THE EFFECTS OF ALLELIC VARIATION FOR GLUTENIN HMW-GS ON DOUGH QUALITY IN SOME BREAD WHEAT CULTIVARS

Altinawi, E.¹; W. Al-Ek²; S. Lawand³ and A. Altaher¹

- 1- Biotechnology Dept. of the General Commission for Scientific Agric. Res. Douma, P.O. Box 113, Damascus, Syria.
- 2- Field Crops Res. Administration of the General Commission for Scientific Agric. Res. Douma, P.O. Box 113, Damascus, Syria.
- 3- Fac. of Agric., Damascus Univ., Syria.

ABSTRACT

The end-use quality of bread wheat is sensitive to cultivar (genotype) and the dominated extreme environmental conditions during the grain filling, as they effect on syntheses and accumulation of storage proteins (gliadin and glutenin) in grain. A field trials were carried out for three bread wheat cultivars (bohoth6, cham4 and cham6) at the four different environment zones in Syria for two seasons 2007 and 2008 to study the effects of the allelic variation in each studied cultivar (genotype) and effect of the dominant environmental conditions at each zone on gluten content and dough quality. Glutenin protein were analyzed by sodium dodecyl sulphate polyacrylamide gel electrophoresis SDS-PAGE that demonstrated presence some allelic variations and the absence (deletion) of some alleles at loci on the chromosomes. Three different allelic were identified at (Glu-A1 and Glu-B1) loci. The gluten content was the best in bohoth6, because there were no any allelic deletion at loci in comparison cham4 and cham6. So, the gluten content reduced when air temperature was raised above 30 °C for long duration coincident with water deficit during the grains filling period. The gluten quality was analyzed by Farinograph test (development time, stability and mixing tolerance), this test confirmed that bohoth6 has the best allelic group for syntheses and accumulation of glutenin in grains. bohoth6 at zone2 and zone4 gave a dough strength ranging from the strongest in season 2007 to the medium grade in season 2008 as a result of effect the unusual extreme climatic conditions. The results indicated that bohoth6 was more tolerant to these extreme environmental conditions than cham4 and cham6.

Keywords: Glutenin, SDS-PAGE, Gluten content, Bread wheat quality, Environmental conditions, Farinograph test and Dough quality.

INTRODUCTION

In Syria, wheat is the most important field crop. The majority of grown wheat is durum wheat (*T. durum*) and bread wheat (*T. aestivum*). Bread wheat is the most widely grown food crop in the world and distinguished for its dough quality and bread making properties. There is significant relationship between the different environmental conditions, syntheses and accumulation of storage protein, and the dough quality, which suggests that climatic conditions and weather models, climatic zones and genotype could be useful or not useful in predicting grain quality (Peterson *et al.*, 1998). The total protein content and the glutenin and gliadin ratio also affect on dough and baking properties (Uthayakumaran *et al.*, 2004). The wheat storage protein genes exhibit a co-dominant Mendelian inheritance (Gupta and Shepherd, 1990, Wrigley, 1996).

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Three genetically unlinked loci Glu-A1, Glu-B1, and Glu-D1 present on homeologous chromosomes 1A, 1B, 1D control the synthesis of the HMW glutenins (MacRitchie and Lafiandra, 2001, Gálová et al., 2002). Correlation studies (Gupta and MacRitchie, 1994) have indicated that dough 'strength' and bread-making potential of flour are positively related with subunits 5+10, 7+8, 17+8, 1 and 2* and negatively associated with subunits 6+8, 2+12 and 20 Goesaert et al., 2005; Lagrain et al., 2006. It was also observed that, where subunits 7+8 and 2* occurred together (Khatkar et al., 1996). Mir Ali, (1995) reported that the studied lines were characterized by having higher SDS sedimentation values due to the presence of subunits 17+18 at Glu-B1 when compared to those that has subunits 7+8. SDS page separation of HMW glutenins from several wheat cultivars have demonstrated a number of alleles at each loci (Payne & Lawerence, 1983; MacRitchie and Lafiandra, 2001). Although grain protein composition depends primarily on genotype, it is significantly affected by the environment factors and their interactions (Triboï et al., 2000; Daniel and Triboï, 2002; Zhu and Khan, 2001). For certain varieties, flour, dough, and baking quality parameters were reported to be altered in response to a short period of heat stress >35 °C, and some of these effects have been linked to an increased gliadins-to-glutenins ratio (Blumenthal et al., 1991, 1993; Wardlaw et al., 1995, 2002). The water deficits severely effect end-use quality for several cultivars of wheat (Guttieri et al., 2001). This study will focus on, 1- Study of the allelic variations at loci of glutenin subunits HMW-GS and their effect on total protein and gluten contents in three bread wheat cultivars and define the suitable environmental zones to each cultivar. 2- Study the effect of the deletion of some important alleles on the end-use quality (dough). Define the most tolerant cultivar to extreme environmental conditions (heat and drought stress).

