b	186.04 B	104.74 abc	102.23 abcde	0.05 ef	85.09 c
	Giza (843)	76.8 i	166.12 ef	121.46 DE	116.21 a	117.86 abcde	0.2 cde	99.73 ab
	Sakha (1)	84.53 hi	181.63 de	133.08 DE	114.86 ab	131.46 ab	0.18 cd	96.30 ab
	Sakha (3)	95.47 hi	202.82 c	149.15 C	112.44 abc	98.33 e	0.07 def	96.95 ab
	Sakha (4)	100.27 h	204.31 c	152.29 C	103.93 abc	126.67 abc	0.11 def	100.26 ab
	<b>Mean</b>	<b>95.84 C</b>	<b>200.97 B</b>	<b>148.4 B</b>	<b>110.44 A</b>	<b>116.04 B</b>	<b>0.12 B</b>	<b>97.42 A</b>
2000 ppm	Nubaria (1)	120.8 g	246.5 b	183.65 B	104.24 abc	116.79 bcde	0.27 bc	98.98 ab
	Giza (843)	77.6 i	158.65 f	118.13 E	104.27 abc	107.16 cde	0.2 cd	96.65 ab
	Sakha (1)	85.33 hi	170.81 ef	128.07 DE	99.78 c	102.02 e	0.43 a	91.36 bc
	Sakha (3)	96 hi	199.67 cd	147.84 C	107.97 abc	106.83 cde	0.34 ab	105.87 a
	Sakha (4)	98.4 h	197.57 cd	147.98 C	100.61 bc	127.82 abc	0.31 abc	93.99 bc
	<b>Mean</b>	<b>95.63 C</b>	<b>194.64 B</b>	<b>145.13 B</b>	<b>103.37 B</b>	<b>112.13 B</b>	<b>0.31 A</b>	<b>100.08 A</b>
Average	Nubaria (1)	125.96 E	262.47 A	194.21 A	108.22 A	112.95 BC	0.11 B	94.69 B
	Giza (843)	78.22 G	164.14 D	121.18 D	109.88 A	109.28 C	0.13 AB	98.79 AB
	Sakha (1)	84.27 G	176.86 C	130.56 C	109.83 A	122.28 AB	0.2 A	93.83 c
	Sakha (3)	96.09 F	201.76 B	148.92 B	110.02 A	110.33 BC	0.14 AB	100.94 A
	Sakha (4)	99.78 F	202.67 B	151.22 B	103.09 A	131.64 A	0.14 AB	98.08 AB
	<b>time</b>	<b>96.86 B</b>	<b>201.58 A</b>					

**Table 3.** Effect of soaking period in saline water on water imbibition (ml/100 seed) and seed volume (cm<sup>3</sup>/100 seed) of five faba bean varieties

