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Genetic Improvements in Pea (*Pisum sativum* L.) Through Irradiation by Gama Rays

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ABSTRACT

Mutation breeding using radiation has been shown to be an effective and important tool in incorporating specific desirable agronomic values characteristics in vegetable crops including pea (*Pisum sativum* L.). In the present study, dry seeds of "Master - B" Pea cultivar were exposed to 0, 10, 20 and 30 Krad doses of gamma rays, and variability in growth characteristics, earliness, and seed yield were investigated in M₁, M₂ and M₃ generations. Analysis of variance showed significant difference among M₁-plants as well as M₂ and M₃-families for the studied traits with higher variability in high doses of gamma ray treatments. In M₃-generation, lines with significant increases in yield parameters such as number of pods per plant and number of seeds per plant were obtained. Mutant line number 8 was out yielded in terms of number of pods and seeds per plant. These results indicate that induced mutation breeding by gamma ray is a valid and effective crop breeding method in pea.

Keywords: Gamma rays, genetic variability, Mutation breeding, pea, seed yield

INTRODUCTION

Pea (*Pisum sativum* L.) belongs to the family Fabaceae and is a relatively major vegetable crop in Egypt. It is one of the world's oldest agricultural products, high yield, short-term crop yield and high protein content. In dried seeds, the protein and carbohydrate content was 19.7% and 56.6% respectively.

Mutation breeding using radiation has been shown to be an important tool in incorporating specific desirable agronomic values characteristics. Gamma-ray irradiation of plant materials is commonly used to induce genetic mutation that alters the number of biochemical processes leading to beneficial genotype changes. The primary injury to plant material due to gamma irradiation is physiological damage, which is mainly limited to the M₁ generation. It can be detected and assessed cytologically as well as by the reaction of the whole organism in general by growth retardation and at higher doses of death (Sparrow, 1961). It is possible to create new variations in a short time using the mutation breeding method. The most used mutation breeding agent is physical mutagens such as ultraviolet (UV) light which leads to breaks on DNA double strand and deletions. Gamma rays and neutrons have high energy radiation applications (Koorneet, 2002). To enhance genetic variation for desirable traits, mutation breeding program can be initiated to develop stabilized mutants with respect to high yield and some other desirable traits following selection in successive advance generations (Dewanjee and Sarkar, 2017). The ease of application irradiation plays an important role on widely spreading of the technique, about 90% of the obtained mutant cultivars

were obtained with gamma-rays method, and 22% with X-rays (Jain, 2010). Reduction-induced mutation is one of the most widely used method to improve direct mutant varieties compared to acclimatization, selection, hybridization, which are laborious, time consuming and also with limited genetic variation (Hanafiah *et al.*, 2010). Any change in growth pattern will ultimately affect maturity and yield. Gamma irradiation of seeds has been shown to have a significant effect on plant growth (Rubaihayo, 1978). Radiation leads to an increase in the formation of reactive oxygen species (ROS) in plant cells due to damage to cell homeostasis and causes progressive oxidative damage and, finally, cell death (Sharma *et al.*, 2012). In M₂-generation of *P. sativum*, gamma irradiation significantly affected morphological and proximate parameters. The most important dose was 7 Krad. That can be used for agricultural purposes to improve pea yield attributes. (Khan *et al.*, 2018).

Irradiating snap bean seeds with gamma rays strongly induced morphological changes. (Eiiyfa *et al.*, 2007). Seven superior mutants were selected when exposed seeds of cowpea to three doses from gamma rays 10, 20 and 30 Krad (Metwally *et al.*, 1998). A significant variation in the protein patterns of cowpea seeds is due to the new expression of some polypeptide, the silence of others, and the expression of third-class polypeptides (Mohammed *et al.*, 2012). Gamma rays are known to affect the development and advancement of plants by affecting cytological, morphogenetic changes in cells and tissues. Several new cultivars in cowpea (Metwally *et al.*, 1998), in tomatoes (Sikder *et al.*, 2013) and mungbean (Sangsiri *et al.*, 2005) have been successfully developed by using gamma rays.

