

Circulating Adipocyte Fatty Acid Binding Protein (A-FABP): Predictor of Metabolic Syndrome in Patients with Type 2DM

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Abstract

Background: The circulating portion of adipocyte fatty acid binding protein (A-FABP) acts as a humoral factor potentially controlling inflammatory responses and insulin action in adipocyte. Most individuals with Type 2 diabetes mellitus (2DM) exhibit intra-abdominal obesity, insulin resistance and subsequent occurrence of metabolic syndrome (MetS).

Aim of Study: This work aimed at evaluating A-FABP as a marker for MetS in type 2DM.

Subjects and Methods: The study included (180) subjects divided into group I (120) adult patients with type 2DM diagnosed according to ADA 2016. They were subdivided into subgroup Ia (60 without MetS) and subgroup Ib (60 with MetS). Group II (control group) included (60) age- and sex-matched apparently healthy subjects. Circulating A-FABP was measured by ELISA. In addition, assay of fasting insulin (F.insulin), lipid profile, hsCRP, and calculation of HOMA-IR.

Results: Highly significant increase in levels of A-FABP, F.insulin, HOMA-IR and hsCRP was observed in subgroup Ia compared to control group (Z: 5.691, 4.902, 5.908, and 0.477; $p < 0.01$; respectively). Sub-group Ib versus sub-group Ia revealed significant elevated levels of A-FABP and F.Insulin (Z: 5.276, $p < 0.01$ and Z: 2.469, $p < 0.05$; respectively) while HOMA-IR and hsCRP showed no significant difference between both subgroups ($p > 0.05$). In subgroup Ib, there was a significant positive statistical correlation between A-FABP and weight and BMI ($r: 0.379$ and 0.386 , $p < 0.05$; respectively). Stepwise multi-regression analysis indicated that triglycerides and A-FABP are the most sensitive independent predictors of MetS in diabetic patients (F-ratio: 64.4 and $p < 0.001$).

Conclusion: High serum A-FABP level is strongly associated with MetS in type 2DM. It is a promising independent marker of MetS and is beneficial in its early diagnosis.

Key Words: Type 2DM – Metabolic syndrome – A-FABP – ELISA – predictor.

Introduction

THE association between excessive body weight and many diseases, particularly type 2 diabetes mellitus has been proved in various studies [1]. Besides lipid accumulation and free fatty acids (FFA) release, adipocytes together with adipose tissue stromal cells produce and release multiple signaling proteins termed adipokines. Adipocyte fatty acid binding protein (A-FABP) is a new adipokine suggested to be linked to obesity, insulin resistance, impaired glucose metabolism and eventually development of metabolic syndrome (MetS) [2,3].

Although A-FABP was originally identified as an abundant cytoplasmic protein in adipocytes, a portion of A-FABP is released into bloodstream. Circulating A-FABP is elevated in obese individuals and correlates positively with the features of the metabolic syndrome, and the incidence of atherosclerosis and cardiovascular diseases [4]. Accordingly, this study investigated the possible association between serum levels of A-FABP and the presence of MetS in type 2 diabetes mellitus, as well as its evaluation as a predictor marker for MetS in these patients.

Subjects and Methods

A- Subjects:

This study was conducted on (120) adult diabetic patients (group I) diagnosed according to the American Association of Diabetes, 2016 criteria for diagnosis of DM [5] presented to Endocrinology Department, Ain Shams University Hospital, Cairo, Egypt, between November 2017 and January 2019, and their matching (60) apparently healthy subjects serving as control group. Group I patients were

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further subdivided into 2 subgroups; subgroup Ia which included 60 diabetic patients without MetS (22 males and 38 females) and subgroup Ib included 60 diabetic patients with metabolic syndrome (26 males and 34 females). Patients enrolled in subgroup Ib were selected according to the inclusion criteria for MetS by National Cholesterol Education Program Adult Treatment Panel III (NCEP ATP III) [6].

Exclusion criteria: Patients were excluded if they have

- Acute infection.
- Acute myocardial infarction.
- Heart failure.
- Malignancy at the time of sampling.

An informed consent was taken from all subjects participating in the study before history taking, physical examination and blood sample withdrawal. The procedures applied in this study were approved by The Ethical Committee of Human Experimentation of Ain Shams University, and are in accordance with the Helsinki Declaration of 1975.