Characters	Imbibed water (ml /100 seed)					
	Salinity (NaCl PPM)	Variety	First 9-hour soaking	Second 9-hour soaking	Total imbibed 18-hour soaking	Mean
Distilled water	Nubaria (1)		117.21 j	72.71 p	189.91 a	<b>126.61 A</b>
	Giza (843)		82.28 o	45.33 tu	127.61 fg	<b>85.07 G</b>
	Sakha (1)		92 lm	28 x	120 hij	<b>80 HI</b>
	Sakha (3)		96.99 kl	53.45 rs	150.43 c	<b>100.29 C</b>
	Sakha (4)		90.33 lmn	43.83 uv	134.17 ef	<b>89.44 EF</b>
	<b>Mean</b>		<b>95.76 C</b>	<b>48.66 F</b>	<b>144.42 A</b>	<b>96.28 A</b>
1000. PPM	Nubaria (1)		102.83 kl	59 r	161.83 b	<b>107.89 B</b>
	Giza (843)		86.82 mno	31.26 wx	118.08 ij	<b>78.72 I</b>
	Sakha (1)		97.22 kl	29.71 wx	126.93 fgh	<b>84.62 G</b>
	Sakha (3)		83.54 no	45.72 tu	129.25 fg	<b>86.17 FG</b>
	Sakha (4)		91.94 lm	51.33 st	143.28 d	<b>95.52 D</b>
	<b>Mean</b>		<b>92.47 D</b>	<b>43.4 G</b>	<b>135.88 B</b>	<b>90.58 B</b>
2000 PPM	Nubaria (1)		95.72 kl	65.83 q	161.55 b	<b>107.7 B</b>
	Giza (843)		87.78 mno	36.89 vw	124.67 ghi	<b>83.11 GH</b>
	Sakha (1)		82.76 no	36.73 vw	119.49 ij	<b>79.66 HI</b>
	Sakha (3)		84.87 mno	45.09 tu	129.97 fg	<b>86.65 FG</b>
	Sakha (4)		95.71 kl	41.3 uv	137.02 de	<b>91.34 E</b>
	<b>Mean</b>		<b>89.37 E</b>	<b>45.17 G</b>	<b>134.54 B</b>	<b>89.69 B</b>
AVERAGE	Nubaria (1)		105.26 D	65.85 H	171.1 A	<b>114.07 A</b>
	Giza (843)		85.62 G	37.83 J	123.45 C	<b>82.3 C</b>
	Sakha (1)		90.66 EF	31.48 K	122.14 C	<b>81.43 C</b>
	Sakha (3)		88.47 FG	48.09 I	136.55 B	<b>91.04 B</b>
	Sakha (4)		92.66 E	50.16 I	142.82 B	<b>92.1 B</b>
	Time		92.53 B	46.68 C	139.21 A	



Table 3. Cont.

Characters	Seed volume (cm <sup>3</sup> /100 seed)				
	Salinity (NaCl ppm)	Variety	Non soaked (row seed)	First 9-hour soaking	Total 8-hour soaking
Distilled water	Nubaria (1)	122.67 mnop	212.58 bc	269.97 a	201.74 A
	Giza (843)	72 r	121.33 nop	145.89 ijklmn	113.07 GH
	Sakha (1)	69.33 r	153.33 ghijkl	161.33 fghijk	128 DEF
	Sakha (3)	86.67 qr	143.83 ijklmn	195.71 cd	142.07 CD
	Sakha (4)	80 r	158.17 ghijkl	192.22 cde	143.46 C
	<b>Mean</b>	<b>86.13 E</b>	<b>157.85 C</b>	<b>193.03 A</b>	<b>145.67 A</b>
1000. PPM	Nubaria (1)	116 p	172.62 defgh	232.12 b	173.58 B
	Giza (843)	68 r	134.39 lmno	147.98 hijklm	116.9 FGH
	Sakha (1)	72 r	156.36 ghijkl	166.38 fghij	131.58 CDE
	Sakha (3)	85.33 qr	160.54 fghijkl	169.17 efghi	138.35 CD
	Sakha (4)	81.33 r	157 ghijkl	184.56 def	140.96 CD
	<b>Mean</b>	<b>84.53 E</b>	<b>156.18 C</b>	<b>180.04 B</b>	<b>140.27 B</b>
2000 PPM	Nubaria (1)	108 pq	168.28 efghij	233.33 b	196.87 B
	Giza (843)	65.33 r	121.78 nop	135.39 klmno	107.5 H
	Sakha (1)	76 r	136.02 klmno	154.43 ghijkl	122.15 EFG
	Sakha (3)	80 r	142.24 jklmn	164.98 fghij	129.08 CDEF
	Sakha (4)	77.33 r	144.17 ijklmn	176.6 defg	132.07 CDE
	<b>Mean</b>	<b>81.33 E</b>	<b>142.5 D</b>	<b>172.95 B</b>	<b>132.026 B</b>
Average	Nubaria (1)	115.56 E	184.49 B	245.14 A	181.73 A
	Giza (843)	68.44 G	125.94 E	143.09 D	112.47 D
	Sakha (1)	72.44 FG	148.57 CD	160.72 C	127.24 C
	Sakha (3)	84 F	148.87 CD	176.62 B	136.5 B
	Sakha (4)	79.56 FG	153.11 CD	184.46 B	139.04 B
	<b>Time</b>	<b>84 C</b>	<b>152.2 B</b>	<b>182.01 A</b>	

