

---

## EARLY DETECTION OF CANDIDA ALBICANS IN NEONATAL INTENSIVE CARE UNITS

**Ragaa A.M. Ibraheim \*; Salah A. I. Ali\*\*; Hanaa Hosney\*\*  
and Hannan S. Ahmad \*\***

Pediatric department, Al-Azhar University for Girls\*  
Clinical pathology department, Zagazig University \*\*

### **ABSTRACT**

**Background and objective:** Candidemia is a one of the most common causes of late-onset sepsis in the neonatal intensive care unit (NICU) and is associated with significant morbidity and mortality. The aim of this work is to evaluate the value of Real-Time PCR for early detection of blood stream *Candida albicans* infection (candidemia) in NICU. **Materials and Methods:** The study included 60 newborn admitted in NICU of Al-Zhraa hospital, Al-Azhar University Hospital for girls, Egypt, during the period from January 2010 to January 2011, who had one or more risk factors for candidemia. A total of 60 blood samples were submitted to testing by blood culture and real-time PCR. 20 cases were diagnosed as candidemia by blood culture method, the other 40 neonates were negative for candidemia by blood culture method considered as control group. **Results:** the blood culture was positive in 20/60 samples and *Candida albicans* was the predominant cause of candidemia 15/20 (75 %) followed by *C. tropicalis* 4/20 (20 %) and *C. glabrata* 1/20 (5 %). Real-time PCR for *Candida albicans* was positive in 14/15 and gave only one false-positive and false-negative result. It had a sensitivity, specificity and accuracy of 93.3 %, 97.8 % and 96.7% respectively. Central venous catheter (CVC) was the most frequent risk factor among candidemic cases 15/20 (75 %), followed by mechanical ventilation (MV), endotracheal tube (ETT) and admission > 10 days in NICU (45 %) for each. Out of 20 patients, 17 had more than one risk factor (Odd Ratio [OR] 22.7; 95%CI 5.31, 96.77;  $p < 0.0001$ ). **Conclusion:** Real-Time PCR can be applied as a rapid diagnostic method for candidemia and can aid in early antifungal therapy and in minimizing the several drawbacks of candidemia. Identification of risk factors is more important in predicting the neonates who are at risk for candidemia.

### **INTRODUCTION**

Candidemia is the fourth most common nosocomial bloodstream infection [1]. Neonatal candidemia accounts for 12% to 15% of late-onset sepsis episodes in the NICU

[2]. Neonates diagnosed with candidiasis are at significant risk for mortality (20%) and morbidity [3, 4, 5]. Pediatric patients in critical condition, particularly neonates, are especially vulnerable

to invasive *Candida* infections (ICI), partly because of their age and severe underlying disease and partly because of the invasive procedures used [6]. Risk factors such as preterm babies with low body weight, staying for long-term periods at intensive care units, subjected to invasive procedures, receiving one or more antibiotic schemes are considered to pose a risk for the development of infections by opportunistic agents [7]. Although *C. albicans* remains the most common fungal pathogen isolated from blood and body tissue, recent literature shows an increased prevalence of non-candida species [8, 9, 10]. The diagnosis of invasive candidiasis is difficult because there are no specific clinical manifestations of the disease and colonization and infection are difficult to distinguish. In the last decade, much effort has been made to develop reliable tests for rapid diagnosis but none of them found widespread clinical use [11]. Traditional diagnostic methods based on culture techniques and biochemical tests require 2 or more days and could be inaccurate [12], although a blood culture positive for candida was considered the gold standard for diagnosis of candidemia [13]. A variety of PCR assays based on the detection of fungal DNA in

sterile human body fluids or tissue samples to allow early diagnosis of fungal infections and to improve the survival rate of patients suffering from invasive infections has been described. However, none of the developed PCR assays have been standardized, resulting in diverging results [13]. The introduction of real-time PCR technology in the detection of fungal infections has increased reliability of PCR results compared to results obtained by conventional PCR methods. Real-time PCR sharply decreases the risk of false-positive results due to PCR product carry-over during gel electrophoresis or enzyme linked immunoassays to check the specificity of the conventional PCR product [14]. The aim of this work is to evaluate the value of risk factors and real-time PCR for early detection of candidemia in NICU patients before development of clinical sepsis.

### **MATERIALS AND METHODS**

Clinical samples were obtained from Sixty newborn admitted in NICU of Al-Zhraa hospital, Al-Azhar University Hospital for girls, Egypt, during the period from January 2010 to January 2011, who had one or more of the risk factors for candidemia. All samples were evaluated by blood culture and real-time PCR.

