EFFECT OF SOME GROWTH REGULATORS AND ANTIOXIDANTS ON GROWTH, YIELD AND SEED CHEMICAL COMPOSITION OF FABA BEAN PLANTS Ismaeil, Faten H. M. and M. M. M. Abd El-All Retenue Den See of Agric Banka Univ

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ABSTRACT

This study was carried out at Experimental Station of Agricultural Botany Department, Faculty of Agriculture, Moshtohor, Benha University during 2009 and 2010 seasons.

All applied growth substances significantly increased morphological characteristics, nodules number formation, pollen fertility, yield, content of N, P, K, crude protein and carbohydrates in seeds of faba bean plant.

The treatments of NAA at 200 or 100 ppm and salicylic acid at 100 or 50 ppm followed by pp333 at 10 or 5 ppm and Ascorbic acid at 100 or 50 ppm were more effective comparing in this respect. Also, the same treatments exhibited the lowest percentages of flowers and pods abscission.

In addition, NAA at 200 ppm and salicylic acid at 100 ppm gave the lowest percentage of abscission, pollen sterility while gave the highest seeds yield as well as significant differences in the anatomical characteristics.

Keywords: Faba bean, NAA, salicylic acid, Paclobutrazol, Ascorbic acid, anatomy and flower abscission.

INTRODUCTION

Faba bean is an important legume crop as a major source of protein and occupies large area of cultivated land in Egypt. Cultivation of faba bean leads to increase of soil nitrogenous compounds (Hungria and Vargas, 2000).

Field crop production in Egypt is concentrated mainly on the arable and around the banks of river Nile and the intensive agriculture system may leads to decrease the productivity of unit-area. Reclamation, cultivating new lands and adopting the most promising agronomic practices are the most important factors for increasing the productivity to close the gap between consumption and production of food crops.

High demand of food in Egypt implies more production of food crops, including protein-containing crops, i.e. leguminous ones. Faba bean *(Vicia faba,* L.) is one of the major field crops grown in Egypt; it is an important source of protein for human and animal consumption and it plays a role in the crop rotation. However, the total production of this crop is still insufficient to cover the local consumption. (Khafaga *et al.,* 2009)

Rhizobium plays a very important role in agriculture by inducing nitrogen-fixing nodules on the roots of legumes such as peas, beans, clover and alfalfa (Downie and Brewin, 2007). Therefore, this symbiosis can relieve the requirements for added nitrogenous fertilizer during the growth of leguminous crops. Flower abscission occurs both before and after fertilization. In some species the mere lack of pollination, after a critical period, activates the abscission zones, and in other species the lack of fertilization, again after a critical period, does so. Unfertilized flowers often abscise due to competition for carbohydrates (Aloni *et al.*, 1996).

Salicylic acid (SA) is an endogenous plant growth regulator. It is involved in various physiological processes of plant growth and development (Klessig and Malamy, 1994) such as induction of flowering and root growth stimulation (Gutiérrez-Coronado *et al*, 1998). SA also plays a major role during the early stages of Rhizobium-legume symbiosis (Rasmussen *et al*, 1991).

Paclobutrazol increased chlorophyll content, this may be partly due to the observed increase in mass of the root system which is the major site of cytokinin biosynthesis. The increase in cytokinin levels was associated with stimulated chlorophyll biosynthesis (Fletcher *et al.*, 2000).

Ascorbic acid, also named vitamin C or ascorbate, is an antioxidant and enzyme co-factor that has multiple functions in plants (Ishikawa *et al.,* 2006).

This present study was designed to investigate the effect of soaked and foliar spray of NAA, pp333, salicylic acid and ascorbic acid on enhancing the growth, nodules number, anatomical studies(stem & leaf), flower abscission, pollen fertility, yield and chemical composition in seeds of faba bean plant.

MATERIALS AND METHODS

This study was carried out at Experimental Station of Agricultural Botany Department, Faculty of Agriculture, Moshtohor, Benha University during 2009 and 2010 seasons.

This investigation aimed to study the response of faba bean plant (*Faba vulgaris* Mill.) Cv. Gridley to seed soaking and foliar spraying with some growth regulators and antioxidants. Seeds were soaked for 3 hours in the assigned concentrations as follow:-

1-The control (distilled water).

2-Naphthaline acetic acid (NAA) at 100 and 200 ppm.

3-Pacloputrazol (PP₃₃₃) at 5 and 10 ppm.

4- Salicylic acid(SA) at 50 and 100 ppm.

5- Ascorbic acid (AsA) at 50 and 100 ppm.

