



## EFFICACY OF CERTAIN FUNGICIDES AND ANTAGONISTIC MICROORGANISMS ON MYCELIAL GROWTH OF *Fusarium oxysporum* ISOLATED FROM DATE PALM IN NORTH SINAI

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### ABSTRACT

In this study the efficacy of three chemical fungicides as well as five different biological control agents were tested *in vitro* against several isolates of *Fusarium oxysporum* from wilted date palm trees grown in North Sinai. The three commercial fungicides were: Tachigaren 30% SL, Rhizolex-T 50% WP and Dithane M-45 80% WP. Whereas the biological control agents included three fungal species namely: *Trichoderma album*, *T. harzianum* and *T. strigosum*. As well as the two bacteria: *Bacillus megaterium* and *B. subtilis*. Tachigaren was the most effective fungicide in reducing the fungal growth of all tested isolates of *F. oxysporum*. Rhizolex T-50 ranked second while Dithane M-45 was the least effective fungicide in reducing the mycelial growth of the tested isolates of *F. oxysporum in vitro*. *Trichoderma harzianum* was the most effective biocontrol agent in inhibiting the mycelial growth of all tested *F. oxysporum* isolates followed by *T. album* and then *T. strigosum*. On the other hand, *Bacillus megaterium* and *Bacillus subtilis* were generally non effective in reducing mycelial growth of the tested isolates of *F. oxysporum in vitro*.

**Kew word:** Certain Fungicides, antagonistic microorganisms, *Fusarium oxysporum* isolated, date palm, North Sinai.

### INTRODUCTION

The date palm (*Phoenix dactylifera* L.) is a major fruit crop in most Arab countries. It has historically been connected with sustaining human life and tradition of the people in the old world as a major agricultural crop. Arab countries possess 70% of the 120 million world's date palms and are responsible for 67% of the global date production **Loutfy (2010)**. *Fusarium* wilt diseases caused by *Fusarium oxysporum* are among the most severe plant diseases in the world. *Fusarium* wilts affect many plant species belonging to all the botanical families. *F. oxysporum* is one of the most common species among soil fungi in cultivated soil all over the world. It includes a large diversity of strains: saprophytic, parasitic and pathogenic.

*Fusarium oxysporum* (Fo) is a ubiquitous inhabitant of soils worldwide and causes diseases such as wilt, yellows and damping-off in different plant species **Ratul and Narendra (2010)**. The fungus can invade plant's roots with its conidial germ tube or mycelium. The roots can be infected directly through the root tips, wounds in the roots or at the formation point of lateral roots. Mycelium enters the xylem vessels branches and produces micro-conidia which are carried upward in the sap stream. Due to the growth of the fungus within the plant's vascular tissue, the plant's water supply is greatly affected. This lack of water induces the leaves stomata to close, the leaves wilt and the plant eventually dies. Infection usually leads to chlorosis, leaf wilting and browning of the vascular system. Several procedures have been attempted for managing

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the fusarium wilt disease in the greenhouse and in field, **Ioannou (1999)**. Bayoud of date palm caused by the fungus *Fusarium oxysporum* f.sp. *albedinis* (Foa) is the most important disease of this crop. It is currently confined to Morocco and the western and central regions of Algeria, having destroyed approximately 15 million trees since it was first discovered, sometime before 1870, **Djerbi (1983)**.

In this research we studied the effects of some chemical fungicides as well as bio-fungicides, on the growth of several isolates of *Fusarium oxysporum* isolated from date palm grown in North Sinai.

## MATERIALS AND METHODS

### Isolation of Fungal Pathogens

*Fusarium oxysporum* cultures were isolated from infected roots of date palm trees showing wilt, drying, yellow leaves and progressive die back from different districts located in four different geographic regions in North Sinai. Root samples were washed carefully with tap water to remove the adhering soil. Diseased root samples were cut into pieces (1cm long). Surface sterilized with dipping in 0.5% sodium hypochlorite solution for 2 minutes, then washed thoroughly with several changes of sterile distilled water. The cut pieces were dried between sterile filter papers. The surface sterilized pieces were transferred to Petri dishes containing acidified potato dextrose agar medium (PDA) then incubated at  $25 \pm 2^\circ\text{C}$  for two weeks then inspected for fungal growth, **Rashed (1991)**, **El-Morsi (1999)**, **Abdalla *et al.* (2001)**, **El-Morsi (2004)** and **Baraka *et al.* (2011)**.