Regarding to varieties, soaking time interaction, data showed the highest seed volume recorded by Nubaria (1) (245.14 cm<sup>3</sup>) for 18 hour soaking, whereas, the lowest seed volume were recorded in all variety row seeds and Giza (843) (125.94 cm<sup>3</sup>) in the first 9 hour soaking only. The interaction between varieties and salinity presented that Nubaria (1) had the highest seed volume (201.74 cm<sup>3</sup>) using distilled water. On the contrary, data showed that Giza (843) recorded lowest seed volume (107.5 cm<sup>3</sup>) using higher NaCl concentration (2000 ppm). The interaction between variety, soaking period and salinity recorded, the higher seed volume of Nubaria (1) (269.97 cm<sup>3</sup>) within 18 hour soaking, whereas, the lowest seed volume was recorded in row seed varieties and Giza (843) in the 1st 9 hour soaking in both distilled water and saline water (2000 ppm NaCl) within the 2nd 9hr of soaking in distilled water (Table 3).

### DISCUSSION

Quality of germination and good sprouting are important for plant emergence and development. Good germination, enough to produce seedlings for plant population and edible sprouts. The seed germination quality is expressed by characters such as percentage and index of germination. Seedlings healthy growth showed good radical growth, which increased best population of plant, particularly in saline conditions. Therefore, a test of germination could be useful tool using it for selection and development varieties for production salted soil or production of edible sprouts using saline water especially in arid zone (Abdallah et al 1991; Ibrahim et al 2016; Khan and Gulzar, 2003; Zhou et al 2010).

Five varieties of faba bean from Egypt showed similar trends to salinity stress with wheat Ibrahim et al (2016); Kandil et al (2012) but in our faba bean experiment, their differences in reduction of water uptake, percentage of germination, and radical initiation. The differences suggest that a test of germination may help in suitable faba bean varieties selected for saline zone soils or for sprout production in arid zone.

In this study, a decreasing in water uptake by weight or volume was showed with NaCl concentrations increasing in faba bean varieties. These agreement with wheat and sorghum reports by Ibrahim et al (2016); Kader and Jutzi, (2002). The osmotic barrier owing to NaCl concentration affected water uptake (Mehmet, 2006). Faba bean

data similar to other researchers (Akbarimoghaddam et al 2011; Ilker, 2011; Mehmet, 2006), who reported that germination of seed and earlier growth of seedling decreased in saline soils and water with different varieties responses. The germination of seed affected by NaCl may be related to generate an external osmotic potential which restrict water of uptake. Moreover, several studies demonstrated reducing of water absorption in ryegrass, wheat and barley (Bağcı et al 2003; Ibrahim et al 2016; Ilker, 2011) under abiotic stress such as salinity or water stresses.

In the current study, the lower percentage of germination and germination index was recorded in 2000 ppm NaCl concentration. The decline in percentage and index of germination using NaCl solution attributed to combined effect of osmotic pressure as reported by Moud and Maghsoudi, (2008); Munns, (2002) and salt concentrations toxicity (Ibrahim et al 2016; Mehmet, 2006; Munns and Tester, 2008) or owing to effect of Cl<sup>-</sup> (Almodares et al 2007) which increase osmotic stress. Higher Na<sup>+</sup> and Cl<sup>-</sup> ions uptake in germination using saline solution can induce toxicity for cells which slows or inhibits the proportion of germination and finally reduces percentage of germination (Taiz and Zeiger, 2003). In our research, the varieties responses differently to three different NaCl concentrations. Data means that there are genetic variations regarding to varieties in respect of abiotic saline stress tolerance. Increasing NaCl concentration showed decrease in percentage and index of germination in all faba bean varieties. Some of the varieties were more tolerant than the others. Result of salinity with variety interaction showed that the germination percentage of varieties at NaCl 1000 ppm Nubaria (1) reached the highest germination percentage (80%) while, Giza (843) had the highest germination percentage of (68%) at the 2000 ppm NaCl concentration. Similar results were recorded by Li, (2008) in Kuntze, Sieb and Stapf; Zhang et al (2010) in barely Kumari and Vishnuvardhan, (2015) in Kodo millet; El-Bastawisy et al (2018) in faba bean. Begum et al (2010) indicated that salt stress induced delay and decrease in germination percentage through higher accumulation of Na<sup>+</sup> and Cl<sup>-</sup> and lower water uptake.