The seeds were sown in 25th of October during 2009-2010 seasons. Plants were sprayed with the same treatments (with 1ml/l of Tween 20 as a wetting and spreading agent) two times:(i.e., vegetative growth stage and the flowering stage).

the following measurements were recorded:-

Growth Parameters:

Root length(cm.), nodules number/ plant, plant height, number of leaves/ plant, total leaf area(cm²)/ plant following the method described by Deriaux *et al.* (1973), dry weights (g / plant) of shoots and roots were

recorded (90 days after sowing). Shoots and roots were dried in an electrical oven at 75 °C till constant dry weight to determine dry weights.

Photosynthetic pigments:-

Chlorophyll a, b and carotenoids were Colorimetrically determined in leaves of faba bean plants at 60 and 85 days from sowing according to the methods described by Nornal(1982).

Anatomical studies

The samples of stem and leaf were taken from the 4th internode from top of the main stem from high concentration of all treatments at 85 days after sowing. The samples specimens were taken then killed and fixed in FAA (5ml. formalin, 5ml. glacial acetic acid and 90ml. ethyl alcohol 70%), washed in 50% ethyl alcohol, dehydrated in series of ethyl alcohols 70,90,95 and 100%, infiltrated in xylene, embedded in paraffin wax with a melting point of 60-63°C, sectioned to 20 microns in thickness (Sass 1951), stained with the double stain method (fast green and safranin), cleared in xylene and mounted in Canada balsam (Johanson, 1940). Sections were read to detect histological manifestation of noticeable responses resulted from other treatments.

The prepared section were microscopically examined, counts and measurements (μ) were taken using a micrometer eye piece. Average of readings from 3 slides/treatment were calculated .

flowering characters:-

a)Total number of flowers / plant:

The total number of the opened flowers per plant through the season were recorded for each treatment of faba bean plants.

b) Abscission percentage: was calculated according to the equation:

No. of flowers/plant -No. of fruits/plant X 100 Abscission %= No. of flowers / plant

c) Pollen grains fertility:

Pollen fertility was estimated by the inspection and counting of fertile and non-fertile pollen grains mounted in dilute iodine solution and microscopically examined using the method described by Shahine (1961).

d) Pods setting percentage:

was calculated according to the following equation:

| | No. of pods / plant | X (00 |
|--------------------|------------------------|----------|
| % of setted pods = | No. of flowers / plant | —— X 100 |

yield and yield components:-

Number of setted pods/ plant , weight of pod(g), pods yield(g)/ plant, weight of seeds/ pod, seeds yield(g)/plant and seed index(weight of 100 seeds (g)) were recorded in the harvest sample(120 days after sowing). Chemical analysis:

Seeds at the harvest (120 days after sowing) were used to determine the following chemical analysis during 2010 season.

-Total nitrogen percentage was determined in the dried seeds by using wet digestion according to Piper (1950), using microkjeldahl method as described by Horneck and Miller (1998). Crude protein = total nitrogen x 6.25 (A. O. A. C., 1990), Phosphorus was determined colorimetrically according to the method of Sandell (1950), Potassium content was determined by flame photometer according to Horneck and Hanson (1998), Total carbohydrates content was determined by using phenol-sulphoric acid method described by Dubois *et al.*,(1956)

Statistical analysis:

All data obtained during both seasons of study were subjected to analysis of variance and significant differences among means were determined according to Snedecor and Corchran (1972). Significant differences among means were distinguished according to the Duncan's multiple range test (Duncan, 1955).

RESULTS AND DISCUSSION

Growth Parameters:

Data in Table (1) revealed that all applied treatments were significantly increased the root length, No. of nodules number/ plant, plant height, No. of branches/plant No. of leaves/plant, total leaf area/ plant and dry weight of root, stem and leaves compared the control. Maximum increments were obtained by applying 200 and 100 ppm of NAA followed by 100 and 50 ppm of salicylic acid and 100ppm of ascorbic acid treatments in 2009 and 2010 seasons.

Gutam *et al.*,(2009) stated that Naphthalene acetic acid(NAA) is a major factor has a stimulating effect on cell division and cell elongation, hence it could increase all vegetative growth and photosynthetic rates(Table, 2).

El-Shraiy and Hegazi(2009) showed that the maximum number of nodules per plant was achieved by the application of SA at 10⁻³ M. Whereas PP₃₃₃ applications prevent nodule formation.

In this connection, Sakhabutdinova, *et al.*, (2003) mentioned that SA at 0.05 mM increased the level of cell division within the apical meristem of seedling roots which caused an increase in plant growth. SA treatment caused accumulation of both ABA and IAA in wheat seedlings.