All subjects included in the study had detailed medical history taking, blood pressure measurement, anthropometric measures (weight, height, BMI calculation and waist circumference), measurement of fasting and 2 hours postprandial serum glucose, glycated haemoglobin (HbA1c), total cholesterol, triglycerides (TG), high-density lipoprotein cholesterol (HDL-C), low-density lipoprotein cholesterol (LDL-C), fasting serum insulin level, highly sensitive C-reactive protein (hsCRP), insulin resistance evaluated using a homeostasis model assessment of insulin resistance (HOMA-IR) and serum A-FABP level.

B- Sampling:

Under complete aseptic conditions, 10mL of fasting venous blood were obtained by a clean venipuncture, two milliliters were placed in EDTA tube for subsequent assay of HbA1c, while the rest was evacuated in two plain test tubes. The serum was separated by centrifugation (1 000x g for 15 minutes). Serum of one tube was immediately assayed for fasting serum glucose and lipid profile (cholesterol, triglycerides, HDL-C and LDL-C), while the serum collected in the other tube was divided in two aliquots and stored at -20°C for subsequent assay of hsCRP, fasting insulin and A-FABP. Two hours postprandial sample for glucose was collected, separated by centrifugation and assayed. Hemolysed samples were discarded. Repeated freezing and thawing was avoided.

C- Methods:

Analytical methods:

HbA1c was assayed by high performance liquid chromatography (HPLC) technique on the Bio-Rad d- 10 hemoglobin testing system (Bio-Rad Laboratories, Inc., 4000 Alfred Nobel Drive, Hercules, California 94547, USA) using reagents supplied by the company. Serum samples were assayed for glucose and lipid profile on the Beckman AU-680 system auto-analyzer (Beckman Coulter, Inc. Diagnostics Division Headquarters 250 South Kraemer Boulevard Brea, California 92821-6232 USA) using reagents supplied by the company.

Fasting serum insulin was assayed by commercially available ELISA kit supplied by Chemux BioScience, Inc (50 South Linden Ave., 7, South San Francisco, CA 94080, USA). Serum A-FABP was assayed by commercially available sandwich ELISA kit supplied by Elabscience Biotechnology Inc., USA (Building B 18, 2nd Phase of Biomedical Park, 858 Gaoxin Road, Donghu Hi-Tec Development Area, Wuhan, Hubei, USA). hsCRP was measured by immunoturbidimetry using reagents supplied from Biosystem S.A (Bio System S.A, Costa Brava, 30.08030 Barcelona Spain) on automated BioMaxima instrument (BioMaxima S.A. 20-277 Lublin, Vetterow 5, Poland).

Calculation of results:

$$\text{HOMA-IR} = \frac{\text{Fasting glucose (m/dL)} \times \text{Fasting insulin (IU/mL)}}{405}$$

Cutoff point to define insulin resistance corresponds to HOMA-IR \geq 3.8 [7].

A-FABP and Fasting insulin:

Construction of a standard curve is done by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and drawing a best fit curve through the points on the graph. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

hsCRP:

Agglutination between hsCRP in a sample and anti hsCRP is detected as an absorbance change (570nm), with the magnitude of the change being proportional to the quantity of hsCRP in the sample. The actual concentration is then determined by the interpolation from a calibration curve prepared from calibrators of known concentration [8].

Statistical methods:

All statistical analyses were done using software version IBM SPSS (Statistical Package for the Social Sciences) statistics (Version 25.0, IBM Corp., USA, 2017-2018). Data were expressed descriptively as median and interquartile range (IQR) for quantitative non parametric values and as percent for qualitative data. Comparative statistics were done using the Wilcoxn Rank Sum test and Kruskall Wallis test for non parametric data. Correlation analysis was performed using Spearman’s rank correlation coefficient (*r*) for non parametric data. Receiver operator characteristic (ROC) curves were constructed and optimal cut-off values for serum A-FABP and triglycerides were established by the best sensitivity and specificity where the right angle at the upper left corner is the best diagnostic threshold (cut-off) of the parameter being varied. Logistic multi-regression analysis was used to search for a panel (independent parameters) that can predict the target parameter (dependant variable). In all statistical analyses, *p*-value >0.05: Non significant; *p*-value <0.05: Significant; *p*-value <0.01: Highly significant.

Results

Descriptive and comparative statistics of different studied parameters among group Ia, Ib (patient subgroup) and group II (control group) using Kruskall-Wallis Test are seen in Table (1).

Table (2) shows statistical comparison between subgroup Ia versus group II Using Wilcoxn Rank Sum Test.

Table (3) shows comparative statistical analysis using Wilcoxn Rank Sum Test, between subgroup Ib versus subgroup Ia.