Approximately 2.5 mL of fresh blood obtained from patients was inoculated aseptically into a blood culture bottle containing 25 mL of Sabouraud's broth (Sigma-Aldrich Chemie GmbH) and incubated at 37°C for 14 days maximum. After 24 hours incubation subculture was done on: Sabouraud's dextrose agar (SDA) with chloramphenicol (Oxoid, UK): plates were incubated aerobically at 37°C for 48 hours. All plates were examined after the given periods for any growth. If no growth was found, subcultures were done every three days for three successive times and if still no growth was seen, cultured plates were considered negative and excluded. Phenotypic identification of *Candida* species was based on germ tube assessment and CHROMagar *Candida* (Hardy Diagnostics Inc.).

#### **Detection of *Candida albicans* by quantitative Real-time PCR:**

The clinical specimens were tested using *Candida albicans*-specific real-time PCR assay. The amplified PCR products were identified with Light Cycler SYBR Green I dye fluorescent based on LightCycler FastStart DNA MasterPLUS SYBR Green I (Roche).

**DNA extraction:** DNA was extracted from 200 µL of serum by

MagNA Pure Compact Nucleic Acid Isolation Kit I. using MagNA Pure Compact instrument for automated purification of DNA [15]. DNA extraction from a clinical sample is a critical step for molecular diagnosis. DNA extraction in the present study was done by an automated system using MagNA Pure Compact nucleic acid isolation kit I with the MagNA Pure Compact instrument. Total time for the automated purification of DNA from 8 samples is about 25 minutes [15]. Automated extraction systems have the advantage over manual extraction in that recovery of nucleic acids is consistent and reproducible, also it keep sample manipulation to a minimum and hence reducing the risk for cross contamination. Finally, once these systems have been validated and proper maintenance procedures are in place, quality control monitoring is less intensive than that required for manual extraction [16].

#### ***Candida albicans*-specific primers:**

Primers for *C. albicans* are specific for the ITS (Internal Transcribed Spacer) of the fungus, with amplicon size 273 bp and melting temperature of 85.7±0.7 °C: Forward: (5'→3') TTTATCAAC TTGTACACCAGA, gene bank accession no. L47111, Reverse:

(5'→3') ATCCCGCCTT  
ACCACTACCG, gene bank  
accession no. L28817 [17].

**PCR Protocol:** The *C. albicans* DNA extracted from serum samples was subjected to a consensus primer mediated PCR method using the LightCycler FastStart DNA MasterPLUS SYBR Green I (Roche). The standard PCRs were carried out in 20 ul. The samples were cycled according to the LightCycler protocol. There was an activation step of Taq polymerase at 95 °C for 10 minutes, followed by 45 cycles of denaturation at 95 °C for 10 s, annealing at 65 °C for 5 s and extension at 72 °C for 10 s. This was followed by melting analysis consisting of 95 °C for 30 s. and then cooling to 35 °C for 60 s. before temperature was raised to 98 °C at a rate of 0.2 °C / s. with continuous fluorescence acquisition. A final cooling step was performed at 40 °C for 10 s [17].

Results were interpreted as positive or negative by the presence or absence of the melting temperature peak of  $85.7 \pm 0.7$  °C and by the effectiveness of quantitative score where the data was saved and processed by the software of the analyzer both can increase the specificity and sensitivity of amplification reactions. The real-time PCR assay analy-

tical detection threshold was 200 copies of *C. albicans* amplicon / ml.

### **STATISTICAL ANALYSIS**

Analysis of the collected data was done using SPSS version 11.0. Data were presented as mean  $\pm$  Standard deviations for continuous variables. For qualitative variables, frequency data were summarized as number and percentage. Chi square ( $X^2$ ), Odd Ratio [OR] and 95% confidence intervals (CI) were applied for comparison of qualitative data. p value considered significant if  $<0.05$ . Sensitivity (true positive / true positive + false negative), specificity (true negative / true negative + false positive), positive predictive value (true positive / true positive + false positive), negative predictive value (true negative / true negative + false negative) and accuracy (true positive + true negative / true positive + false positive + false negative + true negative).

### **RESULTS**

Samples were obtained from 60 neonatal patients at risk for candidemia, 29 were males and 31 were females. A total of 60 samples were analyzed by blood culture and real-time PCR assays. Out of these, 20 showed positive blood cultures. Table 1 shows: the demographic and the distribution

## EARLY DETECTION OF CANDIDA ALBICANS IN NEONATAL INTENSIVE CARE UNITS

Ragaa A.M. Ibraheim; Salah A. I. Ali; Hanaa Hosney and Hannan S. Ahmad

of risk factors in cases and control. Table 2 shows: The risk factors within this population of patients with candidemia. All patients had at least one predisposing risk factor. The most common risk factor was the use of central venous catheter (75%), followed by mechanical ventilation, endotracheal tube, Admission in NICU > 10 days (45%) for each. Table 3 shows: Most of the patients had multiple risk factors. Out of the 20 patients, 12 had three different risk factors and 5

had four risk factors. Only, 3 cases had one risk factor. Table 4 shows: The distribution of Candida species among the blood culture positive patients. Candida albicans, Candida tropicalis and Candida glabrata caused 75%, 20%, 5% of the candidemia respectively. Table 5 shows: Real-time PCR was positive in 14 samples and gave only one false-positive and false-negative result, its diagnostic indices are detailed in Table 5.