### *Fusarium oxysporum* Control *In vitro*

#### Effect of chemical fungicides on the linear growth of *F. oxysporum*

Three fungicides: Hymexazol (Tachigaren 30% SL), Tolclofos- methyl (Rhizolex-T

50% WP) and Mancozeb (Dithane M-45 80% WP) were tested individually at different concentration (10, 50, 100, 500 and 1000 ppm). The effects of the fungicides were assayed by using the poisoned food technique according to the method described by **Abdalla *et al.* (1992)**. Fungicides were added to autoclaved PDA medium before dispensing medium in Petri plates, when temperature of the medium was about  $50^\circ\text{C}$ . Control treatment was PDA medium without fungicides additions.

Disks (5mm) in diameter from 7 days old cultures of the tested fungus were transferred to the center of Petri plates then the plates were incubated at  $27 \pm 2^\circ\text{C}$  for 16 days. After 4, 8, 12 and 16 days of incubation the linear growth of the tested was assessed by measuring two perpendicular diameters in cm and the average was recorded.

### Biological control

The *Trichoderma album* isolate was isolated from the commercial biological control product called "Bio zeid", *Trichoderma strigosum* and *Trichoderma harzianum* were isolated by one gram of dry soil from each sample was added to test tubes containing 9 ml sterile water and each test tube was shaken periodically for approximately 15 minutes. After shaking one ml of suspension was added to test tube containing 9 ml sterile water, and the process was repeated to make serial dilutions up to 1:10000 for each fungus isolation. Aliquots 1 ml of serial dilutions was spread on APDA plates. All plates were incubated at  $25^\circ\text{C}$ . After 2-7 days the isolated microorganisms, were identified according to their morphological characters, **Samuels and Hebbar (2015)**. Selected microorganisms were purified and transferred on to APDA slants; pure cultures were stored at  $4^\circ\text{C}$  for further studies. The antagonistic fungi were identified at Unit of Identification of Microorganisms, Plant Pathology Research Institute, Agricultural Research Center, using BIOLOG™ system.

The *Bacillus megaterium* culture was isolated from the commercial biological control product called “Bio Arc” and the *Bacillus subtilis* culture was isolated from the commercial biological control product called “Clean Root”.

The biological control agents were tested against *F. oxysporum* isolates *in vitro* using acidified potato dextrose agar (APDA) medium in 9cm diameter Petri plates. Five mm diameter disc of a four days old fungal growth of the tested *F. oxysporum* isolates were placed at about two cm from the edge of the plates.

On the opposite side of the Petri plates a five mm disc of the tested antagonistic fungi was placed. Plates were incubated at 25°C and kept under daily observation and data were recorded one week later as linear growth of the *F. oxysporum* cultures as described by **Abd El-Moity et al. (1993)** and **Perveen and Bokhari (2012)**. The bacterium was maintained on PDA medium at 33±2°C. The *Bacillus* sp. isolate was tested against *F. oxysporum* using PDA medium in 9 cm diameter Petri plates. 5 mm diameter discs of the tested fungus was placed two cm from the edge of the plate. On the opposite side of the Petri plates at about 2 cm from the edge a streak of the tested antagonistic bacterium was placed as suggested by **Agarwal et al. (2011)**. These experiments were repeated twice and data were recorded.