The radical length is powerful character for salinity stress since radical is direct touch the media and uptake water from the media or saline solutions and supply it to the whole plant. For this sense, radical length provides a great evidence for

plant response to salt stress (Al-Karaki, 1997; Carpıcı et al 2009; Ibrahim et al 2016). Reduction in growth as a result of saline stress also published by other researcher in other species Li, (2008) in Kuntze, Sieb and Stapf; Carpıcı et al., (2009) on maize; Ibrahim et al 2016 on wheat. The increase in salinity concentration decreased the length of radical and dry weight of faba bean varieties. Similar data reported by Hussein et al (2007); Carpıcı et al (2009) in maize. Genotypes responded variously to salt stress.

Faba bean sprout fresh and dry weight were also recorded in the varieties which having higher germination percentage. Nubaria (1) recorded the highest sprout fresh & dry weight while, Giza (843) recorded the lowest fresh & dry sprout weight. The results in this study agree with Hussein et al (2007) in maize and Ibrahim et al (2016) in wheat. They recorded negative relationship identified between vegetative growth characters and increasing salinity. In addition, the sprout dry weight losses % indicated significant differences among varieties. Giza (843) had the highest losses (20.66%) followed by Nubaria (1) (16.59%) whereas Sakha (1) had the lowest losses of (5.29%). Salinity concentrations showed significant differences regarding sprout weight losses%. This is agree with the results of other researchers (Bahrami and Razmjoo, 2012; Begum et al 2013; Ibrahim et al 2016).

Salt tolerance of faba bean varieties decreased as the NaCl concentrations increased. The lowest salt tolerance index was registered in 2000 ppm NaCl concentration. These results agreed with Carpıcı et al (2009) in maize; Kumari and Vishnuvardhan, (2015) in spinach Alom et al (2016) and Ibrahim et al (2016) in wheat; Kan et al (2016) in soybean.

As the NaCl concentration increases the relative salt injury rate was increased in all faba bean varieties. The relative salt injury rate recorded no damage in all varieties using distilled water while, Sakha varieties recorded the highest injury when using 2000 ppm NaCl concentration data was more pronounced with Sakha (1). This damage increment may be due to change in cell expansion process which is controlled by processes related to cellular water uptake and cell wall extension (Ibrahim et al 2019; Kumari and Vishnuvardhan, 2015; Li, 2008).

Water imbibition was significantly affected by variety, salinity and soaking period and their interactions. Nubaria (1) recorded the highest imbibed

amount of water (114.07 ml) while, the lowest imbibed water (81.43 ml) was recorded in Sakha (1). The seeds water absorption in saline solutions decreased with of NaCl concentration increase. Concerning soaking period data showed the higher imbibed water within 18 hour soaking, moreover the absorption of water in the first 9-hour soaking (92.53 ml) was almost twice the absorption in the second 9-hour soaking (46.68 ml) similar data was obtained by Akbarimoghaddam et al (2011); Ibrahim et al (2016); Ilker, (2011); Kader and Jutzi, (2002); Mehmet, (2006). Begum et al (2010) reported the accumulation of sodium and Chloride ions increased with increasing salinity. The result indicated that salinity induced lower water absorption (Begum et al 2010).