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The decline in the percentage of germination caused by the high doses of radiation applied may have resulted in a decrease in internal growth regulators (Kiong *et al.*, 2008). Gamma rays have induced reduction in plant height may be due to destructions or damage to apical meristem or reduction in the level of amylase activity and to temporary suspension of cell division or delay in mitosis or increase in the production of active radicals that are responsible for lethality or due to the increase in gross structural chromosomal changes induced by radiation (Esnault and Chenal, 2010).

The aim of this study was to induce a large scale of genetic variability by exposing Master-B pea cultivar seeds to different doses of gamma-ray in order to select new lines of pea from the superior mutant that could become a new cultivar and adapted to Egyptian environmental condition.

MATERIALS AND METHODS

The present study was carried out during the three consecutive growing seasons of 2016, 2017 and 2018 in a private farm in the Sidy Salim district, Kafr-Elsheikh Governorate and Sakha Experimental Station at Horticulture Research Institute, Kafr-Elsheikh. This work was completed in two stages, the first stage was started from November 2016 to April 2017 to induce mutation in pea (Master-B cultivar), and the second stage from November 2017 to April 2019 to characterize pea mutants produced and select desired plants in the following generations.

In the first stage, Pea seeds of Master-B cultivar were exposed to three doses of gamma-rays 10, 20 and 30 krad at a dose level of 7.69 rad / sec along with the control "0 Krad" at the Nuclear Research Center, Atomic Energy Establishment, Cairo. The source of gamma-rays was produced from Cobalt 60.

On November 1st in both seasons irradiated and unirradiated seeds (500 seeds per treatment) were sown on two sides of the ridge (0.8x10 m), 20 cm apart in a randomized complete block design with three replications. In the first season 2016, about 200 plants were chosen from each irradiated treatment which called M₁- plants. The seeds of these plants were separately harvested to generate the M₂-seeds.

M₂- seeds from the individual M-plants from each treatment were sown as a family on November 2017 season under field conditions, as well as one row of unirradiated seeds as a control. During the growth period, macro-mutations such as dwarf, vigorous plants, early or late mature plants, and other morphological changes were collected separately to give rise to M₂-families. At harvest stage, seed yield and its components were measured for each plant from each family to determine the differences of M₂ - micro mutations. Data were recorded on plant height, number of branches, pods and seed yield per plant on M₂-plants.

In the second stage, M₃-seeds from 20 selected mutants with the control (unirradiated) were sown in open field on November 2018 for further selections. According to superior in vigorous growth, pod number, juvenility period (Number of days till flowering), high yield and large seeds than the original cultivar (Master B), 11 mutants were selected. The experimental design was a complete randomized block with three replications. For collecting the

data, five plants were uprooted from each plot at the full blooming stage and the following data were recorded: plant height, number of branches/plant, number of pods/plant. In addition, number of seeds/pod and number of seeds/plant, seed index were recorded and the percentage of total protein in dry seed was determined using micro-Kjeldated method according to A.O.A.C. (1995). The total protein percent was calculated by the multiplication of nitrogen values by 6.25%

Statistical analysis

The data on M₂-Mutation were analyzed by calculating minimum value, maximum value (Range of variability) the mean treatment (\bar{X}) variance (S^2), standard error (S.E), coefficient of variation (C.V.%), the ratio of coefficient of variation of treatment and control (C.V.% of treatment/ C.V.% of control) to determine the relative variability induced by gamma rays. In addition, the data on M₁ and M₃-Mutatiowere analyzed of variance according to Snedecor and Cochran (1971).