## RESULTS AND DISCUSSION

### *Fusarium oxysporum* Control *In vitro*

#### Effect of chemical fungicides on linear growth of *F. oxysporum*

*In vitro* studies Table 1 indicate that Tachigaren 30 % SL was very effective in controlling radial growth of *F. oxysporum* at 1000 ppm on tested isolates (D<sub>2</sub>, E<sub>1</sub> and F<sub>7</sub>) 16 days after incubation, but it showed little effect on isolate B<sub>2</sub> 16 days after

incubation. Rhizolex-T 50% WP ranked second in controlling radial growth of *F. oxysporum* isolates (E<sub>1</sub> and F<sub>7</sub>). It was less effective on linear growth of *F. oxysporum* isolate D<sub>2</sub>, and showed no significant effect on isolate B<sub>2</sub> after 16 days of incubation. The fungicide Dithane M-45 80% WP was the least effective fungicide in controlling radial growth of *Fusarium oxysporum* isolates (B<sub>2</sub>, D<sub>2</sub>, E<sub>1</sub> and F<sub>7</sub>) at all tested concentrations 16 days after incubation on APDA medium.

These results are in agreement with those obtained by **Haggag and El-Gamal (2012)** who found that Tachigaren was the most effective fungicide in reducing the mycelial growth of *Fusarium solani*. Similarly, **Song et al. (2004)** demonstrated that Prochloraz and Carbendzim were more efficient in reducing mycelial growth of *F. oxysporum* f.sp. *lycopersici* compared with Tachigaren and Dithane M-45.

#### *In vitro* antagonism of microorganisms on the pathogen *Fusarium oxysporum*

All tested isolates of *Trichoderma strigosum*, *T. harzianum* and *T. album* suppressed the growth of *F. oxysporum* isolates (B<sub>2</sub>, D<sub>2</sub>, E<sub>1</sub> and F<sub>7</sub>), this inhibition occurred before any direct mycelial interaction took place (Table 2 and Plate 1).

*Trichoderma strigosum* isolate significantly suppressed the growth of *F. oxysporum* isolates and it subsequently mycoparasitised the pathogen expect in case of isolate B<sub>2</sub>, *T. strigosum* did not overgrow it.

*Trichoderma harzianum* isolate also significant suppressed the growth of *F. oxysporum* isolates and it subsequently mycoparasitised the pathogen expect in case of isolate B<sub>2</sub>, *T. harzianum* did not overgrow it.

*Trichoderma album* isolates was significantly inhibiting the growth of the pathogen *F. oxysporum* isolates and it subsequently mycoparasitised the pathogen, expect isolate B<sub>2</sub>.

**Table (1): Effect of five concentrations of three fungicides “Tachigaren, Rhizolex and Dithane M-45” on the linear growth (mm) of *F. oxysporum in vitro* isolate (B<sub>2</sub>, D<sub>2</sub>, E<sub>1</sub> and F<sub>7</sub>) 4, 8, 12 and 16 days after incubation.**