The volume (cm<sup>3</sup>) of 100 seeds in NaCl concentration decreased with increasing concentration, seed volume was decreased from (145.67 cm<sup>3</sup>) to (132.26 cm<sup>3</sup>). Volume of seed s showed a highly significant difference among varieties, Nubaria (1) recorded the highest volume (181.73 cm<sup>3</sup>) while, the lowest volume was recorded (112.97 cm<sup>3</sup>) in Giza (843). Regarding soaking period data showed that the imbibed water for 9 hour increased row seed volume by about 81.2% while soaking for 18 hours increased volume by about 116.7% than row seeds and about 19.6% than the volume of seeds soaked for 9 hours. Panuccio et al (2014) also found the volume decrease with increasing salinity through lower water absorption.

## CONCLUSION

It can be concluded the uptake of water decreased with increasing salinity. Giza (843) and Sakha (3) varieties had recorded the highest tolerance to salinity. These genotypes were more tolerant to salinity and recommended to use them in breeding program for increasing faba bean production in Egypt. Therefore, these genotypes could be grown on slightly saline condition. However, this could not be a guarantee for these faba bean genotype to be salt-tolerant in later stages of plant growth. There must be additional experimental research to be conducted at the mature plant growth stage and then recommended salt concentration that provide best separation of faba bean varieties on the stage of germination and vegetative stage.