RESULTS AND DISCUSSION

Effect of gamma-rays on seed germination and M₁-plants

The results presented in Table (1) revealed that increasing rate of gamma ray from 0 to 30krad significantly decreased the percentage of seed germination. The maximum germination percent (90.13%) was observed with the unirradiated seeds (control) compared with irradiated seeds. The lowest germination percent (11.8%) was obtained with the seeds treated by gamma ray at rate 30 krad. The effect of mutagens on seeds is expressed through delayed emergence of roots, reduction in vigor, low metabolic and enzymatic activity, losses of membrane integrity and finally loss of germ inability, suggesting that germination is a radio sensitive phenomenon (Sushil and Dubey, 1997). These results are in agreement with those of Katayani *et al.* (1980) observed the drastic reduction in the percentage of seed germination under higher doses of gamma irradiation in *Phaseolusaureus*.

Regarding to plant height, results presented in Table (1) showed that increasing irradiation doses caused gradual decrease in plant height; where the control treatment gave the highest value in this respect as compared with the lowest values obtained from the high doses of gamma ray treatments. Similarly, Ramesh and Reddi (2002) reported that irradiation by gamma ray decreased plant height of three cultivars of *Oryza sativa* L.. This may be due to destruction or damage happen to apical meristem when high doses of gamma rays used (Patel and Saha, 1974), hampered respiratory enzyme synthesis and reduction in the level of amylase activity (Reddy and Vidyavathi, 1985), or temporary suspension of cell division or delay in mitosis. In the same line, Machaiah and Vakil (1979) mentioned that ionizing radiation causes inactivation of growth regulators leading to retarded plant growth. In addition, Selim *et al.* (1974) obtained that reductions in plant height may be due to an increase in the production of active radicals due to increase radiation doses.

The effects of gamma doses on number of branches and leaves/plant, fresh weight / plant and number of pods / plant were given in Table 1. The results showed that there were no significant differences among the radiation doses. Furthermore, plant dry weight was decreased with

increasing of radiation doses from 0 to 30 krad. The highest dry weight was obtained by the control plants and radiation dose at 10 krad without significant differences. Radiation treatment dose of 10krad significantly increased number of seed per plant followed by control, radiation at 30 and 20krad. The decreasing in plant dry weight could be attributed to reduction plant height, branches and leaf number (Amjad *et al.*, 1993). In addition, radiation lead to increase the formation Reactive oxygen species (ROS) in

plant tell due to damage of cellular homeostasis and cause progressive oxidative damage and finally cell death (Sharma *et al.*, 2012). Bajaj (1970) reported that gamma irradiations causes problems with RNA and protein synthesis and then inhibits growth of plants. The reduction in seed yield and its components with increasing the doses may be due to the reduction in fertility which may be attributed to chromosomal aberrations or attributed to physiological damage (Abd El-Rahman, 2000).

Table 1. Effect of gamma irradiation on growth characteristic and yield of Master B pea cultivar and M₁-plants during 2016 season

Radiation doses(KR)	Germination (%)	Plant height (cm)	No. of branches/Plant	No. of leaves /plant	Plant fresh weight (g)	Plant dry weight (g)	No. of pods/plant	No. of seeds/plant
Control	90.1 a	30.00 a	0.40	9.40	11.70	2.21 a	5.60	29.70 ab
10	76.3 b	25.40 ab	0.30	9.00	10.60	1.80 a	7.10	32.90 a
20	43.7 c	14.60 bc	0.10	7.00	5.40	0.99 ab	4.50	16.00 b
30	11.8 d	3.90 c	0.00	4.40	2.60	0.39 b	5.00	19.90 ab
L.S.Dat5%	11.98	12.19	1.12	5.85	9.36	1.26	3.54	15.57

Means followed by different letters within column are different by Duncan's multiple range test in 0.05 P-significance.

2-Effect of gamma irradiation on M₂-generation

Results in Table 2 show that high doses of gamma rays treatments were significantly induced genetic variability higher than control. The measurements of variation (rang of variability and coefficient of variation) in the treated plants were higher than those of the control. Therefore, the increments in variability in M₂- irradiated generation could be due to genetically variations affecting the plant height (Hussein *et al.* 1974), suggesting were opportunities for the selection of the desirable plant height.