MATERIALS AND METHODS

Seeds of three bread wheat cultivars Bohoth6, Cham4 and Cham6 were sown in three replicated trials at four zones with different environmental conditions for two seasons 2007 and 2008. These zones were zone1: (Ghab research center, Hama), zone2: (Himo research center, Al-Hassake), zone3: (Idleb research center, Idleb), zone4: (Malikieh research center, Al-Hassake) in each zone, the daily meteorological data was recorded (air temperature and rainfall) during the grain filling period from 15 April to 30 May (Table 1). The climatic stress was calculated as number of daily extreme temperature \geq 30 °C during the grain filling period at each zone. The grain filling period was defined based on the average maturity of cultivars grown in the trials.

Table	1:	Environr	nenta	al factor	s (the	extreme	high 1	tempei	ratures	(heat
		stress)	and	rainfall	(wate	r stress)	durin	g the	grain	filling
		period								

Peries									
The dominant climatic		Seaso	n 2007		Season 2008				
conditions	zone1	zone2	zone3	zone4	zone1	zone2	zone3	zone4	
Rainfall during filling period (mm)	97.4	35.2	80.0	50.5	37.8	3.5	28.1	17.2	
Number days (≥ 30 °C)	20	19	17	18	16	32	18	30	

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At the harvesting stage, the grains were harvested and brought to the seed biotechnology laboratory and biotechnology laboratory in General Commission for Scientific Agricultural Research, Douma, Damascus.

Near-infrared reflectance (NIR): the percentage of the total protein content (GPC) and wet gluten content (WG) and Sodium Dodecyle Sulphate sedimentation volume SDSs volume of sample grains was determined by Near-infrared reflectance (NIR) spectroscopy using an Infratec 1241 (Grain Analyzer, made in FOSS company, Tecator, Höganås, Sweden).

Milling of the Wheat flour for Farinograph test: Reduction flour was mixed to determine the dough strength of the flour by farinograph. Brabender Farinograph-E worldwide standard was used to assess the rheological properties of wheat flour in accordance (AACC 54-21). Farinograph is measured in Brabender Units (BU), which refers to the dough resistance to mixing. The Farinograph was used to identify the gluten quality and to measure dough characteristics of flours during the test such as development time (FDT) is the needed time for dough and water to mix together to develop strength, dough stability (FST), dough mixing tolerance (MT). And the Farinograph results were interrelated in comparison to the Farinograph parameters (Table 2).

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According to (Williams <i>et al</i> ., 1986).										

(Rating)	SDS sedimentation volume (ml)	FDT (min)	FST (min)	MT (BU)
Exceptionally weak	Less than 20	-	-	-
Very weak	20-29	0-2	0-2	200-250
Weak	30-39	2-4	2-4	150-199
Near of weak	40-49	-	-	-
Medium strength	50-59	4-6	4-7	100-149
Near to strong	-	6-8	7-10	-
Strong	60-69	8-10	10-15	50-99
Very strong	70-79	Over 10	Over 15	0-49
Exceptionally strong	Over 80	-	-	-

FDT, development time; FST, Dough Stability, MT, Mixing Tolerance; BU, Brabender Units.

Sodium Dodecyle Sulphate Polyacrylamid Gel Electrophoresis (SDS-PAGE): It is generally known that Gluten strength is associated with certain High Molecular Weight (HMW) Glutenin subunits. Thus, the High Molecular Weight (HMW) Glutenin composition of wheat is determined utilizing sodium dodecyle sulphate polyacrylamid gel electrophoresis (SDS-PAGE).

Preparation of Samples for SDS-PAGE Electrophoresis:

HMW-GSs were analyzed by SDS-PAGE method. Dual slab electrophoretic system Hoefer SE 600 (gel size -16×18 cm) was used for the analyses. Three grains of every cultivar were tested for four zones in two seasons 2007 and 2007. The remaining pellet after extraction of gliadin protein was extracted for the obtaining on glutenins according to a sequential procedure (Singh *et al.*, 1991). Electrophoresis of glutenin subunits was

performed on SDS-PAGE according to (peňa *et al.*, 2007). Allele identification was by using the standard wheat varieties and respective HMW-glutenin profiles were Pavon: (2*, 17+18 and 5+10), Opata: (2*, 13+16 and 2+12) Liu *et al.*, 2008, and Pitic62: (1, 7+8 and 2+12) Cornish, 2007. HMW glutenin alleles at *Glu-A1*, *Glu-B1* and *Glu-D1* loci were identified and allelic variations were rated by numbering each HMW glutenin sequentially, based on mobility in SDS-PAGE and the classification of (Payne and Lawrence, 1983).