**Table (1): Demographic characteristics and risk factors in cases and control.**

Variables	Cases (No = 20)		Control (No = 40)	
	No	%	No	%
CVC	15	75.0%	11	27.5%
MV	9	45.0%	8	20.0%
ETT	8	40.0%	6	15.0%
Postnatal Steroids	4	20.0%	8	20.0%
Antibiotics	4	20.0%	8	20.0%
Admission > 10 days	9	45.0%	3	7.5%
GA < 32 weeks	5	25.0%	2	5.0%
VLBW <1500 gram	5	25.0%	2	5.0%
Multiple risk factors	17	85.0%	8	20.0%
Males	5	25.0%	24	60.0%
Age in days (on admission)	3.95±2.39		4.83 ±2.34	

CVC; Central venous catheter, MV; Mechanical ventilation, ETT; Endotracheal tube, VLBW; Very low birth weight, GA: gestational age.

**Table (2): The risk factors for predicting cases of candidemia in NICU patients..**

Variables	Candidemia (N=20)		P-value	OR	95% Confidence Interval	
	N	%			Lower	Upper
CVC	15	75	0.00	7.91	2.32	26.98
MV	9	45	0.04	3.27	1.01	10.57
ETT	8	40	0.04	3.78	1.09	13.13
Postnatal Steroid	4	20	0.64	1.00	0.26	3.82
Antibiotics	4	20	0.54	1.18	0.30	4.61
Admission >10 days	9	45	0.001	10.09	2.32	43.88
GA < 32 weeks	5	25	0.04	6.33	1.11	36.27
VLBW < 1500 grams	5	25	0.04	6.33	1.11	36.27

CVC; Central venous catheter, MV; Mechanical ventilation, ETT; Endotracheal tube, VLBW; Very low birth weight, GA: gestational age, OR; Odd ratio, *p* significant if < 0.5.

**Table (3): Multiple risk factors for predicting cases of candidemia in NICU patients.**

Multiple risk factors	Candidemia (N=20)		P	OR	95% Confidence Interval	
	N	%			Lower	Upper
CVC, MV and ETT	8	40	<0.0001	22.7	5.31	96.77
CVC, VLBW, GA< 32w and Admission > 10 days	5	25				
Steroids, Antibiotics and Admission > 10 days	4	20				
Total	17	85				

CVC; Central venous catheter, MV; Mechanical ventilation, ETT; Endotracheal tube, VLBW; Very low birth weight, GA: gestational age, OR; Odd ratio, *p* significant if < 0.5.

**Table (4): Distribution of various Candida species isolated from blood in cases of neonatal candidemia.**

Species	Blood culture	
	No	%
<i>C. albicans</i>	15	75.0 %
<i>C. tropicalis</i>	4	20.0 %
<i>C. glabrata</i>	1	5.0 %
Total	20	100.0 %

**Table (5): Sensitivity and Specificity of real-time PCR for diagnosis of Candida albicans among studied groups.**

Test	Culture		Sensitivity	Specificity	Predictive value		Accuracy
	+ve	-ve			+ve	-ve	
PCR	+ve	14	93.8 %	97.8%	93.9%	97.8%	96.7 %
	-ve	1					

PCR; Polymerase chain reaction, +ve; Positive, -ve; Negative

## DISCUSSION

The present study emphasizes the importance of candidemia among neonatal intensive care unit (NICU). Candidemia is not only associated with a significant mortality but also extends the duration of hospital stay and increases the cost of medical care [18]. All these drawbacks are related to many factors among

them; most of clinicians do not pay attention to risk factors till clinical sepsis develop or even after that. A septic-appearing newborn routinely receives antibacterial agents at the time of clinical presentation. In contrast, many clinicians will only start antifungal therapy when a positive culture is reported or when the patient fails to improve after a

period of antibacterial agents. On the other hand, the routine blood cultures for candidal identification up to species level and susceptibility till reporting results take also a long time that can aggravate these drawbacks. This study, aimed to evaluate the value of risk factors and real-time PCR in early diagnosis of candidemia before appearing of clinical sepsis.

Numerous studies have identified risk factors, among them are: receiving broad-spectrum antibiotic therapy and steroids [19, 20]. □ Infants with positive or negative culture showed no difference between receiving broad-spectrum antibiotic therapy ( $p = 0.54$ ), and steroids ( $p = 0.64$ ) (table 1).