Data presented in Table 3 show that the estimates for number of branches per plant under doses of gamma rays was not statistically variable in spite of the appearance of some differences among the treatments means for this trait. This response for number of branches per plant to radiation might be due to the balance between the stimulating effect

of the lower doses of mutagen and inhibiting effect of the higher doses (Badr *et al.*, 2000). Furthermore, the depression effect of the high mutagen doses on vegetative traits maybe attributed to the active disturbances of some enzyme involved in the synthesis of the growth (Abd El-Rahman, 2000). Data presented in Table 3 showed that the number of pods per of pea plant cultivar Master B was affected by the different mutagenic treatments in M₂-generation. The response of seed yield per plant to mutagenic treatments differed in the cultivar Master after irradiated with gamma rays. The highest percentage of mutants in the M₂-generation was obtained and selected 11 mutants with more seeds per pod, larger seeds and plant height than initial form (Khan *et al.* 2018) on pea reported that gamma irradiation significantly affected morphological and proximate and yield parameters in M₂- generation.

Table 2. Effect of gamma irradiation in M₂- generation on plant height and number of branches of pea plants cv."Master B" during 2017 season

Radiation doses(KR)	Number of plants	Plant height (cm)				No. of branches					
		Min.	Max.	$\bar{x} \pm S.E$	C.V. %	C. V. treat		C. V. Cont		C. V. Cont	
Control	450	45	63	56.10 ± 5.57	17.21	1.00	1.20	2.10	1.80 ± 0.33	17.21	1.00
10	381	52	62	57.00 ± 3.00	9.11	0.53	1.00	2.00	1.60 ± 0.30	33.23	1.93
20	217	57	84	67.70 ± 8.33	21.31	1.23	2.00	2.50	2.30 ± 0.14	11.16	0.65
30	59	51	85	65.80 ± 10.13	26.77	1.55	0.80	2.50	1.60 ± 0.49	55.17	3.20

Table 3. Effect of gamma irradiation in M₂- generation on number of pods and seeds of pea plants cv."Master B" during 2017 season

Radiation Doses (Krad)	Number of plants	Number of pods/plant					Number of seeds/plant				
		Min	Max	$\bar{x} \pm S.E$	C.V.%	C. V. treat		C. V. Cont		C. V. Cont	
Control	450	9.00	19.00	14.10 ± 2.98	36.84	1.00	42.00	52.00	48.30 ± 3.40	12.19	1.00
10	381	9.00	14.00	12.10 ± 1.48	21.31	0.58	34.00	68.00	57.00 ± 11.44	34.75	2.85
20	217	12.00	24.00	16.70 ± 0.99	41.36	1.12	51.00	108.00	73.40 ± 17.83	42.07	3.45
30	59	6.00	33.00	17.20 ± 8.23	82.85	2.24	25.00	91.00	57.20 ± 18.94	57.42	4.71

3- Effect of gamma rays on M₃-generation

In M₃-generation, 11 lines were selected based on superiority in vegetative growth and flowering traits, and high yield with large seeds than the original cultivar (Master B). The results presented in Table 4 showed that there were significant differences between the mutant lines for plant height and number of branches. The tallest line was obtained by Mutant line 10, while the shortest one obtained with line 4 compared with original cultivar (control). In addition, mutant line 8 had the higher number of branches per plant than the original cultivar. On the other hand, mutant line 6 had the lowest number of branches per plant.