RESULTS AND DISCUSSION

Analysis of HMW-GSs Protein: The effects of the HMW glutenin protein is evident from the results that were obtained by sodium dodecyl sulphate polyacrylamide gel electrophoresis SDS-PAGE Analysis in this study. Analysis results of the SDS-PAGE of the other cultivars demonstrated absence (deletion) of some alleles at loci on the chromosomes. Three different allelic were identified, two different alleles 2*, null (N) were corresponded to the Glu-A1 locus, and two different alleles (7+8 and 17+18), were corresponded to the Glu-B1 locus and one allele 2+12 was corresponded to Glu-D1 locus. This the last allele 2+12 was present into three studied cultivars. The HMW proteins present in three studied cultivars (cham4, cham6 and bohoth6) that were tested by using SDS-PAGE (Table 3; Figure 1 and 2).

Cultivere	HMW glutenin alleles							
Cultivars	Glu-A1	Glu-B1	Glu-D1					
Cham4	-	17+18	2+12					
Cham6	2*	7+8	2+12					
Bohoth6	2*	17+18	2+12					

Table 3: Compositions and deletion of glutenin alleles HMW-GS in three studied cultivars (cham4 cham6 and bohoth6).

SDS-PAGE analysis was used to further verify the HMW-GSs protein allelic patterns of the studied cultivars in both seasons 2007 and 2008. But, when the gels of both seasons were compared together, the obtained results by electrophoresis to determine the responsible alleles from dough strength were similarly and they didn't change when the climatic conditions were changed at four studied environmental zones in season 2008. These results indicated the subunits (alleles) don't affect by the extreme climatic conditions, viz the responsible genes in genetic loci (Glu-A1, Glu-B1 and Glu-D1) on chromosome 1 expressed despite of the dominant bad climatic conditions in season 2008 at zone2 and zone4 (Table 1). This result was similar to that reported by Dupont et al., (2006). However, each studied cultivar included 3-5 bands of HMW-GSs. Cham4 has subunit (null), 17+18 and 2+12 at loci (Glu-A1, Glu-B1 and Glu-D1), respectively. Cham6 has subunit 2*, 7+8 and 2+12 at loci (Glu-A1, Glu-B1 and Glu-D1), respectively. Also, Bohoth6 has subunit 2*, 17+18 and 2+12 at loci (Glu-A1, Glu-B1 and Glu-D1), respectively (Figure 1, 2 Table 3).

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When the comparison between loci to three cultivars was noticed that subunit 17+18 at Glu-B1 locus were present in both cham4 and bohoth6 and subunits 2+12 at Glu-D1 locus were present in all studied three cultivars. Also, subunits 7+8 at Glu-B1 locus were present in cham6 and subunits 2* at Glu-A1 locus were present in both cham6 and bohoth6 (Figure 1 and 2). However, the subunit 20 Glu-B1 loci and subunits 5+10 in Glu-D1 don't present in none from the three studied cultivars. This results indicated the HMW-GSs allelic pattern compositions in cham4 (17+18 and 2+12) are to bohoth6, and in cham6 are (2* and 2+12) are similar to bohoth6 at same locus. However, all subunits significantly affected on some of the quantitative and qualitative traits of GPC and WG contents (Table 4 and 5). So that some tests like SDSs volume and Farinograph as (FAB, FDT, FST and MT) were affected with changing both quantitative and qualitative traits of GPC and WG. The (2*, 17+18 and 7+8) are positively influenced on content of both traits and increasing of SDSs volume. This result was indicated by correlation studies to (Gupta and MacRitchie, 1994) who have confirmed that dough 'strength' and bread-making potential of flours are positively related with subunits 17+8, 7+8, and 2* and negatively associated with subunits (null) and

2+12. Also, the results in this study were similar to those by Gregova *et al.* (2006); Goesaert *et al.*, 2005; Lagrain *et al.*, 2006). Also, Branlard and Dardevet (1985) confirmed subunits 2* and 17+18 are associated with a strong dough. In general, when the alleles 2* and 17+18 are together in bohoth6, they show highest positive effects to GPC and WG. This result is explained that the wheat storage protein genes exhibit a co-dominant Mendelian inheritance (Gupta and Shepherd, 1990).