Using Ranked Sperman Correlation Test, correlation analysis of A-FABP and other studied parameters in subgroup Ib, revealed a significant positive statistical correlation between A-FABP and weight as well as BMI (*r*:0.379 and 0.386, *p*<0.05; respectively), in the studied population. However no significant statistical correlation is seen between A-FABP and other studied parameters (*p*>0.05).

Stepwise multi-regression analysis in Table (4) examined the effect of different parameters on A-FABP. Each non significant parameters is removed in a stepwise manner to reach the final step with TG, which indicated that TG and A-FABP are the most sensitive independent predictors of MetS in diabetic patients, and eliminates the role of HOMA-IR, F.Insulin and hsCRP as predictors of the disease, with (F. Ratio=64.4 and *p*=0.001) and the coefficient variant zero reflects that prediction formula for MetS in type 2 DM = -0.175+0.005(TG) + 0.095 (A-FABP).

Table (5) shows Receiver Operating Characteristic Curve (ROC) curve analysis for diagnostic performance of TG, A-FABP and their combination for discriminating sub-group Ib from combined sub-group Ia and group II (AUC; TG: 0.678, A.FABP: 0.904,combined TG and A-FABP:0.987).

Table (1): Descriptive and comparative statistics of different studied parameters among SubGroups Ia,Ib and Group II.

Parameter	Subgroup Ia (n=60) Median (Q1-Q3)	Subgroup Ib (n=60) Median (Q1-Q3)	Control group (II) (n=60) Median (Q1-Q3)	H	<i>p</i>
Weight (Kg)	83 (68.75-90.25)	100 (84.5-110)	72.5 (68-85.25)	23.543	<0.01
Height (cm)	166.5 (160-170.25)	165 (160-172.5)	170 (162.25-175)	2.763	>0.05
BMI (Kg/m ²)	28.25 (26.5-32)	35.5 (29-40)	26 (24-28.25)	35.238	<0.01
Waist (cm)	86 (78-95.75)	109 (94.5-120.5)	83.5 (78.75-91.25)	27.429	<0.01
FBS (mg/dL)	138.5 (101.75-173.75)	125.5 (101.5-158.25)	80 (75.75-86)	52.657	<0.01
2hPP (mg/dL)	191.5 (147.25-262.75)	203.5 (170.75-275)	97 (89.75-104)	51.223	<0.01
HbA1c (%)	6.9 (6.15-9.25)	7.35 (6.357-8.6)	5.2 (5-5.3)	56.846	<0.01
T.Chol (mg/dL)	170.5 (144.25-206.25)	191 (164-239.25)	165 (139-191)	5.937	>0.05
TG (mg/dL)	122.5 (87.5-148)	148 (111.5-196.5)	93 (84.5-122.25)	19.683	<0.01
HDL-C (mg/dL)	50.5 (43.25-57.25)	44.5 (39.75-49.75)	54 (46.5-60.25)	9.035	<0.05
LDL-C (mg/dL)	100.5 (77.5-117.75)	111.5 (86.25-150.75)	111 (97-123.75)	3.052	>0.05
F.Insulin (μU/mL)	11.5 (5-20.5)	19 (10.75-25)	8.5 (5.375-12)	22.785	<0.01
HOMA-IR	3.95 (2.175-7.9)	5.6 (3-9.375)	1.65 (0.975-2.225)	39.109	<0.01
A-FABP (ng/mL)	3.4 (0.2-6)	10.5 (8.25-14.25)	1.5 (0.675-2.625)	41.389	<0.01
hsCRP (mg/dL)	0.4485 (0.153-0.578)	0.6185 (0.282-1.218)	0.155 (0.079-0.235)	2.6106	<0.01

p-value >0.05: Non significant. *p*-value <0.05: Significant. *p*-value <0.01: Highly significant.

Table (2): Comparative statistics between SubGroup Ia versus Group II and Subgroup Ib versus Group (II) as regard different studied parameters.

Parameter	Group	Subgroup Ia vs Control group (II)		Subgroup Ib vs Control group (II)	
		Z	p	Z	p
F.Insulin (μ IU/mL)		1.986	<0.05	4.902	<0.001
HOMA-IR		4.533	<0.01	5.908	<0.001
A-FABP (ng/mL)		1.371	>0.05	5.691	<0.001
hsCRP (mg/dL)		0.366	<0.01	0.477	<0.001

p-value >0.05: Non significant.
 p-value <0.05: Significant.
 p-value <0.01: Highly significant.