It is obvious from Table 4 that there were significant differences in number of days till flowering among M₃-lines selected and the original cultivar (Master B). Mutant line 3 had the shortest days to flowering compared to original cultivar with 9 days differences. Significant differences among 11 mutant lines and original cultivar were also observed for seed yield and its components (number of pods

per plants, number of seeds per pod and seed index) as indicated in Table 4. In general, most of mutant lines were produced more pods and heavy seeds than the original cultivar, while the behavior of number of seeds per pod differed among the mutants and the original cultivar. Mutant line number 8 was produced the highest number of pods per plant. Furthermore, mutants lines 5, 8, 9, and 11 were produced the highest number of seeds per plant. In the same line, Metwall *et al.* (1992) found that gamma rays was increased significantly seed yield of cowpea plants which were expressed by number of pods per plant, number of seeds per pod as well as the value of seed index.

Regarding to seed protein percentage, data in Table 4 indicate that one mutant line No. 3 contain higher values than the original cultivar. On the other hand, mutant line No. 7 was decreased significantly in their protein content. Khan *et al.* (2018) found that irradiated pea in M₂ generation reported was varied in seed protein.

Table 4. Growth characteristic, flowering date, seeds yield and components of 11 mutants of pea comparing with cv. "Master B" during 2018 season.

Number of line	Plant height (cm)	No. of branches/plant	No. of days till flowering	No. of pods /plant	No. of seeds /pod	No. of seeds /plant	Seed index	Protein % in seeds
Control	63.80 g	1.50 cde	38 a	13.80 c	9.50 a	78.00 bc	14.90 g	19.89 ab
1	92.80 e	1.00 de	38 a	14.30 c	7.00 cd	79.30 bc	17.20 e	15.07 d
2	78.80 f	2.80 ab	38 a	20.80 bc	8.30 b	96.80 abc	18.80 c	18.61 bc
3	97.50 e	2.30 abc	47 d	20.30 bc	6.00 d	71.00 c	20.40 a	21.40 a
4	38.00 h	1.50 cde	38 a	6.00 d	7.80 bc	30.00 d	14.10 h	19.20 bc
5	103.00 de	2.00 bcd	44 b	19.00 bc	7.00 cd	114.50 a	18.70 c	19.64 abc
6	118.00 bc	0.80 e	43 c	18.30 bc	8.00 bc	93.80 abc	18.90 c	18.47 bc
7	129.30 ab	1.00 de	43 c	22.80 b	7.50 bc	107.80 ab	18.00 d	17.61 c
8	102.30 de	3.30 a	43 c	29.30 a	7.30 bc	128.30 a	19.60 b	18.80 bc
9	112.00 cd	1.00 de	43 c	17.00 bc	7.80 bc	118.30 a	17.10 e	18.19 bc
10	136.00 a	0.80 e	43 c	18.30 bc	7.50 bc	104.30 abc	16.00 f	19.75 ab
11	60.30 g	2.50 abc	43 c	17.00 bc	9.50 a	122.50 a	16.80 e	19.74 ab
L.S.Dat5%	12.07	1.002	0.004	6.13	0.97	30.79	0.57	1.81

Means followed by different letters within column are different by Duncan's multiple range test in 0.05 P-significance.

CONCLUSION

Induced mutation is a valuable tool for inducing new genetic variability in vegetable crops including pea. In the present study, genetic variability was observed for all morphological and yield measured characters in M₁-plants and M₂ generation, and new mutant families of pea with novel morphological traits and higher seed yield were selected in M₃-generation. These elite mutant families could be used in the future breeding program in pea for development of new cultivars adapted for Egyptian condition.

REFERENCES

- A. O. A. C. (1995). Association of official Analytical Chemists. Official Methods of Analysis. 15th Ed. Washington D. C., USA.
- Abd El-Rahman, M. M. (2000). Inducing genetic variability in *Citrullus colocynthis* by using gamma irradiation. J. Agric. Sci. Res. Mansoura Univ., 25(1): 193-1991.
- Amjad, M.; R. Rahim; A. Ali and M. Ayyub (1993). Variation in growth characteristics of pea under the Influence of gamma irradiation. Pak. J. Agri, Sci., 30 (1): 102-104.