Our Statistical analysis revealed the following significant risk factors associated with *Candida* spp. isolation: central intravascular catheter (CVC), mechanical ventilation (MV), endotracheal intubation (ETT), prolonged hospitalization, VLBW and gestational age (GA) < 32 ( $p < 0.05$ ) (table 2). All of these extensively reported as major factors for candidemia. [19, 21-23]. □ CVC was the most frequent risk factor in the positive blood culture cases (75%). It was the only risk factor in two positive cases, while was associated with other risk factors in 13 cases (table

3). We observed that most of patients with positive blood cultures had multiple risk factors ( $P < 0.001$ , OR =22.7, 95% CI =5.31 – 96.77) (table 3). Several other studies have illustrated the link between the presence of multiple risk factors (e.g. duration of intubation and length of NICU hospitalization or antibiotics and steroids) and emergence of fungal sepsis [19, 24- 26].

*Candida albicans* was the most common isolate (75%) as reported by others [27]. Over the years, non-*albicans* *Candida* infections have become more frequent as major causes of neonatal candidemia [28 - 30]. *Candida parapsilosis* was reported to be most common isolate in several studies [23, 31- 33]. Our data shows *Candida tropicalis* to be the second most common agent (20%). Mari'a t al., reported that *Candida tropicalis* represented 10% of patients with candidemia in his study, which may be acquired by horizontal transmission [34]. Its ability to produce clusters has been one of its major pathogenic components. However, *C. tropicalis* has not been thoroughly studied and could potentially play an important role in nosocomial infections [35, 36]. Our data revealed that *Candida glabrata* was the least cause of

neonatal candidemia (5%). Although it was considered to be less pathogenic but studies have shown no significant differences in morbidity among species [37].

The technique used in this study was the real-Time PCR for *Candida albicans*. So, the comparison was applied according to whether the case was positive or negative for *Candida albicans* by blood culture while other *Candida* species were not included to make this comparison possible.

Several PCR assays have been developed for the diagnosis of candidemia, in this study, we evaluated the efficiency of real-time PCR for the diagnosis of *Candida albicans* as a cause of candidemia in NICU patients at high-risk for candidemia. Real-Time PCR assay was directly performed from serum samples obtained from these patients. To date, the optimal biological specimen for the molecular detection of candidemia is a matter of debate. Some authors have used serum [38, 39 ], whole blood samples [40, 41] or blood culture samples [42] for the detection and identification of *Candida* spp. for diagnosis of candidemia.

The real-time PCR assay identified 14 of 15 positive specimens detected by blood culture with one

false-negative and one false-positive results. Negative PCR in ultimately culture-positive samples represents a serious problem in making a diagnosis and may return to a deficient DNA extraction, small amounts of target DNA in the sample, as well as the small amount of sample submitted to PCR, or PCR inhibitors. The melting curve analysis designed in this study detected *Candida albicans* DNA from 14 patients with a positive blood culture and gave only one false-positive result when compared with the reference standard, blood culture with a sensitivity, specificity, PPV, NPV and accuracy of 93.9% 97.8% 93.9% 97.8% and 96.7% respectively. It should also be noted that the inexpensive blood culture system used in this study may have a lower sensitivity than widely used automated culture systems moreover, the ability of the automated system (MagNA Pure Compact) used for extraction to recover minute amounts of DNA in an efficient manner may thus preferentially inflate the false-positive rate or represented true false-positive results caused by PCR contamination.

Lack of standardization in the molecular diagnosis of candidemia made many discrepancies in sensitivity and specificity of several studies [27]. Our results



were matched with some studies [43- 45] but did not match with others [13, 42, 46- 48]. In general, the introduction of real-time PCR technology together with automated extraction systems in the detection of fungal infections have increased the reliability of PCR results compared to results obtained by conventional PCR methods. The identification of species via melting curve analysis with species-specific hybridization probes further increases specificity, as one mismatch in the probe binding site would lead to an altered melting temperature ( $T_m$ ). The fast turnaround time of less than 2 h is another advantage of the real-time PCR technology [49].

Our study was focused on *Candida albicans* because it was the most common pathogen isolated during routine work as processed by others [50]. The differentiation between clinically relevant *Candida* species is of great value in guiding antifungal therapy [51]. Van Burik et al. and White et al. described panfungal PCR assays that can only detect the genus *Candida* [52, 53]. Recently, Schabereiter-Gurtner et al. [54] described a Light Cycler assay that allows the simultaneous detection and identification of various *Candida* and *Aspergillus*

species; its clinical efficiency needs to be assessed in prospective studies.

### CONCLUSION

In conclusion: Real-Time PCR can be applied as a rapid diagnostic method for candidemia and can aid in early antifungal therapy and in minimizing the several drawbacks of candidemia. Identification of risk factors is more important in predicting the neonates who are at risk for candidemia.