| Treatments  |                      | Rate growth rate of <i>Fusarium oxysporum</i> isolates after incubation (day) |            |            |            |           |
|---|----------------------|---|------------|------------|------------|-----------|
|   |                      | 4 days  | 8 days     | 12 days    | 16 days    |           |
| Fungal isolate “B <sub>2</sub> ” from Bear Al Abd | Control              | 30.00 a   | 50.00 fgh  | 90.00 a    | 90.00 a    |           |
|   | Tachigaren 30% SL    | 10 ppm  | 21.50 ghi  | 44.00 jklm | 60.50 jkl  | 76.15 ij  |
|   |                      | 50 ppm  | 19.75 ijkl | 42.87 lmn  | 56.50 mn   | 78.75 hi  |
|   |                      | 100 ppm   | 16.62 n    | 37.25 pq   | 50.25 op   | 82.00 fgh |
|   |                      | 500 ppm   | 4.50 vw    | 13.50 DE   | 27.25 z    | 22.60 yz  |
|   |                      | 1000 ppm  | 3.62 w     | 11.75 E    | 16.75 B    | 18.87 A   |
|   | Rhizolex-T 50 % WP   | Control   | 30.00 a    | 50.00 fgh  | 90.00 a    | 90.00 a   |
|   |                      | 10 ppm  | 23.00 defg | 41.00 mno  | 71.00 hi   | 90.00 a   |
|   |                      | 50 ppm  | 17.00 mn   | 34.00 rs   | 61.50 jk   | 90.00 a   |
|   |                      | 100 ppm   | 12.25 q    | 25.50 wx   | 46.00 qrs  | 72.82 kl  |
|   |                      | 500 ppm   | 9.00 tu    | 20.50 zA   | 41.50 tu   | 62.75 p   |
|   | Dithane M-45 80 % WP | Control   | 30.00 a    | 50.00 fgh  | 90.00 a    | 90.00 a   |
|   |                      | 10 ppm  | 22.25 efgh | 46.28 ij   | 69.25 hi   | 88.75 abc |
|   |                      | 50 ppm  | 19.25 kl   | 40.56 nno  | 59.75 jklm | 80.50 gh  |
|   |                      | 100 ppm   | 16.18 no   | 36.81 pqr  | 53.43 no   | 86.50 bed |
| 500 ppm   |                      | 14.31 p   | 31.93 stu  | 50.43 op   | 84.25 def  |           |
| Fungal isolate “D <sub>2</sub> ” from Rafah       | Tachigaren 30% SL    | Control   | 30.00 a    | 50.00 fgh  | 90.00 a    | 90.00 a   |
|   |                      | 10 ppm  | 14.50 op   | 30.50 tu   | 37.25 vw   | 53.10 rs  |
|   |                      | 50 ppm  | 12.06 qr   | 26.25 wx   | 38.25 uv   | 49.62 t   |
|   |                      | 100 ppm   | 10.21 st   | 24.50 wxy  | 34.25 wxy  | 47.82 tu  |
|   |                      | 500 ppm   | 0.00 x     | 0.00 F     | 0.00 D     | 2.50 B    |
|   | Rhizolex-T 50 % WP   | Control   | 30.00 a    | 55.00 cd   | 82.00 c    | 90.00 a   |
|   |                      | 10 ppm  | 24.55 cd   | 54.25 de   | 80.25 cd   | 90.00 a   |
|   |                      | 50 ppm  | 10.28 rst  | 31.25 stu  | 49.00 pq   | 75.45 jk  |
|   |                      | 100 ppm   | 9.08 tu    | 25.57 wx   | 38.00 v    | 56.45 q   |
|   |                      | 500 ppm   | 0.00 x     | 16.00 BCD  | 18.75 B    | 42.82 v   |
|   | Dithane M-45 80 % WP | Control   | 30.00 a    | 55.00 cd   | 82.00 c    | 90.00 a   |
|   |                      | 10 ppm  | 25.01 c    | 50.00 fgh  | 77.00 de   | 89.50 ab  |
|   |                      | 50 ppm  | 21.20 hij  | 42.50 lmn  | 69.50 hi   | 89.75 ab  |
|   |                      | 100 ppm   | 19.77 ijkl | 38.25 op   | 59.50 jklm | 85.25 def |
|   |                      | 500 ppm   | 19.55 jkl  | 42.00 lmn  | 53.25 no   | 71.37 lm  |
|   | 1000 ppm             | 14.65 op  | 32.75 st   | 44.50 rst  | 65.00 op   |           |

**Table (1): Con. Effect of five concentrations of three fungicides “Tachigaren, Rhizolex and Dithane M-45” on the linear growth (mm) of *F. oxysporum* *in vitro* isolate (B<sub>2</sub>, D<sub>2</sub>, E<sub>1</sub> and F<sub>7</sub>) 4, 8, 12 and 16 days after incubation.**