So here, present both the alleles 2* and 17+18 together showed the highest positive effects to GPC and WG, accordingly both traits effected positively on SDSs volume and farinograph test (FDT and FST) and these results were confirmed by Mir Ali (1995). Whereas, bohoth6 has subunits 17+18 that contributed to increase SDSs volume more than cham6 which has subunits 7+8 and 2+12 together. Because subunits 7+8 and 2+12 together have minimum positive effects on the mean of SDSs volume, and this also reported by Khatkar et al, (1996). Thus, in our study, cham6 that has subunits 7+8 showed minimum positive effects on the mean of SDSs volume in comparison with bohoth6 that has subunits 17+18 that expresses a higher gluten contents than subunits 7+8 in cham6. So, the deletions in the glutenin loci resulted in significantly (p<0.05) reduced the mean of GPC and WG by comparison between the means of three studied cultivars in each season alone (Table 4 and 5). The deletions to subunit 2* at Glu-A1 as in cham4, subunits 17+18 in cham6 and subunits at Glu-B1 reduced the quality of GPC and WG in flours of both cultivars, because subunit 2* and subunits 17+18 affect positively on the dough strength. So, bohoth6 had a strong dough in its flour, because it has at both loci (Glu-A1 and Glu-B1) the positive subunits 2* and 17+18 that result in a high dough strength. Gupta and MacRitchie (1994) have indicated that dough 'strength' and bread-making potential of flour are positively related with subunits 7+8, 17+8 and 2*. Anyhow, the Farinograph tests had indicated significant effects of GPC and WG contents on dough strength at four studied environmental zones in both the seasons 2007 and 2008. Cham4 which has a the deletion (null) and 17+18, cham6 which has subunits 2* and 7+8, and bohoth6 which has subunits 2* and 17+18. These cultivars in season 2007 were affected by the usually extreme climatic conditions at four studied zones, so cham4 and cham6 produced medium GPC and WG, but bohoth6 produced high GPC and WG (Table 4 and 5). While, GPC and WG in Cham4 and cham6 in season 2008 lowed to medium, and GPC and WG in bohoth6 lowed to the medium dough according to Farinograph tests (FDT, FST and MT) and SDSs volume and the studied cultivars means (Table 2, Figures 4, 6, 8, and 10). Because they were affected by the unusually extreme climatic conditions (Table 1). In fact, presence of the subunits 17+18 and 2+12 together in the genomic material of bohoth6 resulted in stronger dough than the dough which has subunits 7+8 and 2+12 together as in cham 6, perhaps because the subunits 17+18 are responsible of synthesis glutenin proteins more than subunits 7+8 as indicated by Mir Ali (1995). The deletion in cham4 to subunit 2* at Glu-A1 locus negatively effected on the GPC and WG of flour, after that on the dough and bread quality. Because the deletion changes the composition of GPC and WG during duration synthesis and accumulate total proteins (gliadin and

glutenin) during the grain filling period and consequently the end-use quality (Table 4 and 5). Shwery *et al.*, (2003) demonstrated that allelic differences among genotypes affect the amounts and the properties of HMW-GS polymers of wheat gluten and the bread making properties of individual genotypes. Therefore, the deletion some alleles of glutenin subunits at Glu-A1 and Glu-B1 loci impacted negatively on the parameters of Farinograph test FDT, FST and MT and SDSs volume (Table 3 and figures 3, 5, 7 and 9). The dough mixing strength was significantly reduced, because the deletion of subunits at Glu-A1 loci (null) and Glu-B1 loci (Table 3).

The low amount of glutenins formed a weak gluten network was not able to extend itself, because it has the weak structure causes reduction to the dough stability and the network ruptures quickly. These results were agreed with (Branlard and Dardevet, 1985; MirAli, 1995; Gregova *et al.* (2006); Goesaert *et al.*, 2005; Lagrain *et al.*, 2006). Also, many studies reported that when some HMW glutenins are deleted, the dough mixing strength and bread making quality will be reduced (Lawrence *et al.*, 1988; MacRitchie and Lafiandra, 2001). The stability FST in bohoth6 in both seasons (Figure 5 and 6). This results were similar to many results obtained by (Weegels *et al.*, 1994; Lafiandra *et al.*, 1999; Branlard *et al.*, 2001).In both seasons, the means of GPC and WG in bohoth6 were better than the means in deleted cham4 and cham6 despite that the means of GPC and WG were reduced in season 2008.