- Badr, M.; O. El-Shennawy; M. Mostafa and F. EL-Tony (2000). Effect of gamma irradiation, Methyl methane sulphonate and their combination on growth flowering and induced variability in *Tagetes erecta* L. J. Agric. Sci. Mansoura Univ., 25(6): 3587 – 3604.
- Bajaj Y, P. S. (1970). Effect of gamma irradiation on growth, RNA, protein, and nitrogen content of bean callus cultures [https:// academic.oup.com /aob articles abstract 34/5/1989/148656](https://academic.oup.com/aob/articles-abstract/34/5/1989/148656).
- Dewanjee, S. and K. K. Sarkar (2017). Evaluation of Performance of Induced Mutants in Mungbean [*Vigna radiata* (L.) Wilczek]. ISSN: 0250-5371 / Online ISSN: 0976-0571
- Eiyyfa, K.; O. M. Ahmed; S. Shaharudin and D. AbdulRahman (2007). Gamma radio sensitivity study on snap bean (*Phaseolus vulgaris*) J. Agris Reas., 2(9): 844-848.
- Esnault, M. A and C. Chenal (2010). Ionizing radiation advances in plant response. Environmental and Experimental Botany, 68:231- 237.

- Hanafiah, D. S.; T. Maningtyas; S. Yahya and D. Wirnas (2010). Induced mutations by gamma ray irradiation to Agronomy bean (*Glycine max*) variety. *Nusantara Bioscience*, 2(3): 121-125.
- Hussein, H. A. S., A. R. Selim and I. I. S. EL-Shawaf (1974). EMS and gamma-rays induced mutations in *Pisum sativum* L. I. Effect on the frequency and spectrum of M₂ chlorophyll mutations. *Egypt J. Genet. Cytol.*, 3(1) 106-116.
- Jain, S. M. (2010) Mutagenesis in crop improvement under the climate change. *Romanian Biotechnological Letters*, 15(2); 88-106.
- Katyayani, M.; D. Rao; S. Raos and K. Murthy (1980). Gamma irradiation induced physiological variabilities in *Phaseolus aureus*. *Roxb. J. Indian Bot. Soc.*, 59: 153-156.
- Khan, W.; M. Zahir and N. Akhtar (2018). Gamma radiation induced mutation in M₂ generation of pea (*Pisum sativum*) applied Biology, 7(2): 832-837.
- Kiong, A.; A. Lingpick; S. H. Grace Lai and A. R. Harun (2008). physiological responses of Orthasiphonstamineus plantlets to gamma irradiation. *American Eurasian Journal of Sustainable Agriculture*, 2: 135-149.
- Koornneet, M. (2002). Gene identification classical mutagenesis in higher plants, in Philip M.
- Machaiyah, J. D. and U. K. Vakil (1979). The effect of gamma irradiation in the formation of alpha amylase isoenzyme in germinating wheat. *Envir. Exp. Bot.*, 19: 337-348.
- Metwally, E. I.; A. A. Ali and A.Y. Mazrouh (1992). Selection of high yielding mutant of cowpea (*Vignasinesis* L.) after irradiations. *Annals of Agric. Sci. Moshtohor*, 3(1): 221-231.
- Metwally, E. I.; A.M. Hewedy; M. Hafez and M. A. Morsy (1998). Kafr El-Sheikh I and Kahal new cultivars of Cowpea, *J. Agric. Sci. Mansoura Univ.* 23(8):3887-3897.
- Mohammed, A. H.; H. I. Mohammed; B. M. Zaki and A. M. Mogazy (2012). Pre-exposure to gamma rays alleviated the harmful effect of salinity on Cowpea plants. *Journal of Stress Physiology and Biochemistry*, 8(4): 199-2171.
- Patel, J. D. and J. J. Saha (1974). Effect of gamma irradiation on seed germination and organization of shoot apex in *Solanum melongena* and *Capsicum annum*. *Phytomorphology*, 25:174 - 180
- Ramesh, D. V. and T. V. V. S. Reddi (2002). Induced morphological variability among three genotypes of rice *J. Cyto. Genet.*, 3: 115-120.
- Reddy, K. J. and M. Vidyavathi (1985). Effects of sumithion on the germination, growth chromosomal aberration and the enzyme amylase of *Dolichos bifloras* L. *Indian bot. Soc.*, 64 : 88-92.
- Rubaihayo, P. R. (1978). Utilization of gamma-rays for soybean improvements, *Egypt. J. Genet. Cytol.*, 5(1): 136-140.
- Sangsiri, C.; W. Sorajjapinun and P. Srinives (2005). Gamma Radiation Induced Mutations in Mungbean. *Sci. Asia*. 31: 251-255.
- Selim, A. P.; H. A. Hussein and I. I. S. Kishawaf (1974). EMS and gamma-rays induced mutation in *Pisumsativum* L. II. Effects of gamma rays on M1-generation seedling bight and fertility. *Egypt J. Genet. Cytol.*, 3: 170.192.
- Sendecor, G. W. and W. G. Cochran (1971). *Statistical Methods* Iowa State University Press, U.S.A.
- Sharma, P.; A. B. Tho; R. S. and M. D. Pessaraki (2012). Reactive oxygen species, oxidative damage and antioxidant defense mechanism in plant under stressful conditions, *Journal of Botany*, 1.26.
- Sikder, S.; P. Biswas; P. Hazra; S. Akhtar; A. Chattopadhyay; A. M. Badigannavar and S. F. D. Souza (2013). Induction of mutation in tomato (*Solanum lycopersicum* L.) by gamma irradiation and EMS. *Indian Journal of Genetics and Plant Breeding*, 73(4), 392-399.
- Sparrow, A. H. (1961). Types of ionizing radiations and their cytogenetic effects. *Mutation and Plant Breeding*, NAS-NRC., 891: 55-119
- Sushil, K. and D. K. Dubey (1997). Effect of mutagens on pollen traits in M₁ and their possible role as indicators of micro mutations in latter generations in Khesari (*Lathyrus sativus* L.) var. P-sos. *J. Indian Bot. Soc.*, 75:89-89.