Table (3): Comparative statistics between SubGroup Ib versus SubGroup Ia as regard different studied parameters.

Parameter	Z	p
F.Insulin (μ IU/mL)	2.469	<0.05
HOMA-IR	1.715	>0.05
A-FABP (ng/mL)	5.276	<0.01
hsCRP (mg/dL)	0.172	>0.05

p-value >0.05: Non significant.
 p-value <0.05: Significant.
 p-value <0.01: Highly significant.

Table (4): Stepwise logistic multi-regression analysis for different studied parameters using groups as dependent variable.

Item	Reg. Coef.	t	p	Sig.	F-ratio	p
(Constant)	-0.175	-1.237	0.219	>0.05		
TG (mg/dL)	0.005	5.233	0	<0.01		
A.FABP (ng/mL)	0.095	8.93	0	<0.01		
					64.368	<0.01

p-value >0.05: Non significant.
 p-value <0.05: Significant.
 p-value <0.01: Highly significant.

Table (5): Diagnostic performance of A-FABP, TG and their combined use for discriminating between Sub-Group Ib and combined SubGroup Ia with Group II.

Parameter	Cutoff	% Specificity	% Sensitivity	% NPV	% PPV	% Efficiency
TG.	144mg/dL	86.7	53.3	78.8	66.7	75.6
A-FABP	8ng/mL	96.7	76.7	89.2	92	90
Combined TG and A-FABP	88mg/dL and 8ng/mL	98.3	100	100	96.8	98.9

Discussion

Patients with MetS have two- to three-fold increased risk for the development of cardiovascular morbidity and mortality [9]. In nearly 20-25% of apparently healthy individuals and in 45% of patients with clinical manifestations of atherosclerosis and MetS is present [10].

Adipose tissue plays a key role in the development of insulin resistance and its pathological sequelae, such as type 2 diabetes, MetS and non-alcoholic fatty liver disease. Dysfunction in the adipose tissue response to storing excess fatty acids as TG can lead to adipose tissue inflammation and spillover of fatty acids from this tissue and accumulation of fatty acids as lipid droplets in ectopic sites, such as liver and muscle [11].

The adipokine A- FABP, also known as FABP4 or aP2, is not abundantly expressed in adipocytes, but also produced in macrophages, endothelial cells and glial cells. A-FABP functions as a lipid chaperone that regulates trafficking, fluxes and signaling of FFAs, and has an important role in linking lipid metabolism with inflammation [12].

In humans, a promoter polymorphism, T-87C, of the A-FABP gene that resulted in reduced adipose tissue A-FAPB mRNA expression was found to be associated with reduced risk for type 2 DM and cardiovascular diseases [13]. Based on these findings, the present study hypothesized that serum A-FA BP level may be a useful marker for identifying DM patients susceptible to developing MetS as well as its evaluation as a predictor marker for MetS.

The results of the present study showed a highly significant increase in serum A-FABP levels in diabetic patients with MetS when compared to the diabetic subjects without MetS. Furthermore, elevated serum A-FABP levels were positively correlated with markers of MetS namely weight and BMI in sub-group Ib. These findings are in accordance with those of Xu et al. [14], Yeung et al. [15] and Mauvais [16]. These results are explained by the fact that adipose tissue is probably a significant contributor of A-FABP into the circulatory system through Extracellular vesicles. Therefore; A-FABP might clinically serve as a marker for adiposity which in turn is an important component of MetS [14,16].

Subcutaneous adipose tissue functions as a storage depot for excess fatty acids, visceral adipose tissue is more closely linked to the adverse metabolic and inflammatory profile observed in indi-

viduals with obesity and insulin resistance [17]. When individuals become obese, their excess caloric intake is stored in the form of triglycerides within the adipocytes of white adipose tissue. Excess fatty acids are stored in existing mature adipocytes leading to an increase in their size (hypertrophic expansion) [18]. Larger adipocytes tend to be more dysfunctional, and they become insulin resistant resulting in increased lipolysis due to resistance to the anti-lipolytic effects of insulin [19]. Failure of angiogenesis and provision of an adequate blood supply to hypertrophic adipocytes leads to necrosis, macrophage infiltration into adipose tissue and inflammation and adipokine release specially A-FABP. "Spillover" of fatty acids unable to be retained in subcutaneous adipocytes leads to an increase in the visceral fat compartment and eventually flux of fatty acids into ectopic sites, stored as intracellular lipid droplets in tissues, such as liver and the pancreas. The formation of ectopic fat is closely linked to the development of IR and Type 2 DM and metabolic disease [20].