| Treatments  |                                  |                        | Growth rate of <i>Fusarium oxysporum</i> isolates after incubation (day) |            |           |           |         |
|---|----------------------------------|------------------------|--|------------|-----------|-----------|---------|
|   |                                  |                        | 4 days   | 8 days     | 12 days   | 16 days   |         |
| Fungal isolate “E <sub>1</sub> ” from Al Shaikh Zewayed | Tachigaren<br>30% SL             | Control                | 30.00 a  | 65.00 a    | 90.00 a   | 90.00 a   |         |
|   |                                  | 10 ppm                 | 22.50 efgh   | 48.50 ghi  | 68.00 i   | 85.75 cde |         |
|   |                                  | 50 ppm                 | 18.50 lm   | 41.25 lmno | 58.50 klm | 82.95 efg |         |
|   |                                  | 100 ppm                | 19.00 l  | 34.25 qrs  | 53.00 o   | 75.45 jk  |         |
|   |                                  | 500 ppm                | 3.42 w   | 14.25 DE   | 18.75 B   | 25.50 y   |         |
|   |                                  | 1000 ppm               | 0.00 x   | 0.00 F     | 0.00 D    | 0.00 B    |         |
|   | Rhizolex-T<br>50% WP             | Control                | 30.00 a  | 65.00 a    | 90.00 a   | 90.00 a   |         |
|   |                                  | 10 ppm                 | 20.90 hijk   | 47.50 hi   | 57.75 lm  | 72.50 kl  |         |
|   |                                  | 50 ppm                 | 11.42 qrs  | 29.50 uv   | 43.25 rst | 69.00 mn  |         |
|   |                                  | 100 ppm                | 8.87 tu  | 19.00 zAB  | 33.00 xy  | 44.67 uv  |         |
|   |                                  | 500 ppm                | 0.00 x   | 14.50 CDE  | 22.37 A   | 35.40 w   |         |
|   |                                  | 1000 ppm               | 0.00 x   | 0.00 F     | 0.00 D    | 20.90 zA  |         |
|   | Fungal isolate “E <sub>1</sub> ” | Dithane M-45<br>80% WP | Control  | 30.00 a    | 65.00 a   | 90.00 a   | 90.00 a |
|   |                                  |                        | 10 ppm   | 24.87 c    | 52.05 def | 75.50 ef  | 90.00 a |
|   |                                  |                        | 50 ppm   | 19.07 l    | 44.12 jkl | 62.75 j   | 90.00 a |
| 100 ppm   |                                  |                        | 21.42 ghi  | 43.00 klmn | 61.50 jk  | 90.00 a   |         |
| 500 ppm   |                                  |                        | 22.20 fgh  | 47.25 hi   | 75.75 ef  | 90.00 a   |         |
| 1000 ppm  |                                  |                        | 23.05 defg   | 48.25 ghi  | 77.50 de  | 90.00 a   |         |
| Tachigaren<br>30% SL                                    |                                  | Control                | 30.00 a  | 63.00 a    | 90.00 a   | 90.00 a   |         |
|   |                                  | 10 ppm                 | 10.50 qrst   | 27.02 vw   | 43.00 st  | 71.00 lm  |         |
|   |                                  | 50 ppm                 | 9.57 tu  | 21.50 yz   | 35.50 vwx | 56.25 qr  |         |
|   |                                  | 100 ppm                | 7.85 u   | 17.50 ABC  | 31.25 y   | 31.00 x   |         |
| Fungal isolate “F <sub>7</sub> ” from Al Arish          | Rhizolex-T<br>50% WP             | 500 ppm                | 0.00 x   | 0.00 F     | 0.00 D    | 0.00 B    |         |
|   |                                  | 1000 ppm               | 0.00 x   | 0.00 F     | 0.00 D    | 0.00 B    |         |
|   |                                  | Control                | 29.00 a  | 63.00 a    | 90.00 a   | 90.00 a   |         |
|   |                                  | 10 ppm                 | 27.02 b  | 59.00 b    | 77.25 de  | 90.00 a   |         |
|   |                                  | 50 ppm                 | 15.22 nop  | 42.50 lmn  | 71.75 gh  | 90.00 a   |         |
|   |                                  | 100 ppm                | 11.45 qrs  | 26.00 wx   | 44.00 rst | 66.62 no  |         |
|   | Dithane M-45<br>80% WP           | 500 ppm                | 0.00 x   | 12.85 E    | 27.50 z   | 50.00 st  |         |
|   |                                  | 1000 ppm               | 0.00 x   | 0.00 F     | 0.00 D    | 21.10 zA  |         |
|   |                                  | Control                | 29.00 a  | 63.00 a    | 90.00 a   | 90.00 a   |         |
|   |                                  | 10 ppm                 | 27.07 b  | 51.25 efg  | 88.50 ab  | 90.00 a   |         |
|   |                                  | 50 ppm                 | 23.00 defg   | 50.25 fgh  | 72.50 fgh | 90.00 a   |         |
|   |                                  | 100 ppm                | 23.50 cdef   | 46.00 ijk  | 74.75 efg | 90.00 a   |         |
|   |                                  | 500 ppm                | 24.00 cde  | 58.00 bc   | 86.25 b   | 90.00 a   |         |
|   |                                  | 1000 ppm               | 27.00 b  | 65.25 a    | 90.00 a   | 90.00 a   |         |
| LSD 0.05  |                                  |                        | 1.777  | 3.014      | 3.342     | 3.257     |         |