	zones.					
Cultivars	Season	zone1	zone2	zone3	zone4	Cultivar mean
Cham4		13.01Cc±	16.0Bc±	11.9Dc±	15.1Bc±	14.51d±
		0.515	2.006	0.885	0.452	0.348
Cham6		14.01Db±	17.01Ab±	14.80Da±	16.1Ad±	14.85C±
	2007	0.137	0.154	1.668	0.201	0.001
Roboth 6	2007	14.6Ca±	18.01Aa±	12.7Db±	16.5Ba±	15.20a±
Bonotino		0.377	1.852	0.782	0.251	0.349
Means		13.87±	17.01±	13.13±	15.90±	
		0.980	1.653	1.720	1047	
Cham4		13.05Bc	12.01Dc	13.15Ac ±	11.90Dc	12.35
Cham4		±0.545	±0.149	0.312	±0.138	±0.722
Chame		14.40Ab±	12.25Cb±	14.20Bb ±	12.25Cb±	13.46±
Chanto	2009	0.178	0.718	0.111	0.139	0.094
Boboth6	2000	15.50Aa±	12.50Da±	15.20Ba ±	13.30Ca±	14.13±
Donotho		0.367	0.569	0.201	0.001	0.816
Moone		14.32±	12.25±	14.18±	12.48±	
Wearts		1.008	1.056	0.874	0.826	
Gra	Grand mean			14.138		
LSD 0.05	Cultivar X zone			0.035		
	CV%			0.2		

Table 4: Analysis of variances of total protein content (GPC) content in three studied cultivars at four environmental zones in two studied seasons 2007 and 2008 and the means of cultivars and zones.

LSD, Least significant difference of the means (5% level); CV, Coefficients of variation.The numbers with superscript capital letters (A, B, C and D) refer to significant variation among varieties, while numbers with superscript small letters (a, b, c and d) refer to significant variation among zones.

	0000			ine meane	el ealtra				
Cultivars	Season	zone1	zone2	zone3	zone4	Cultivar means			
Cham4		32.1Dc±	39.14Ab±	34.8Bb±	34.6Cc ±	35.16±			
		0.01	1.837	0.81	2.657	2.177			
Cham6		35.1Db±	37.4Cc±	39.5Ba±	40.9Ab ±	38.23±			
	2007	0.055	2.968	2.445	0.578	0.888			
Bohoth6	2007	35.6Ca±	41.9Ba±	34.2Dc±	42.8Aa ±	38.63±			
		0.045	1.132	3.255	2.078	1.288			
Moone		34.27±	39.48±	36.17±	39.43±				
IVICALIS		3.070	2.143	1.17	2.097				
Cham4		33.2Ab	24.01cD	32.01Bc±	31.55Cc±	30.19±			
		±0.652	±0.258	0.345	0.565	0.450			
Cham6		32.5Bc±	24.25bC	32.5Bb	33.15Aa±	30.60 ±			
	2000	0.456	±0.091	±0.2625	0.628	0.078			
Bohoth6	2000	33.4Ba±	24.45aD	34.01Aa	33.10Cb±	31.24 ±			
		0.196	±0.349	±0.608	0.062	0.562			
Moone		33.03±	24.24±	32.84±	32.60±				
IVICALIS		2.356	6.441	2.1625	1.9225				
Grand	mean	33.961							
LSD	0.05	0.194							
Cultivar	X zone			0.104					
CV	%			0.35					

Table	5:	Analysis	of	variances	of	wet	gluten	content	(WG)	in	three
		studied	cult	ivars at fo	ur	envir	onment	al zones	in two) st	udied
		seasons	200	07 and 2008	3 ai	nd th	e means	s of cultiv	vars an	d z	ones.

The interactions of subunit 2* with subunits 17+18 in bohoth6 resulted in the best composition and higher contents of the GPC and WG than cham4 that has subunits 17+18, and cham6 that has only subunit 7+8. These results in our study are a strong evidence on co-dominant to both subunit 2* and subunits 17+18 together. Therefore, bohoth6 which has subunits 2*, 17+18 had larger means of GPC and WG than both of the cultivars cham4 and cham6 that have (subunits 17+18, subunits 2* and 7+8) respectively. Peña *et al.*, (1995) confirmed common wheat varieties possessing five HMWG subunit components generally have stronger gluten character than the ones possessing three or four components. Too, the results in this study indicated bohoth6 has next subunits 2*, 17+18 and 2+12 at (Glu-A1, Glu-B1 and Glu-D1) respectively, whereas it express the higher glutenin percentage into the extreme climatic conditions (high temperatures and water deficit). Also, other similar results to our results were confirmed by (Mir Ali, 1995; Gregova *et al.*, (2006); Goesaert *et al.*, 2005; Lagrain *et al.*, 2006).