دراسة الاختلافات الوراثية في البسلة باستخدام اشعة جاما على ابراهيم على مصرى ، عاطف محمد السيد فياض و داليا ابراهيم طاهر معهد بحوث البساتين – مركز البحوث الزراعية – الجيزة – مصر

يعتبر استخدام الطفرات عن طريق الاشعاع من اهم الطرق الفعالة والهامة في التحسين الوراثي في محاصيل الخضر وخاصة المحاصيل ذاتية التلقيح مثل البسلة. لذلك اجريت الدراسة خلال ثلاثة مواسم متتالية ابتداء من 2017 حتى 2019 على نباتات البسلة صنف ماستر بي المنزرعة بمزرعة خاصة بمحافظة كفر الشيخ – مصر. وذلك بهدف استحداث اختلافات وراثية يمكن استخدامها في تحسين محصول البسلة. لذا تم استخدام جرعات مختلفة من اشعة جاما (0, 10, 20, 30) كيلو راد على خصائص النمو المختلفة والتكبير والمحصول ومكوناته بالإضافة الى الاختلافات الوراثية على الجيل الاول والثاني والثالث (M₁, M₂, M₃) اظهرت النتائج وجود فروق معنوية بين نباتات الجيل الاول من الناحية المورفولوجية بالإضافة الى وجود اختلافات وراثية بين نباتات الجيل الثاني والثالث وخاصة مع زيادة جرعة الاشعاع (30 كيلو راد). كما اوضحت النتائج وجود زيادة معنوية في سلالات الجيل الثالث المنتخبة وخاصة السلالة رقم 8 في خصائص المحصول مثل (عدد القروون / نبات, عدد البذور / نبات). لذلك توصي الدراسة الى ان استخدام اشعة جاما من افضل الطرق الفعالة في تحسين البسلة.