Table (2): The antagonistic effect of the bio-control agent: *T. strigosum*, *T. harzianum*, *T. album*, *Bacillus megaterium* and *B. subtilis* against *F. oxysporum* isolates growth (mm) *in vitro* 8 days after incubation.

| Origin of fungal isolates                               | Treatment            | Main linear growth of <i>F. oxysporum</i> (mm) |         |           |
|---|----------------------|--|---------|-----------|
|   |                      | Microorganism against <i>F. oxysporum</i>      | Control | * PGI (%) |
| Fungal isolate "B <sub>2</sub> " from Bear AIAbd        | <i>T. strigosum</i>  | 25.50 e  | 90      | 71.66     |
|   | <i>T. harzianum</i>  | 26.75 e  |         | 70.27     |
|   | <i>T. album</i>      | 19.72 fgh                                      |         | 78.17     |
|   | <i>B. megaterium</i> | 50.00 a  |         | 44.44     |
|   | <i>B. subtilis</i>   | 45.00 b  |         | 50        |
| Fungal isolate "D <sub>2</sub> " from Rafah             | <i>T. strigosum</i>  | 25.05 e  | 90      | 72.16     |
|   | <i>T. harzianum</i>  | 18.35 gh                                       |         | 79.61     |
|   | <i>T. album</i>      | 24.82 e  |         | 72.42     |
|   | <i>B. megaterium</i> | 45.00 b  |         | 50        |
|   | <i>B. subtilis</i>   | 37.00 d  |         | 5.88      |
| Fungal isolate "E <sub>1</sub> " from AI Shaikh Zewayed | <i>T. strigosum</i>  | 26.00 e  | 90      | 71.11     |
|   | <i>T. harzianum</i>  | 18.72 gh                                       |         | 79.20     |
|   | <i>T. album</i>      | 20.87 fg                                       |         | 76.81     |
|   | <i>B. megaterium</i> | 48.00 a  |         | 46.66     |
|   | <i>B. subtilis</i>   | 42.00 c  |         | 53.33     |
| Fungal isolate "F <sub>7</sub> " from AI Arish          | <i>T. strigosum</i>  | 22.07 f  | 90      | 75.47     |
|   | <i>T. harzianum</i>  | 16.91 h  |         | 81.21     |
|   | <i>T. album</i>      | 17.97 gh                                       |         | 80.03     |
|   | <i>B. megaterium</i> | 48.00 a  |         | 46.66     |
|   | <i>B. subtilis</i>   | 45.00 b  |         | 50        |

LSD 0.05

\* PGI (Percent Growth Inhibition) =  $\frac{\text{Colony growth in control plate} - \text{Colony growth in intersecting plate}}{\text{Colony growth in control plate}} \times 100$

- Each treatment represented the mean of six replicates for two experiment.