The variances of GPC and WG means at studied environmental zones were noticed (Table 4 and 5). In season 2007 at zone2 and zone4 the means of GPC were 17.01% and 15.9% respectively, and WG were 39.48% and 39.43% respectively, the means of GPC at zone2 and zone4 were higher than the means 13.87% and 13.13% of both zone1 and zone3 respectively, Also, the means of WG at zone2 and zone4 were 34.27% and 36.17% of both respectively, (Table 4 and 5), despite there wasn't any significant difference between the means at zone2 and zone4. But, in season 2008 the means of GPC at zone2 and zone4 were 12.25% and 12.48% respectively, they were lower than the means 14.32% and 14.18% of both zone1 and zone3, respectively. But, the means of WG at zone2 and zone4 were 24.24%

and 32.6% respectively, they were lower than the means 33.03% and 32.84% of both zone1 and zone3 respectively (Table 4 and 5). Depend on this the results confirmed that the extreme environmental conditions (heat and water stress) that was dominant at zone2 and zone4 in season 2008 affected on syntheses and accumulations of total protein and gluten (storage proteins), because storage proteins accumulate from approximately 6 day to the end of grain-filling (Panozzo et al., 2001). Also, the extreme environmental conditions that was dominant in season 2008 at zone2 and zone4 maybe resulted in shorten the grains filling period. Jamieson et al., (2001) has confirmed such as the fact, whereas reported that high temperature and/or drought effect the balance of protein fractions. Was reported by Blumenthal et al., (1993) that for certain varieties, flour, dough, and baking quality parameters are altered in response to a short period of heat stress >35 °C. and some of these effects have been linked to an increased gliadins-toglutenins ratio (Blumenthal et al., 1991) and decreases in the proportion of the high molecular weight glutenins (HMW) (Wardlaw et al., 2002). So that, the means of GPC% and WG% in all studied zones in season 2007 were high, but they decreased in season 2008 in generally (Table 4 and 5). Was noticed that the decreasing average of the means GPC and WG when the comparison between the means at zone1 and zone3 in both seasons was very little. While the decreasing average of the means of GPC and WG in zone2 and zone4 in both seasons was very high when the comparison between these means. Anyway, the percentage of GPC and WG in bohoth6 at zone2 and zone4 were ((12.5% and 13.30%), (24.45% and 33.15%), respectively higher than the percentages of GPC and WG in both cultivars cham4 and cham6, whereas the percentages of GPC in cham4 were ((12.01% and 11.9%), respectively and to WG were (24.01% and 31.55%)) respectively, and cham6 were ((12.25% and 12.25%), respectively and to WG were (24.25% and 33.15%)), respectively (Table 4 and 5). Thus, this results confirmed that bohoth6 is distinguished from cham4 and cham6, because the first cultivar has the improving alleles to dough strength at both Glu-A1 and Glu-B1 loci and it is more tolerant to extreme environmental conditions as that was dominant at zone2 and zone4 in season 2008. In this results, the interaction between cultivar (genotype) and the extreme environmental conditions indicated bohoth6 was the distinguished cultivar. Other studies similar to our study reported that although grain protein composition depends primarily on genotype, it is significantly affected by environment factors and their interactions (Triboï et al., 2000; Zhu and Khan, 2001).



CONCLUSION

This study indicated that bohoth6 has the subunits 2*, 17+18 and 2+12 at (Glu-A1, Glu-B1 and Glu-D1) respectively, and cham6 has the subunits 2*, 7+8 and 2+12 at (Glu-A1, Glu-B1 and Glu-D1) respectively, so cham6 expressed lower glutenin than bohoth6, but it was higher than cham4 which has the subunits 17+18 and 2+12 at (Glu-B1 and Glu-D1) respectively. The deletions in the alutenin loci resulted in significant (p<0.05) reduction of the mean of GPC and WG by comparison between the means of three studied cultivars and bohoth6 was the best one cham4 and cham6 that have the deletion at their loci. Also, bohoth6 had a strong dough result to the increasing of the GPC and WG content. Farinograph test and SDSs volume had indicated a presence to significant effects of the GPC and WG contents on dough strength at four studied environmental zones in both seasons. Bohoth6 was more tolerant to these extreme environmental condition than cham4 and cham6, so it gave the highest content of GPC and WG and best dough strength at same both zones (zone1 and zone2). Thus suggests that bohoth6 can perform better than cham4 and cham6 if it grows in regions of Syria having similar extreme environmental conditions to the one dominated in both zone2 and zone4 during season 2008 season.