Also, results showed that foliar application of paclobutrazol (10 ppm) recorded the highest values of number of branches/plant, No. of leaves/plant and dry leaves weight/plant. Meanwhile, paclobutrazol at 5 or 10ppm reduced plant height by inhibiting GA3 production which is responsible for cell elongation(Grossmann,1992)

Paclobutrazol (200 ppm) caused marked the highest values of fresh and dry weights/plant, leaf area, number of pods/plant, number of seed/pod, 100 seed weight and seed yield (Khafaga *et al.*, 2009).

Photosynthetic pigments:-

Data in Table (2) showed that photosynthetic pigments content in leaves (i.e., chlorophyll a & b, Chl.(a+b) and Carotenoids) were increased with pp₃₃₃ at 10 or 5ppm, NAA at 200 or 100ppm and salicylic acid at 100 or 50ppm in plants aged 60 and 85 days in two seasons of 2009 and 2010.

Also, it could be noticed that paclobutrazol gave the highest values in chlorophyll content, this may be partly due to the observed increase in mass of the root system which is the major site of cytokinin biosynthesis. The increase in cytokinin levels was associated with stimulated chlorophyll biosynthesis (Fletcher *et al.*, 2000).

This observation was supported by Sakhabutdinova, *et al.*, (2003) who reported that, leaves treated with high concentrations of SA (5 mM) accumulated more ChI and carotene isomers. Leaves treated with SA enhanced SOD activities and accumulated H_2O_2 before the detection of the SA-mediated inactivation of H_2O_2 -degrading enzymes.

Moreover, Ismaeil, (1995) and Fletcher *et al.*, (2000) found stimulating effects of paclobutrazol on the photosynthetic pigments in some other plants.

| Table | (2): | Effect | of | some | growth | re | gulators | and | anti | oxidan | ts on |
|-------|------|--------|------|----------|----------|-----|----------|-------|------|--------|-------|
| | | photo | syr | nthetic | pigmen | Its | concentr | ation | of | faba | bean |
| | | leave | s dı | uring 20 | 010 seas | on. | | | | | |

| Characters | Photosynthetic pigments mg/g fresh weight | | | | | | | | | |
|----------------------------|---|------|------|----------|---------|---------|-------------|----------|--|--|
| | Chl. | A | Chl. | B Chl. A | | + b | Carotenoids | | | |
| | 60 | 85 | 60 | 85 | 60 dava | 05 | 60 dava | 05 davia | | |
| Treatments | days | days | days | days | ou days | oo days | 60 days | ob days | | |
| Control(distilled water) | 0.67 | 0.69 | 0.36 | 0.40 | 1.03 | 1.09 | 0.18 | 0.28 | | |
| NAA at 100 ppm | 1.39 | 1.44 | 0.85 | 0.82 | 2.24 | 2.26 | 0.46 | 0.51 | | |
| 200 ppm | 1.44 | 1.48 | 0.87 | 0.88 | 2.31 | 2.36 | 0.48 | 0.58 | | |
| PP ₃₃₃ at 5 ppm | 1.50 | 1.53 | 0.93 | 0.96 | 2.43 | 2.49 | 0.56 | 0.64 | | |
| 10 ppm | 1.57 | 1.56 | 0.95 | 0.99 | 2.52 | 2.55 | 0.59 | 0.66 | | |
| Salicylic acid at 50 ppm | 1.34 | 1.40 | 0.69 | 0.70 | 2.03 | 2.10 | 0.43 | 0.48 | | |
| 100 ppm | 1.37 | 1.43 | 0.74 | 0.77 | 2.11 | 2.20 | 0.45 | 0.51 | | |
| Ascorbic acid at 50 ppm | 1.22 | 1.24 | 0.62 | 0.69 | 1.84 | 1.93 | 0.33 | 0.40 | | |
| 100 ppm | 1.33 | 1.31 | 0.64 | 0.69 | 1.97 | 2.00 | 0.35 | 0.43 | | |

Anatomical studies:-

Stem anatomy:-

Table (3) and Fig. (1) show that the highest increase was existed with the application of pp_{333} at 10 ppm. These increase was 6552.32µ compared with the 3561.83µ that of the control.

The increase in the stem diameter was due to the increase in both hollow pith and stem wall(cuticle , epidermis, collenchyma, parenchyma, vascular bundle and parenchymatous pith thickness).