### REFERENCES

1. Christopher C. Blyth, Sharon C. A. Chen, Monica A. Slavin, Carol Serena, Quoc Nguyen, Deborah Marriott, David Ellis, Wieland Meyer, Tania C. Sorrell (2009): Not Just Little Adults: Candidemia Epidemiology Molecular Characterization, and Antifungal Susceptibility in Neonatal and Pediatric Patients *Pediatrics* Vol. 123 No. 5 May 1, pp. 1360 -1368.
2. Feja KN, Wu F, Roberts K, et al. (2005); Risk factors for candidemia in critically ill infants: a matched case-control study. *J Pediatr.* 147:156-161.
3. Benjamin DK, Jr, Stoll BJ, Fanaroff AA, et al. (2006): Neonatal candidiasis among extremely low birth weight infants: risk factors, mortality rates, and neurodevelopmental outcomes at 18 to 22 months. *Pediatrics.* 117:84-92.
4. Benjamin DK, DeLong E, Cotten CM, et al. (2004): Mortality

- following blood culture in premature infants: increased with Gram-negative bacteremia and candidemia, but not Gram-positive bacteremia. *J Perinatol.* 24:175–180.
5. **Stoll BJ, Hansen NI, Adams-Chapman I, et al. (2004):** Neurodevelopmental and growth impairment among extremely low-birth-weight infants with neonatal infection. *JAMA.* 292:2357–2365.
  6. **Joanna Filioti, Kleomenis Spiroglou, Emmanuel Roilides (2007):** Invasive candidiasis in pediatric intensive care patients: epidemiology, risk factors, management, and outcome. Volume 33, Issue 7 / July , Pages 1272 - 1283
  7. **Solorzano Santos F, Reyna Figueroa J, Torres Munoz MJ, Diaz Luna JL. (2008):** Neonatal candidiasis: therapeutic options. *Drugs Today (Barc).* Nov;44 Suppl 4:25-30.
  8. **Manno G, Scaramuccia A, Rossi R, Coppini A, Cruciani M. (2004):** Trends in antifungal use and species distribution among *Candida* isolates in a large paediatric hospital. *Int J Antimicrob Agents.* 24 (6):627– 628
  9. **Makhoul IR, Kassis I, Smolkin T, Tamir A, Sujov P (2001):** Review of 49 Neonates with Acquired Fungal Sepsis: further Characterization. *Pediatrics* 107: 61-66.
  10. **Benjamin DK Jr, Stoll BJ, Fanaroff AA, McDonald SA, Oh W, Higgins RD, Duara S, Poole K, Laptook A, Goldberg R, (2006):** National Institute of Child Health and Human Development Neonatal Research Network Neonatal candidiasis among extremely low birth weight infants: risk factors, mortality rates, and neurodevelopmental outcome at 18 to 22 months. *Pediatrics* 117: 84-92.
  11. **Lain A., Elguezabal N., Brena S., Garcia-Ruiz J.C., del Palacio A., Moragues M.D. and Ponton J. (2007):** Diagnosis of invasive candidiasis by enzyme-linked immunosorbent assay using the N-terminal fragment of *Candida albicans* hyphal wall protein 1. *BMC Microbiol.* ; 7 : 35.
  12. **Arancia S., Carattoli A., La Valle R., Cassone A. and De Bernardis F. (2006):** Use of 65 kDa mannoprotein gene primers in Real Time PCR identification of *Candida albicans* in biological samples. *Molecul. & Cell. probes.* 15 : 2570-577.
  13. **Moreira-Oliveira M.S., Mikami Y., Miyaji M. , Imai T., Schreiber A.Z. and Moretti M.L. (2005):** Diagnosis of candidemia by polymerase chain reaction and blood culture: Prospective study in a high-risk population and identification of variables associated with development of candidemia. *European Journal of Clinical Microbiology and Infectious Diseases* , vol. 24 , P. 721-726.
  14. **Ferns R.B. (2006):** Evaluation of the role of real-time PCR in the diagnosis of invasive aspergillosis. *Leuk. Lymphoma* 47 : 15-20.
  15. **Reischl (2005):** Automated rapid isolation of bacterial DNA from various samples using the MagNA Pure Compact system. *Biochemica* 2 : 12-15.
  16. **Espy M.J., Uhl J.R. , Sloan L.M., Buckwalter S.P., Jones M.F., Vetter E.A., Yao J.D.C., Wengenack N.L., Rosenblatt J.E., Cockerill F.R. III, and Smith T.F. (2006):** Real-Time PCR in clinical