## REFERENCES

- Abdallah M.M. 2008. Seed Sprouts, a Pharaoh's Heritage to Improve Food Quality. *Arab Universities J. of Agric. Sci.* **16**, 469-478.
- Abdallah M.M., Jones R., El-Beltagy A. and Elmore C. 1991. Interaction of Salinity and Herbicides in Tomato. *Yemen J. Agric. Sci.* **1**, 108-121.
- Akbarimoghaddam H., Galavi M., Ghanbari A. and Panjehkeh N. 2011. Salinity Effects on Seed Germination and Seedling Growth of Bread Wheat Cultivars. *Trakia J. of Sci.* **9**, 43-50.
- Al-Karaki G.N. 1997. Barley response to salt stress at varied levels of phosphorous., *J. of Plant Nutrition (USA)*. **20**, 1635-1643.
- Almodares A. and Hadi D. 2007. Effect of salt stress on germination percentage and seedling growth in sweet sorghum cult. *Biological Sci.* **7**, 1492-1495.
- Alom R., Hasan M.A., Islam M.R. and Wang Q.F. 2016. Germination characters and early seedling growth of wheat (*Triticum aestivum* L.) genotypes under salt stress conditions. *J. of Crop Sci. and Biotechnology*. **19**, 383-392.
- Bağcı S.A., Ekiz H. and Yilmaz A. 2003. Determination of the salt tolerance of some barley genotypes and the characteristics affecting tolerance. *Turkish J. of Agric. and Forestry* **27**, 253-260.
- Bahrami H. and Razmjoo J. 2012. Effect of salinity stress (NaCl) on germination and early seedling growth of ten sesame cultivars (*Sesamum indicum* L.). *Int. J. of Agric. Sci.* **2**, 529-537.
- Baskin J.M., Baskin C.C. and Dixon K.W. 2006. Physical dormancy in the endemic Australian genus *Stylobasium*, a first report for the family Surianaceae (Fabales). *Seed Sci. Research* **16**, 229-232.
- Begum F., Ahmed I., Nessa A. and Sultana W. 2010. The effect of salinity on seed quality of wheat. *J. of the Bangladesh Agric. Univ.* **8**, 19-22.
- Begum M.A.J., Selvaraju P. and Venudevan B. 2013. Saline stress on seed germination. *Acedemic J.* **8**, 1420-1423.
- Bojović B., Đelić G., Topuzović M. and Stanković M. 2010. Effects of NaCl on seed germination in some species from families Brassicaceae and Solanaceae. *Kragujevac J. Sci.* **32**, 83-87.
- Carpıcı E.B., Celik N. and Bayram G. 2009. Effects of salt stress on germination of some maize (*Zea mays* L.) cultivars. *African J. of Biotechnology* **8**, 4918-4922.
- Copeland L.O., Lawrence O. and McDonald M.B. 2001. Principles of seed science and technology. Kluwer Academic Publishers.
- Duc G., Bao S., Baum M., Redden B., Sadiki M., Suso M.J., Vishniakova M. and Zong X. 2010. Diversity maintenance and use of *Vicia faba* L. genetic resources. *Field Crops Research*. **115**, 270-278.
- El-Bastawisy Z.M., El-Katony T.M. and Abd El-Fatah S.N. 2018. Genotypic variability in salt tolerance of *Vicia faba* during germination and early seedling growth. *J. of King Saud Univ., Sci.* **30**, 270-277.
- Erba D., Angelino D., Marti A., Manini F., Faoro F., Morreale F., Pellegrini N. and Casiraghi M.C. 2019. Effect of sprouting on nutritional quality of pulses. *Int. J. of Food Sci. and Nutrition* **70**, 30-40.
- FAO 2017. FAOSTAT. URL <http://www.fao.org/faostat/en/>
- Ghazi A.K. 1998. Seed size and water potential effects on water uptake, germination and growth of lentil. *J. of Agronomy and Crop Sci.* **181**, 237-242.
- Hendawey M.H. and Younes A.M.A. 2013. Biochemical evaluation of some faba bean cultivars under rainfed conditions at El-Sheikh Zuwayid. *Annals of Agric. Sci.* **58**, 183-193.
- Hossain M.S. and Mortuza M.G. 2006. Chemical Composition of Kalimatar, a Locally Grown Strain of Faba Bean (*Vicia faba* L.). *Pakistan J. of Biological Sci.* **9**, 1817-1822.
- Hussein M.M., Gaballah M.S. and Balbaa L.K. 2007. Salicylic Acid and Salinity Effects on Growth of Maize Plants. *Research J. of Agric. and Biological Sci.* **3**, 321-328.
- Ibrahim M.E.H., Zhu X., Zhou G. and Nimir N.E.A. 2016. Comparison of germination and seedling characteristics of wheat varieties from China and Sudan under salt stress. *Agronomy J.* **108**, 85-92.
- Ibrahim M.H., Abas N.A. and Zahra S.M. 2019. Impact of Salinity Stress on Germination of Water Spinach (*Ipomoea aquatica*). *Annual Research & Review in Biology*. pp. 1-12.
- Ilker N. 2011. Effects of salinity stress on water uptake, germination and early seedling growth of perennial ryegrass. *African J. of Biotechnology* **10**, 10418-10424.
- Jamil M., Deog Bae L., Kwang Yong J., Ashraf M., Sheong Chun L. and Eui Shik R. 2006. Effect of Salt (NaCl) Stress on Germination and Early Seedling Growth of Four Vegetables Species. *J. of Central European Agric.* **7**, 273-282.