As for the total number of vascular bundles, all treatments increased them. The highest increase in this number was with the treatment of pp₃₃₃ at 10 ppm. Also, the applied treatments increased phloem & xylem thickness and no. of xylem vessels.

Hence, of interest to note that different applied treatments increased stem diameter that reached to maximum with the treatment of NAA at

 $200ppm(6250.54\mu)$ followed by salicylic acid at 100 $ppm(5574.80\mu)$ and ascorbic acid at 100 $ppm(5574.80\mu).$

In general, the stimulatory effects of applied treatments upon the anatomical features of treated plants could be attributed to the effect upon cambium activity. Increment of cambium activity could mainly attributed to the increase of endogenous hormones level especially auxins and cytokinins(Ismaeil, 1995 and Zewail, 2011) confirming the findings of the present study.

leaf anatomy:

Data in Table (4) revealed that all applied treatments significantly increased the anatomical features of faba bean leaf

With regard to the blade thickness, it was increased with different used treatments to reach its maximum value (622.80 μ) with PP333 at 10 ppm followed by Salicylic acid at 100 ppm (558.75 μ) and NAA at 200 ppm (541.13 μ) compared with the control(403.50 μ).

Table (3): Effect of some growth regulators and antioxidants on the mean counts and measurements of certain anatomical features of main stem of faba bean plants at 85 days after sowing during 2010 seasons.

| Treatments | Control | NAA at | PP ₃₃₃ at 10 | Salicylic | Ascorbic | |
|---|----------------------|---------|-------------------------|--------------------|--------------------|--|
| Histological Characteristics (μ) | (distilled water) | 200 ppm | ppm | acid at 100 ppm | acid at 100 ppm | |
| Stem diameter | 3561.83 | 6250.54 | 6552.32 | 5574.80 | 4530.85 | |
| Cuticle layer thickness | 11.25 | 13.28 | 17.10 | 12.60 | 11.70 | |
| Epidermal thickness | 36.90 | 50.18 | 53.33 | 46.05 | 42.30 | |
| Thickness of collenchyma layers | 62.10 | 90.00 | 97.65 | 86.40 | 85.50 | |
| Thickness of parenchyma layers | 156.15 | 281.03 | 283.50 | 237.60 | 180.00 | |
| No. of vascular bundles | 22 | 28 | 29 | 25 | 24 | |
| Thickness of fibers layers | 126.90 | 258.30 | 319.95 | 252.40 | 150.30 | |
| Thickness of phloem layers | 79.88 | 119.25 | 119.48 | 115.65 | 100.00 | |
| Cambium region thickness | 37.35 | 88.43 | 90.90 | 72.15 | 60.45 | |
| Xylem thickness in vascular bundle | 246.51 | 328.50 | 353.25 | 304.05 | 248.85 | |
| Length of vascular bundle. | 490.64 | 794.48 | 880.58 | 744.25 | 559.60 | |
| No. of xylem vessels in vascular bundle | 60 | 65 | 68 | 64 | 57 | |
| Diameter of the widest xylem vessel in Vascular bundle | 45.00 | 63.00 | 67.05 | 60.25 | 56.25 | |
| Parenchymatous pith thickness | 498.50 | 861.30 | 869.40 | 738.00 | 678.70 | |
| Hollow pith thickness | 1050.75 | 2070.00 | 2149.20 | 1845.00 | 1415.25 | |

For mesophyll tissue, the thickness of both palisade and spongy tissues were increased with different applied treatments. Here, palisade tissue thickness was 117.90μ in the control but was increased to reach 181.80, 163.35, 148.50 and 137.70 μ with Salicylic acid at 100 ppm, PP333 at 10 ppm, NAA at 200 ppm and Ascorbic acid at 100 ppm, respectively. Also spongy tissues was 193.50 μ in the control but was increased to reach 336.60, 284.18, 273.15 and 221.40 μ with PP333 at 10 ppm, NAA at 200 ppm and Ascorbic acid at 100 ppm, NAA at 200 ppm, Salicylic acid at 100 ppm and Ascorbic acid at 100 ppm, respectively.