- microbiology : Applications for routine laboratory testing. Clinical microbiology reviews. January P. 165-256 , vol. 19 , No. 1.
17. **Rong Bu , Rajeev K. Sathiapalan , Muna M. Ibrahim, Ibrahim Al-Mohsen, Edna Almodavar, Marina I. Gutierrez and Kishor Bhatia (2005):** Monochrome LightCycler PCR assay for detection and quantification of five common species of *Candida* and *Aspergillus*. *Journal of Medical Microbiology* ; 54 , 243-248.
  18. **Mukta N. Chowta, Prabha Adhikari, A. Rajeev and Ashok K. Shenoy (2007):** Study of risk factors and prevalence of invasive candidiasis in a tertiary care hospital. *Indian J Crit Care Med* April-June , Vol 11 Issue 2.
  19. **Saiman L, Ludington E, Pfaller M, Rangel-Frausto S, et al., (2000):** Risk factors for candidemia in Neonatal Intensive Care Unit patients. The National Epidemiology of Mycosis Survey study group. *Pediatr Infect Dis J.* Apr;19(4):319-24.
  20. **Kristina N. Feja, Fann Wu, Kevin Roberts, Maureen Loughrey, Mirjana Nesin, Elaine Larson, Phyllis Della-Latta, Janet Haas, Jeannie Cimiotti, and Lisa Saiman (2005):** Risk factors for candidemia in critically ill infants. A matched case-control study. *J Pediatr.* August; 147(2): 156–161.
  21. **Benjamin D, et. al. (2003):** Empirical therapy for neonatal candidemia in very low birth weight infants. *Pediatrics*, 112 (3): 543-7.
  22. **Gupta N, et. al. (2001):** Candidemia in neonatal intensive care unit. *Indian Journal of Pathol Microbiol*; 44(1): 45-8.
  23. **Benjamin DK, et. al. (2000):** When to suspect fungal infection in neonates: a clinical comparison for *Candida albicans* and *Candida parapsilosis* fungemia with coagulase - negative staphylococcal bacteremia. *Pediatrics*. 106 (4):712-8.
  24. **L. Saiman, E. Ludington, J. D. Dawson et al., (2001):** “Risk factors for *Candida* species colonization of neonatal intensive care unit patients,” *Pediatric Infectious Disease Journal*, vol. 20, no. 12, pp. 1119–1124.
  25. **Bendel CM; Wiesner SM; Garni RM; Cebelinski E; Wells CL (2002):** Cecal colonization and systemic spread of *Candida albicans* in mice treated with antibiotics and dexamethazone. *Pediatr Res.* 51(3):290-5
  26. **R. P. Wenzel, (1995):** “Nosocomial candidemia: risk factors and attributable mortality,” *Clinical Infectious Diseases*, vol. 20, no. 6, pp. 1531–1534.
  27. **Kaufman D and Fairchild KD. (2004):** Clinical microbiology of bacterial and fungal sepsis in very low birth weight infants. *Clin Microbiol rev*; 17(3): 638-680.
  28. **B. J. Stoll, N. Hansen, A. A. Fanaroff et al., (2002):** “Late-onset sepsis in very low birth weight neonates: the experience of the NICHD Neonatal Research Network,” *Pediatrics*, vol. 110, no. 2, part 1, pp. 285–291.
  29. **Makhoul IR, Kassis I, Smolkin T, Tamir A and Sujov P. (2001):** Review of 49 Neonates With Acquired Fungal Sepsis: Further