- Kader M.A. and Jutzi S.C. 2002. Temperature, osmotic pressure and seed treatments influence imbibition rates in sorghum seeds. *J. of Agronomy and Crop Sci.*, **188**, 286-290.
- Kan G., Ning L., Li Y., Hu Z., Zhang W., He X. and Yu D. 2016. Identification of novel loci for salt stress at the seed germination stage in soybean. *Breeding Sci.*, **66**, 530-541.
- Kandil A., Sharief A. and Elokda M. 2012. Germination and Seedling Characters of Different Wheat Cultivars under Salinity Stress. *J. of Basic and Applied Sci.*, **8**, 585-596.
- Khan M.A. and Gulzar S. 2003. Germination responses of *Sporobolus ioclados*: a saline desert grass. *J. of Arid Environments* **53**, 387-394.
- Khan M.A., Ungar I.A. and Showalter A.M. 2000. Effects of sodium chloride treatments on growth and ion accumulation of the halophyte *Haloxylon recurvum*. *Communications in Soil Sci. and Plant Analysis*. **31**, 2763-2774.
- Khatun M., Hafiz M.H.R., Hakim M. and Siddiqui M.N. 2013. Responses of Wheat Genotypes to Salt Stress in Relation to Germination and Seedling Growth. *Int. J. of Bio-resource and Stress Management* **4**, 635-640.
- Kumari R.P. and Vishnuvardhan Z. 2015. Effect of salinity on growth, protein and antioxidant enzymes in three Kodo millet (*Paspalum scrobiculatum*) germplasm. *Int. J. of Current Microbiology and Applied Sci.*, **4**, 475-483.
- Kumawat S., Gothwal D.K., Kumawat K.R. and Sharma R.K.M. 2017. Effect of salt stress on seed germination and early seedling traits in fenugreek (*Trigonella foenum-graecum* L.) genotypes grown under different salinity levels, *J. of Pharmacognosy and Phytochemistry* **6(5)**, 776-781.
- Li Y. 2008. Effect of salt stress on seed germination and seedling growth of three salinity plants. *Pakistan J. of Biological Sci., PJBS*. **11**, 1268-1272.
- Manz, B., 2005. Water Uptake and Distribution in Germinating Tobacco Seeds Investigated in Vivo by Nuclear Magnetic Resonance Imaging. *Plant Physiology* **138**, 1538-1551.
- Mehmet A., 2006. Effects of NaCl on the Germination, Seedling Growth and Water Uptake of Triticale. *Turkish J. of Agric. and Forestry* **30**, 39-47.
- Moud A.M. and Maghsoudi K. 2008. Salt Stress Effects on Respiration and Growth of Germinated Seeds of Different Wheat (*Triticum aestivum* L.) Cultivars. *World J. of Agric. Sci.* **4**, 351-358.
- Munns, R., 2002. Comparative physiology of salt and water stress. *Plant, cell & environment*. **25**, 239-250.
- Munns R. and Tester M. 2008. Mechanisms of Salinity Tolerance. *Annual Review of Plant Biology* **59**, 651-681.
- Okçu G., Kaya M.D. and Atak M. 2005. Effects of salt and drought stresses on germination and seedling growth of pea (*Pisum sativum* L.). *Turkish J. of Agric. and Forestry.*, **29**, 237-242.
- Orozco-Segovia A., Márquez-Guzmán J., Sánchez-Coronado M.E., Gamboa De Buen A., Baskin J.M. and Baskin C.C. 2007. Seed anatomy and water uptake in relation to seed dormancy in *Opuntia tomentosa* (Cactaceae, Opuntioideae). *Annals of Botany*. **99**, 581-592.
- Panuccio M.R., Jacobsen S.E., Akhtar S.S. and Muscolo A. 2014. Effect of saline water on seed germination and early seedling growth of the halophyte quinoa. *AoB PLANTS*. **6**, plu 047.
- Santos J.M.A.P. dos, Francisco de A. de Oliveira, J.F. de M., Targino, Ana J. de O., Santos, Costa, L.P. da and Dos S.T. 2018. Saline stress and potassium/calcium ratio in fertigated eggplant. *Revista Brasileira de Engenharia Agrícola e Ambiental.*, **22**, 770-775.
- Schopfer P. and Plachy C. 1984. Control of Seed Germination by Abscisic Acid: II. Effect on Embryo Water Uptake in *Brassica napus* L. *Plant Physiology* **76**, 155-160.
- Silva B.M. da S.E., Silva C. de O.E., Mõro F.V. and Vieira R.D. 2018. Seed anatomy and water uptake and their relation to seed dormancy of *Ormosia paraensis* Ducke. *J. of Seed Sci.*, **40**, 237-245.
- Siswanto J., Prabuwo A.S. and Abdulah A. 2013. Volume Measurement of Food Product with Irregular Shape Using Computer Vision and Monte Carlo Method: A Framework. *Procedia Technology* **11**, 764-770.
- Snedecor G.W. and Cochran W.G. 1980. *Statistical Methods*. 7th Edition, Iowa State University Press, Ames.
- Taiz, L. and Zeiger E. 2003. *Plant physiology*. 3rd edn., Annals of Botany. Oxford Univ. Press.
- Zhang H., Irving L.J., McGill C., Matthew C., Zhou D. and Kemp P. 2010. The effects of salinity and osmotic stress on barley germination rate: sodium as an osmotic regulator. *Annals of Botany* **106**, 1027-1035.
- Zhou G., Ma B.L, Li J., Feng C., Lu J. and Qin, P. 2010. Determining Salinity Threshold Level for Castor Bean Emergence and Stand Establishment. *Crop Sci.* **50**, 2030-2036.