In the present study, notably, the stepwise linear regression analysis adds power to our findings that the best increase in the sensitivity for A-FABP to detect MetS patients was achieved by adding TG. Therefore, A-FABP and TG levels were being the least most sensitive predictor for development of MetS with type 2 DM in studied population. These findings are in accordance with the previous findings of Terra et al. [21] and Li et al. [22] who found through a logistic regression analysis that A-FABP concentration is the only cytokine significantly increases in patients with MetS than patients without. The relationship between A-FABP and lipid metabolism was partly explained by several studies Oram and Lawn, [23]; Makowski et al. [24] and Park et al. [25] who reported that A-FABP altered the cholesterol efflux pathway in macrophages. A-FABP is a critical regulator of the peroxisome proliferator-activated receptors-liver X receptor α -adenosine triphosphate (ATP)-binding cassette A1 (ABCA1) pathway and contributes to the control of cholesterol trafficking in macrophages. (ABCA1) regulates the rate-limiting step in HDL biogenesis via the efflux of intracellular cholesterol, which might stimulate transfer of triglycerides to HDL and triglycerides catabolism. However, the pathologic effect of A-FABP at a cellular level requires clarification with further researches [25].

In an attempt to find a laboratory tool that can discriminate diabetic patients with metabolic syndrome from diabetic patients without MetS combined with control group, ROC analysis was constructed for A-FABP and the best chosen cut-off

in discriminating between groups was 8ng/mL showing 76.7% diagnostic sensitivity, 96.7% diagnostic specificity, 92% positive predictive value, 89.2% negative predictive value 90% diagnostic efficacy and area under the curve (AUC) of 0.904. These remarkable findings were improved in combined use of A-FABP and TG reaching excellent diagnostic performance in discriminating diabetic patients with MetS versus those without, where the AUC approached 1.0 (0.987). The best diagnostic cut-off levels of the A-FABP and TG were 8ng/mL and 88mg/dL, respectively, achieving diagnostic sensitivity, diagnostic specificity, positive predictive value, negative predictive value and diagnostic efficacy of 100%, 98.3%, 96.8%, 100% and 98.9%, respectively. To our knowledge no other studies had discussed the association between A-FABP and TG as diagnostic tool for MetS.

In accordance with our findings, a previous study by Stejskal and Karpisek [26] conducted on 67 healthy subjects and 71 subjects with metabolic syndrome in a Caucasian population showed that A-FABP at a cut-off level of 16.4ng/mL revealed sensitivity of (40%), specificity of (99%) and AUC was 0.77.

hsCRP as a marker of inflammation and a part of the cardiac risk assessment was analyzed to explore its pathologic effect in MetS and its correlation with A-FABP in such patients. In our study, hsCRP levels didn't significantly differentiate between patients with MetS from those without, as well as it was not correlated with A-FABP levels, these in agreement with Grundy [27] and Makowski et al. [24] studies. On the other hand, other studies hypothesized that A-FABP could be a marker for inflammation and MetS through a positive relationship between A-FABP and systemic inflammation reflected by C-reactive protein [28].

Although, several studies showed that A-FABP has a causal role in systemic inflammation, leading to the development of MetS, it appears that the relationship between A-FABP and MetS was independent of the presence of inflammatory markers. The mechanism remains unclear and excessive productions of A-FABP by obese adipose tissue do not fully overlap those of the inflammatory pathways [25].

A cornerstone in our study was the assessment of the correlation between A-FABP, F.insulin and HOMA-IR. A remarkable finding was that the non significant correlation between A-FABP, F.Insulin and HOMA-IR in diabetic patients with MetS.

These results are in agreement with those of Eckel et al. [29]; Xu et al. [30]; Tso et al. [31] and Park et al. [25] who explained that plasma A-FABP level was found to be a strong predictor of metabolic syndrome in type 2 DM patients independently of the traditional risk factors including insulin resistance and glycemic indexes. In addition study of Karpisek et al. [32] suggests that an elevated baseline A-FABP level may enhance the MetS phenotype via mechanisms other than insulin resistance.

On the other hand, Tso et al. [31] and Li et al. [22] reported that elevated baseline serum A-FABP levels predict the development of type 2 DM and development of insulin resistance as A-FABP positively correlated with insulin levels. Although the precise mechanisms explaining the role of A-FABP in glucose metabolism are not yet fully understood, the link between A-FABP and adipose tissue may affect glucose metabolism [33].