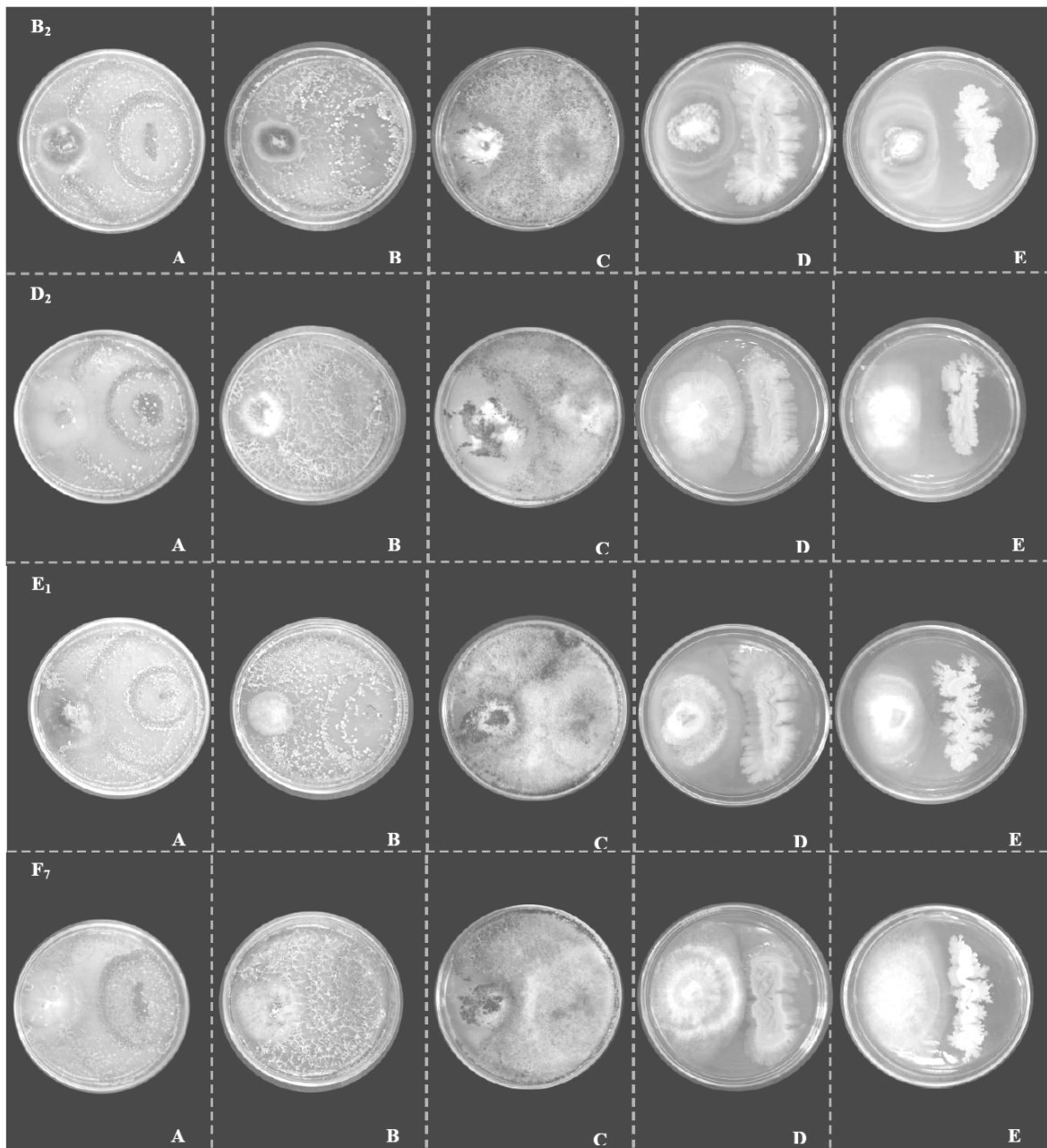
Similar results were obtained by **Perveen and Bokhari (2012)**. They studied the antagonistic potential of *T. harzianum* and *T. viride* against *F. oxysporum* isolated from date palm. They concluded that all tested isolates of *Trichoderma* showed appreciable inhibition for the mycelial growth of *F. oxysporum*.