- Characterization. *Pediatrics*; 107; 61-66.
30. **M. S. Rangel-Frausto, T. Wiblin, H. M. Blumberg et al., (1999):** "National epidemiology of mycoses survey (NEMIS): variations in rates of bloodstream infections due to *Candida* species in seven surgical intensive care units and six neonatal intensive care units," *Clinical Infectious Diseases*, vol. 29, no. 2, pp. 253–258.
31. **Rodriguez D, Almirante B, Park BJ, Cuenca- Estrella M, Planes AM, Sanchez F et al., (2006):** on behalf of the Barcelona candidemia project study group. *Candidemia in Neonatal Intensive care units. Pediatric Infect Dis J*; 25: 224-229.
32. **R. G. Faix, (1992):** "Invasive neonatal candidiasis: comparison of *albicans* and *parapsilosis* infection," *Pediatric Infectious Disease Journal*, vol. 11, no. 2, pp. 88–93.
33. **A. Lupetti, A. Tavanti, P. Davini et al., (2002):** "Horizontal transmission of *Candida parapsilosis* candidemia in a neonatal intensive care unit," *Journal of Clinical Microbiology*, vol. 40, no. 7, pp. 2363–2369.
34. **Mari´a L. Avila-Aguero, Alejandro Canas-Coto. Et al., (2005):** Risk factors for *Candida* infections in a neonatal intensive care unit in Costa Rica. *International Journal of Infectious Diseases* (2005) 9, 90-95
35. **Roilides E, Farmaki E, Evdoridou J, Francesconi A, Kasai M, Filioti J, et al. (2003):** *Candida tropicalis* in a neonatal intensive care unit: epidemiologic and molecular analysis of an outbreak of infection with an uncommon neonatal pathogen. *J Clin Microbiol*;41:735-41.
36. **Finkelstein R, Reinhertz G, Hashman N, Merzbach D. (1993):** Outbreak of *Candida tropicalis* fungemia in a neonatal intensive care unit. *Infect Control Hosp Epidemiol*;14:587-90.
37. **Fairchild KD, Tomkoria S, Sharp EC, Mena FV. (2002):** Neonatal *Candida glabrata* sepsis: clinical and laboratory features compared with other *Candida* species. *Pediatr Infect Dis J*; 21: 39-43.
38. **Wahyuningsih R, Freisleben HJ, Sonntag HG, Schnitzler P. (2000):** Simple and rapid detection of *Candida albicans* DNA in serum by PCR for diagnosis of invasive candidiasis. *J Clin Microbiol*; 38: 3016–3021.
39. **Metwally L., Fairley D.J., Coyle P.V., Hay R.J., Hedderwick S., McCloskey B., O'Neill H.J., Webb C.H. and Mc Mullan R. (2008):** Comparison of serum and whole blood specimens for the detection of *Candida* DNA in critically ill , non-neutropenic patients. *J Med Microbiol* . Oct; 57 (Pt 10).
40. **Einsele H, Hebart H, Roller G et al. (1997):** Detection and identification of fungal pathogens in blood by using molecular probes. *J Clin Microbiol*; 35: 1353–1360.
41. **Loeffler J, Hebart H, Brauchle U, Schumacher U, Einsele H. (2000):** Comparison between plasma and whole blood specimens for detection of *Aspergillus* DNA by PCR. *J Clin Microbiol*; 38: 3830–3833.
42. **M. Khlif, C. Mary, H. Sellami , A. Sellami , H. Dumon , A. Ayadi and S. Ranque (2009):** Evaluation of nested and real-time PCR assays in the diagnosis of candidaemia. *Clinical*

- Microbiology and Infection. Volume 15, Issue 7, pages 656–661, July
43. **White P.L., Archer A.E. and Barnes R.A. (2005):** Comparison of non-culture based methods for detection of systemic fungal infections, with an emphasis on invasive *Candida* infections. *J. Clin. Microbiol.*, 43 (5): 2181-2187.
  44. **Maaroufi Y, Ahariz N, Husson M, Crokaert F. (2004):** Comparison of different methods of isolation of DNA of commonly encountered *Candida* species and its quantitation by using a real-time PCR-based assay. *J Clin Microbiol*; 42: 3159–3163.
  45. **Mc Mullan R., Metwally L., Coyle P.V., et al., (2008):** A prospective clinical trial of a real-time polymerase chain reaction assay for the diagnosis of candidemia in nonneutropenic, critically ill adults. *Clin Infect Dis.* March 15 ; 46 (6) : 890-6.
  46. **Alam F.F., Mustafa A.S. and Khan Z.U. (2007):** Comparative evaluation of (1,3)-beta-D-glucan, mannan and anti-mannan antibodies, and *Candida* species-specific sn PCR in patients with candidemia. *BMC Infect Dis.* Sep. 2007 4; 7 (1): 103
  47. **Chang HC, Leaw SN, Huang AH, Wu TL, Chang TC. (2001):** Rapid identification of yeasts in positive blood cultures by a multiplex PCR method. *J Clin Microbiol*; 39: 3466–3471.
  48. **Morace G., Sanguinetti M., Posteraro B., Cascio G.L. and Fadda G. (1999):** Identification of various medically important *Candida* species in clinical specimens by PCR-restriction enzyme analysis. *J. Clin. Microbiol.* 35:667-672.
  49. **Claudia S., Brigitte S., Manfred L.R., Alexander M.H. and Birgit W. (2007):** Development of novel Real-Time PCR assay for detection and differentiation of eleven medically important *Aspergillus* and *Candida* species in clinical specimens. *Journal of clinical microbiology*, March 2007, P. 906-914, vol. 45, No.3.
  50. **Pryce TM, Kay ID, Palladino S, Heath CH. (2003):** Real-time automated polymerase chain reaction (PCR) to detect *Candida albicans* and *Aspergillus fumigatus* DNA in whole blood from high-risk patients. *Diagn Microbiol Infect Dis*; 47: 487–496.
  51. **Polak A. (2003):** Antifungal therapy – state of the art at the beginning of the 21st century. *Prog Drug Res*; 59-190.
  52. **White PL, Shetty A, Barnes RA. (2003):** Detection of seven *Candida* species using the Light-Cycler system. *J Med Microbiol*; 52: 229-238.
  53. **White PL, Barton R, Guiver M et al. (2006):** A consensus on fungal polymerase chain reaction diagnosis? A United Kingdom–Ireland evaluation of polymerase chain reaction methods for detection of systemic fungal infections. *J Mol Diagn*; 8: 376–384.
  54. **Schabereiter-Gurtner C, Selitsch B, Rotter ML, Hirschl AM, Willinger B. (2007):** Development of novel real-time PCR assays for detection and differentiation of 11 medically important *Aspergillus* and *Candida* species in clinical specimens. *J Clin Microbiol*; 45: 906–914.