Selective deletion of A-FABP in adipocytes resulted in reduced expression of inflammatory cytokines in macrophages, whereas the same deletion in macrophages leads to enhanced insulin signaling and glucose uptake in adipocytes [34]. Therefore, there is a possibility that A-FABP might affect interactions between adipocytes and macrophages, leading to altered insulin sensitivity and glucose metabolism.

The results of the present study showed a highly significant increase in serum A-FABP levels in diabetic subjects with MetS, when compared to diabetic patients without MetS and healthy control group. Furthermore, elevated serum A-FABP levels were positively correlated with markers of MetS weight and BMI. Further studies are required to elucidate the relationship between A-FABP and development of DM and medication used.

Conflict of interest:

All authors declare that they have no financial or institutional interests related to this manuscript.

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دور البروتين الدهني الرابط للأحماض الدهنية بالدم في التنبؤ بمتلازمة الأيض في مرضى السكري من النوع الثاني

خلفية: يعمل الجزء الموجود بمصل الدم من البروتين الدهني الرابط للأحماض الدهنية كعامل خلطي يحتمل أن يتحكم في الاستجابات الالتهابية وعمل الأنسولين في الخلايا الدهنية. وقد معظم الأفراد الذين يعانون من مرض السكري النوع الثاني يعانون من السمنة داخل البطن، ومقاومة الأنسولين وحدوث متلازمة الأيض.

الهدف: تهدف هذه الدراسة إلى تقييم البروتين الدهني الرابط للأحماض الدهنية كعلامة لحدوث متلازمة الأيض في مرضى السكري من النوع الثاني.

المواد وطرق البحث: شملت الدراسة (١٨٠) من الأفراد مقسماً إلى مجموعتين: المجموعة الأولى (١٢٠) من المرضى البالغين المصابين بمرض السكري النوع الثاني الذين تم تشخيصهم وفقاً للمنظمة الأمريكية لمرضى السكري ٢٠١٦. وتم تقسيم هذه المجموعة إلى مجموعتين فرعيتين Ia (٦٠) مريض سكري من النوع الثاني بدون متلازمة الأيض ومجموعة فرعية Ib (٦٠) مريض سكري من النوع الثاني مع متلازمة الأيض. وشملت المجموعة الثانية (المجموعة الضابطة) (٦٠) من الأصحاء من نفس الفئة العمرية والجنس. تم قياسه البروتين الدهني الرابط للأحماض الدهنية في مصل الدم بتقنية فحص الإتصاص المناعي المرتبط بالأنزيم ELISA بالإضافة إلى ذلك، فحص الأنسولين الصائم، مجموعة الدهون في المصل، hsCRP، وحساب HOMA-IR.

النتائج: لوحظ زيادة كبيرة في مستويات البروتين الدهني الرابط للأحماض الدهنية، الأنسولين الصائم، HOMA-IR و hsCRP في المجموعة الفرعية Ia مقارنة بالمجموعة الضابطة (على التوالي $p < 0.01$ (Z: 5.691, 4.902, 5.908, and 0.477). كشفت المقارنة بين المجموعة الفرعية Ib مقابل المجموعة الفرعية Ia عن مستويات مرتفعة كبيرة من مستويات البروتين الدهني الرابط للأحماض الدهنية، الأنسولين الصائم (على التوالي $p < 0.01$ (Z: 5.276, $p < 0.05$ and Z: 2.469). بينما HOMA-IR و hsCRP لم تظهر أى اختلاف كبير بين المجموعتين الفرعيتين ($p > 0.05$) في المجموعة الفرعية Ib كان هناك ارتباط إحصائي إيجابي كبير بين مستويات البروتين الدهني الرابط للأحماض الدهنية والوزن ومؤشر كتلة الجسم ($r = 0.379$ و $p < 0.05$ على التوالي) أشار تحليل الانحدار المتعدد التدريجي إلى أن الدهون الثلاثية و A-FABP هما الأكثر استقلالية في توقع حدوث متلازمة الأيض في مرضى السكري من النوع الثاني (نسبة F: 64.4 و $p < 0.001$).

الخلاصة: يرتبط مستوى المصل المرتفع للبروتين الدهني الرابط للأحماض الدهنية بشدة مع حدوث متلازمة الأيض في مرضى السكري من النوع الثاني. كما أنه مؤشر مستقل واعد لمتلازمة الأيض حيث يفيد في تشخيصها المبكر.