*Bacillus megaterium* was non-significant in controlling radial growth of all tested *Fusarium oxysporum* isolates 8 days after incubation, no inhibition zones were

observed with any of these isolates after incubation on PDA medium.

*Bacillus subtilis* was very effective in controlling radial growth of *F. oxysporum* isolates (B<sub>2</sub>, D<sub>2</sub> and E<sub>1</sub>) except isolate F<sub>7</sub> it was non-significant in controlling radial growth of the pathogen after incubation on PDA medium.

Several researchers have investigated bio-control strategies using various bacterial species to control *F. oxysporum* f.sp. *albedinis*



**Plate (1): The antagonistic effect of *T. strigosum* (A), *T. harzianum* (B), *T. album* (C), *B. megaterium* (D) and *B. subtilis* (E) against *F. oxysporum* isolates growth *in vitro* after 8 days incubation**

which cause the most destructive disease of date palm, called Bayoud disease. For instance, **Dihazi *et al.*, (2012)**, found that *Bacillus amyloliquefaciens* and *Burkholderia cepacia* significantly inhibited growth and sporulation of *F. oxysporum* f.sp. *albedinis*. Also, selected three bacterial species namely: *B. pumilus*, *Rahnella aquatilis*,

*Bacillus oereus* for their high inhibition toward mycelial growth of *F. oxysporum* f.sp. *albedinis*.

These success in finding biological control agents against *F. oxysporum* are providing hope for establishing biocontrol strategy against *Fusarium* wilt of date palm.

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## المخلص العربي

## تأثير بعض المسببات الفطرية والكائنات البيولوجية المضادة علي نمو فطر الفيوزاريوم أوكسيسبورم المعزول من نخيل البلح في شمال سيناء

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تم في هذا البحث دراسة تأثير بعض المبيدات الكيميائية المتداولة وكذلك بعض الفطريات والبكتريا المستخدمين في مجال مكافحة الحيوية علي عدة عزلات من الفطر *Fusarium oxysporum* والتي تم عزلها من جذور أشجار نخيل مصابة معملياً في محافظة شمال سيناء، في هذا البحث تم استخدام ثلاث مبيدات فطرية تجارية هي التاتشجرين والرايزوليكس والدايثين م-٤٥ وذلك بخمس تركيزات هي (١٠-٥٠-١٠٠-٥٠٠-١٠٠٠ جزء في المليون) بالإضافة إلي الكائنات الحية الدقيقة المستخدمة في المقاومة الحيوية مثل فطر الترايكوديرما هارزيانم والترايكوديرما البوم والترايكوديرما سترابجوزم وبكتريا الباسيليس ميجاتيريوم والباسيليس سيبتيليس، كان المبيد الفطري تاتشجرين هو أكثر تأثيراً في تثبيط النمو الميسليومي للمسبب المرضي الفيوزاريوم أوكسيسبورم في المعمل يليه الرايزوليكس ثم الدايثين م-٤٥ الذي أظهر أقل تأثير في تثبيط نمو الفطر. فطريات الترايكوديرما كانت الأعلى تأثيراً في تثبيط نمو الفيوزاريوم أوكسيسبورم حيث كان الفطر تراكوديرما هارزيانم هو أكثر الأجناس المضادة لنمو المسبب المرضي في المعمل يليه ترايكوديرما البوم ثم ترايكوديرما سترابجوزم بينما بكتريا الباسيليس ميجاتيريوم لم تظهر أي تأثير يذكر في تثبيط نمو الفطر، أما البكتريا باسيليس سابتيليس كان لها تأثير واضح في تثبيط نمو الفطر عدا العزلة F7 لم يكون لها أي تأثير مضاد مع المسبب المرضي.

**الكلمات الاسترشادية:** المسببات الفطرية، الكائنات البيولوجية المضادة، فطر الفيوزاريوم أوكسيسبورم، نخيل البلح، شمال سيناء.

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