## الكشف المبكر عن فطر الكانديدا البيكانز في مرضى العناية المركزة للأطفال حديثي الولادة

رجاء عبد السلام محمد إبراهيم\* - صلاح السيد إبراهيم على\*\* - هناء حسنى\*\*  
حنان سمير أحمد\*\*

\*قسم الأطفال - كلية طب الأزهر بنات - \*\*قسم الباثولوجيا الإكلينيكية  
كلية الطب - جامعة الزقازيق

يعتبر وجود فطر الكانديدا بالدم واحدا من أهم الأسباب التي تؤدي ظهور تسمم الدم المتأخر في الأطفال حديثي الولادة وقد تسببت في حدوث نسبة عالية من الاعتلال و الوفيات.

والهدف من هذا البحث : هو تقييم طريقة تفاعل البلمرة المتسلسل في الزمن الحقيقي للتشخيص المبكر لعدوى الكانديدا البيكانز بالدم في مرضى العناية المركزة للأطفال حديثي الولادة.

وقد شملت هذه الدراسة ستون مريضا من أطفال أدخلوا للعناية المركزة للأطفال حديثي الولادة وكان يوجد عندهم سببا واحدا أو أكثر يؤدي للإصابة بفطر الكانديدا بالدم. ستون عينة دم خضعت للفحص باستخدام طريقة مزارع الدم التقليدية وبطريقة تفاعل البلمرة المتسلسل في الزمن الحقيقي لاكتشاف العدوى . وأصفر البحث عن وجود عشرون حالة مصابة بفطر الكانديدا بالدم وأعتبر الأربعةون حالة الأخرى كمجموعة ضابطة.

و قد أوضحت هذه الدراسة أن : وجد أن مزارع الدم التقليدية كانت إيجابية في عشرون عينة من الستون عينة محل البحث وكان فطر الكانديدا البيكانز الأكثر شيوعا بين حالات المصابة الكانديدا بالدم (75%) ويليه فطر الكانديدا تروبيكانز (20%) ثم

فطر الكانديدا جلابراتا (5%) . تفاعل البلمرة المتسلسل فى الزمن الحقيقي للكشف عن فطر الكانديدا البيكانز كان إيجابيا فى عينة أربعة عشر عينة من الخمسة عشر عينة التي تم كشفها بطريقة مزارع الدم التقليدية كما أوضح البحث أن هذه الطريقة أعطت حالة إيجابية كاذبة وأخرى سالبة أوضحت الدراسة أن حساسية هذه الطريقة 93.3% ودرجة تحديدها 97.8% والدقة 96.7%. ووجود القسطرة الوريدية من أشهر عوامل الخطورة للإصابة بهذه العدوى ويليها التنفس الصناعي والأنبوبة الحنجرية والحجز أكثر من عشرة أيام بالمحضن (45%) لكل عامل. من العشرين مريضا المصابين بالكانديدا فى الدم كان هناك سبعة عشر مريضا لديهم أكثر من عامل من عوامل الخطورة المسببة للعدوى بالكانديدا فى الدم (22.7).

ونستنتج من هذه الدراسة : أن معرفة عوامل الخطورة هو الأهم فى توقع الأطفال الأكثر عرضة لحدوث عدوى الكانديدا فى الدم بالرغم من أن مزارع الدم تعتبر هي المقياس الذهبى أو الطريقة الأكيدة لتشخيص عدوى الكانديدا بالدم ولكن حساسيتها ودقتها فى تشخيص عدوى الكانديدا بالدم منخفضة ومضيعة للوقت. أن تفاعل البلمرة المتسلسل فى الزمن الحقيقي هو اختبار ذو حساسية ودرجة تحديد ودقة عالية فى تشخيص عدوى الكانديدا بالدم وطبقا للدراسة الحالية فإن تفاعل البلمرة المتسلسل فى الزمن الحقيقي تستخدم كطريقة سريعة فى تشخيص الكانديدا بالدم وتساعد على العلاج المبكر لاستخدام مضادات الكانديدا وتقلل الآثار الجانبية العديدة للكانديدا بالدم.