REFERENCES

- Blumenthal, C.S.; E.W.R. Barlow and C.W. Wrigley (1993). Growth environment and wheat quality: the effect of heat stress on dough properties and gluten proteins. J Cereal Sci 18: 3–21.
- Blumenthal, C.S.; I.L. Batey; F. Bekes; C.W. Wrigley and E.W.R. Barlow (1991). Seasonal changes in wheat-grain quality associated with high temperatures during grain filling, Australian Journal of Agricultural Research 42:1. 21 – 30.
- Branlard, G. and M. Dardevet (1985). Diversity of grain protein and bread wheat quality. II. Correlation between high molecular subunits of glutenin and flour quality characteristics. J. Cereal Sci. 3: 345-354.
- Branlard, G.; M. Dardevet; R. Saccomano; F. Lagoutt and J. Gourdon (2001). Genetic diversity of wheat storage proteins and bread wheat quality. Euphytica 119, 59–67.
- Daniel, C. and E. Triboï (2002). Changes in wheat protein aggregation during grain development: effects of temperatures and water stress. European Journal of Agronomy 16, 1–12.
- Dupont, F. M.; W. J. Hurkman; W. H. Vensel; C. Tanaka; K. M. Kothari; O.K. Chung and Altenbach (2006). Protein accumulation and composition in wheat grain: effects on S. B. mineral nutrients and high temperature. Eur. J. Agron., 25:96-107.
- Gálová, Z.; I. Michalik; H. Knoblochova, and E. Gregova (2002). Variation in HMW glutenin subunits of different species of wheat. Rostilinna Vyroba. 48: 15-19.
- Goesaert, H.; K. Brijs; W.S. Veraverbeke; C.M. Courtin; K. Gebruers and J.A. Delcour (2005). Wheat flour constituents: how they impact bread quality, and how to impact their functionality, Trends Food Sci. Tech. 16:12-30.

- Gregova, E.; J. Hermuth; J. Kraic and L. Dotlacil (2006). Protein heterogeneity in European wheat landraces and obsolete cultivars. Additional information II. Genetic Resour. Crop Evol., 53: 867–71.
- Gupta, R. B. and F. MacRitchie (1994). Allelic variation at glutenin subunit and gliadin loci Glu-1, Glu-3 and Gli-1 of common wheats. II. Biochemical basis of the allelic effects on dough properties. J. Cereal Sci. 19:19-29.
- Gupta, R.B. and K.W. Shepherd (1990). Two-step one-dimensional SDS-PAGE analysis of LMW Subunits of glutenin. 1. Variation and genetic control of the subunits in hexaploid wheats. Theoretical and Applied Genetics, 80:65-74.
- Guttieri, M.J., J.C. Stark; K. O'Brien and E. Souza (2001). Relative sensitivity of spring wheat grain yield and quality parameters to moisture deficit. Crop Science 41, 327–335.
- Jamieson, P.D.; P.J. Stone and M.A. Semenov (2001). Towards modelling quality in wheat—from grain nitrogen concentration to protein composition. Aspects of Applied Biology 64, 111–126.
- Khatkar, B.S.; A.E. Bell and J.D. Schofield (1996). A Comparative Study of the Inter-Relationships - Between Mixograph Parameters and Bread-Y I Making Qualities of Wheat Flours and Glutens. J Sci. Food Agric 1996, 72:71-85.
- Lafiandra, D.; S. Masci; C.S. Blumenthal and C.W. Wrigley (1999). The formation of glutenin polymer in practice. Cereal Foods World 44, 572–578.
- Lagrain, B.; K. Brijs and J.A. Delcour (2006). Impact of redox agents on the physicochemistry of wheat gluten proteins during hydrothermal treatment, J. Cereal Sci. 44, pp 49-53.
- MacRitchie, F. and D. Lafiandra (2001). The use of near-isogenic wheat cultivars to determine protein composition-FDSnctionality relationship. Cereal Chem. 78(5): 501-506.
- Mir Ali, N. (1995) .Performance of high-protein mutant lines of Triticum aestivum (L.) under semi-arid conditions of Syria Field Crops Research, Volume 41:2, 101-108.
- Panozzo, J.F.; H.A. Eagles and M. Wootton (2001). Changes in protein composition during grain development in wheat. Australian Journal of Agricultural Research 52, 485–493.
- Payne, P. I. and G. J. Lawerence (1983). Catalogue of alleles for the complex gene loci, Glu-A1, Glu-B1, Glu-D1 which code for high-molecular-weight subunits of glutenin in hexaploid wheat. Cereal Res. Communs. 11: 29-35.
- Peña, R.J.; J. Zarco-Hernandez, and A. Mujeeb-Kazi, (1995). Glutenin subunit compositions and breadmaking quality characteristics of synthetic hexaploid wheats derived from Triticum turgidum x Triticum tauschii (coss.) Schmal Crosses, Journal of Cereal Science, 21:1, 15-23.
- Peterson, C.J.; R.A. Graybosch; D.R. Shelton and P.S. Baenziger (1998), Baking quality of hard winter wheat: Response of cultivars to environment in the Great Plains, Euphytica 100: 157–162.
- Shwery, P.R.; N.G. Halford and D. Lafiandra (2003). Genetics of wheat gluten proteins. Advances in Genetics. 49: 111-184.
- Triboï, E.; A. Abad; A. Michelena; J. Lloveras; J.L. Ollier; C. Daniel (2000). Environmental effects on the quality of two wheat genotypes. 1. quantitative and qualitative variation of storage proteins. European Journal of Agronomy, 13, pp 47–64.