## تأثير الملوحة على صفات النبات لخمس أصناف من الفول

[180]

محمود عادل أحمد على\* - ممدوح محمد فوزي عبدالله - نشوي عطية أبو العزم - أحمد أبو اليزيد

قسم البساتين - كلية الزراعة - جامعة عين شمس - ص.ب. 68 - حدائق شبرا 11241 - القاهرة - مصر

\*Corresponding author: [mahmoud\\_adel489@agr.asu.edu.eg](mailto:mahmoud_adel489@agr.asu.edu.eg)

Received 25 July, 2019

Accepted 12 November, 2019

الملح لها تأثيراً سلبياً على امتصاص المياه ونسبة الإنبات وطول الجذير، الوزن الطازج والجاف للنبات، وحجم البذور ومعدل الإصابة النسبي بالملح. بكل تركيزات الملوحة، أظهرت الأصناف الخمس درجات مختلفة لتحمل الملوحة. سخا (3)، (4) وجيزة (843) أفضل مؤشر لتحمل الملوحة من نوبارية (1) وسخا (1). نوبارية (1)، جيزة (843) حصل على أعلى نسبة إنبات ومؤشر الإنبات، وسخا (1) كان أقصر طول الجذير وأكثر حساسية لمعدل الإصابة بالملح. وعلاوة على ذلك، الأنماط الجينية جيزة (843)، وسخا (3) يمكن اعتبارها متحملة للإجهاد الملحي مقارنة بتلك الأخرى. كما يمكن استخدام هذه الأنماط الجينية في برنامج تربية لتعزيز زراعة الفول في الأراضي المستصلحة حديثاً. وتشير البيانات إلى ان اختبار إنبات البذور كأداة اختيار مفيدة لاستخدامها في تطوير سلالات فول جديدة للإنتاج في التربة المالحة.

**الكلمات الدالة:** إنبات بذور فول الفول، امتصاص الماء، معدل إصابة الملح النسبية، مؤشر تحمل الملح، شخصية البرعم، إجهاد الملوحة

## الموجز

تتمثل إحدى الخصائص الفريدة في المراحل الأولى من النبات هو الانبات. الإنبات هو التعبير الظاهري لمختلف الأنشطة الأيضية في البذور. تعرض النبات للإجهادات غير الحيوية مثل الملوحة التي تؤثر على العمليات الفسيولوجية، والتشريحية، ونظم النمو ونمو النبات. الإنتاج الزراعي في المناطق القاحلة وشبه القاحلة منخفض يعزى للعديد من العوامل على سبيل المثال تراكم الأملاح في التربة والمياه، مجموع مساحة الأراضي تحت الملوحة هو حوالي 953 مليون هكتار. الهدف من هذا البحث كان لتقييم صفات الإنبات لخمس أصناف من الفول [نوبارية (1)، جيزة (843)، سخا (1)، سخا (3)، سخا (4)] تم استخدام المحلول الملحي الذي يحتوي على (0، 1000 و 2000 جزء في المليون كلوريد الصوديوم). واستخدمت لدراسة امتصاص الماء، نسبة الإنبات، مؤشر الإنبات، ومعدل الإصابة بالملح، طول الجذير، الوزن الطازج والجاف والفقد في الوزن وحجم البذور باستخدام المحلول الملحي. وأشارت النتائج إلى أن التركيزات العالية من