- Uthayakumaran, S.; I.L. Batey; and C.W. Wrigley (2004). On-the-spot identification of grain variety and wheat-quality type by Lab-on-chip capillary electrophoresis. J. Cereal Sci. 41: 371-374.
- Wardlaw, I.F. and L. Moncur, (1995). The response of wheat to high temperature following anthesis: I The rate and duration of kernel filling. Aust J Plant Physiol 22: 391-397.
- Wardlaw, I.F., Blumenthal C., Larroque O., Wrigley C.W. (2002). Contrasting effects of chronic heat stress and heat shock on kernel weight and flour quality in wheat. Functional Plant Biology 29, 25–34.
- Weegels, P.L., R.J. Hamer and J.D. Schofield (1994). Functional properties of wheat glutenin. J. Cereal Sci. 23: 1-18.
- Williams, P.H.; F. Jaby El-Haramein; H. Nakkoul and S. Rihawi (1986). Crop quality evaluation methods and guidelines, International center for agricultural research in the dry zones, Technical manual No.14, pp 2-31. Wrigley, C.W. (1996). Gaint proteins with flour power. Nature. 381: 738-739.
- Zhu, J. and K. Khan (2001). Effects of genotype and environment on glutenin polymers and breadmaking quality. Cereal Chemistry, 78, 125–130.

تأثير التغيرات الأليلية للجلوتينين على نوعية العجين لبعض أصناف القمح الطري عماد التيناوي ، وليد العك ، سلام لاوند و عبد الله الطاهر ١- قسم التقانات الحيوية في الهيئة العامة للبحوث العلمية الزراعية-دوما-دمشق-سورية. ٣- للية الزراعة-جامعة دمشق-سورية.

تعتبر نوعية الناتج النهائي للقمح الطري حساسة للصنف والظروف البيئية السائدة خلال فترة امتلاء الحبة حيث يؤثران على اصطناع وتكدس بروتينات التخزين في الحبة. نفذت الاختبارات الحقلية لثلاثة أصناف قمح طري (بحوث ، شام ٤ وشام ٢) في أربع مناطق مختلفة بيئياً لموسمين زراعيين ٢٠٠٧ و٢٠٠٨ لدراسة تأثيرات التغيرات الأليلية في كل صنف مدروس وتأثيرات الظروف البيئية السائدة في كل منطقة على محتوى الجلوتين ونوعية العجين. حللت بروتينات التخزين بطريقة BDS-PAGE التي أوضحت وجود بعض محددين في المواقع وغياب بعض الأليلات من مواقعها على الكروموزومات. الأليلات الثلاثة المختلفة كانوا أي حذف أليلية و غياب بعض الأليلات من مواقعها على الكروموزومات. الأليلات الثلاثة المختلفة كانوا محددين في المواقع (Bure 1 معرفي النائية معار المعام على الكروموزومات. الأليلات الثلاثة المختلفة كانوا أي حذف أليلي في مواقعه الوراثية معارنة مع الشام ٤ والشام ٦. أيضاً انخفض محتوى الجلوتين عندما ارتغعت درجة الحرارة إلى ٣٠م وما فوق لفترة طويلة مترافقة مع نقص الماء خلال فترة امتلاء الحبة. أجريت تحاليل باختبارات الفارينوغراف (وقت تطور العجينة وثبات العجين والقدرة على تحرا الحبوب أعلي الحقوال باختبارات الفارينوغراف (وقت تطور العجينة وثبات العجين والقدرة على تحمل العجن)، أثبت هذا الاختبار أن المعوث البينوغراف (وقت تطور العجينة وثبات العجين والقدرة على تحمل العجن)، أثبت هذا الاختبار أن المنطقين البيئيتين قوة عجين تراوحت بين القوة في الموسم ٢٠٠٧ إلى المتوسطة في الموسم ٢٠٠٨ نتيجة المنطقين البيئيتين أو عجين تراوحت بين القوة في الموسم ٢٠٠٧ إلى المتوسطة في الموسم ٢٠٠٨ نتيجة المنطقين المحتويزين شام ٤ ورام عنوالا عنيادية. بينت النتائج أن الصنف بحوث أكثر تحملا للجفاف من

الكلمات المفتاحية: جلوتينين، SDS-PAGE، محتوى الجلوتين، نوعية القمح الطري، الظروف المناخية، اختبار الفارينوغراف ونوعية العجين.

- قام بتحكيم البحث
- أ.د / مسعد عبد العزيز أبو رية
 أ.د / خالد على أبو شادى

كلية الزراعة – جامعة المنصورة مركز البحوث الزراعية