Fig. (1): Transverse sections (X = 25) through 4 th internode of the main stem of faba bean plants at 60 days after sowing as affected by different applied treatments.

| Where: | (A): Control(distilled water) | (B): NAA at 200 ppm | (C): PP ₃₃₃ at 10 ppm |
|--------|-------------------------------|--------------------------|----------------------------------|
| | (D): Salicylic acid at 100 pp | m (E): Ascorbic acid | at 100 ppm |
| | ep= Epidermis | co= Cortex | fi=fiber tissue |
| | ph= phloem tissue | cam=cambium region | xy= Xylem tissue |
| | cvb=cortical | vascular bundle pi= pith | hpi=hollow pith |

Fig. (2): Transverse sections (X = 25) through 4 th leaflet of faba bean plants at 60 days after sowing as affected by different applied treatments.

| Where: | (A): Control(distilled water) | (B): NAA at 200 ppm | (C): PP ₃₃₃ at 10 ppm |
|--------|---|--|----------------------------------|
| | (D): Salicylic acid at 100 p | pm (E): Ascorbic acid at 1 | 00 ppm |
| | uep= upper epidermis spo=spongy tissue | lep= lower epidermis co= collenchyma tissue | pl=palisade tissue |
| | ph= phloem tissue | oft=outer fiber tissue | ift=inner fiber tissue |

The applied treatments increased in most of the midrib anatomical features such as thickness of both uppermost and lowermost collenchyma tissue and dimensions of main vascular bundle as well as thickness of phloem tissue, xylem tissue, number and diameter of xylem vessels in main vascular bundle. These increases were more obvious with the pp₃₃₃ and NAA treatment. The results specially increment of the conductive tissues (xylem and phloem) are great importance because they could be involved in the interpretation about why vigorous growth and high yielded pods were existed with different applied treatments specially with pp₃₃₃ at 10 ppm.

flowering characters:-

Data in Table(5) clearly demonstrate that all applied treatments significantly enhanced total number of flowers/ plant, Pollen grains fertility and Pods setting percentage as compared with the untreated plants. On the other hand, the all applied treatments were decreased abscission percentage

Data in Table (5) show fertility and sterility of pollen grains. The treatments especially NAA at 200 or 100 ppm and salicylic acid at 100 or 50ppm increased pollen fertility over the control plant and other treatments. These results could illustrate the increase or decrease of yield as the fertility of pollen grains could be an indication for egg fertility. Meanwhile, the antioxidant treatments might decreased flower abscission percentage by the protection of flower to abscise through increase those substances responsible for scavngening of free radicals and exchange

these radicals to beneficial substances to cause protection of membranes and all cell organelles in plant cell. The above mentioned increase flower number and flower setting as well Eaid (2010) and Zewail (2011)

Maximum values of growth parameter, Total number of flowers, Pollen grains fertility and Pods setting percentage were recorded by NAA and salicylic acid. So, these increments led to increase yield and yield components. These results were in agreement with those of Ismaeil (1995), Fletcher *et al.*(2000), Ishikawa *et al.*(2006), Riad *et al* (2008) and Zewail (2011).

yield and yield components:-

Higher yield and yield components (No. of pods, weight of pod(g), Pods yield(g)\ plant, Weight of seeds\ pod, Seed yield (g)\ plant and Seed index(g))of faba bean as affected by NAA and salicylic acid followed by pp₃₃₃ and ascorbic acid treatments in Table(5) may be attributed to the indirect effect of most materials in many biochemical processes SA applied on basil stimulated the growth and yield by enhancing photosynthesis and nutrient uptake (Gharib, 2006).

The obtained increase of the final seed yield could be attributed to that increase firstly in growth characteristics such as branches number, bacterial nodules number, total leaf area and dry weight (Table, 1) and secondly may be due to that increase in photosynthetic pigments content (Table, 2).

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Thereby, increase of all substances and bioconstituents synthesis and their translocation from leaf and different plant organs to seeds Zewail (2011). NAA increased flower number and flower setting (Table, 5) and increased of all physiological substances to bring plant growth and development and highest production in from of seeds and all different part of plant production of this plant during 2009 and 2010 seasons Qbal *et al.*,(2009)

Chemical analysis:

Data presented in Table (6) indicate that the NAA at 200 or 100 ppm and salicylic acid at 100 or 50ppm followed by $pp_{333}at$ 10 or 5ppm and ascorbic acid at 100 or 50 ppm treatments increased nitrogen, phosphorus, potassium and crude protein contents in seeds. On the other hand, High increase in total carbohydrates was obtained from NAA at 200 or 100 ppm and pp₃₃₃at 10 or 5ppm followed by salicylic acid at 100 or 50ppm.

Table (6): Effect of some growth regulators and antioxidants on some minerals (mg/g D.W.), total carbohydrates, crude protein of faba bean seeds during 2009 and 2010 seasons.

| protein of raba bean seeds during 2005 and 2010 seasons. | | | | | | | | | | | |
|--|-------|---------------------|------|----------------------------|------------------------|--|--|--|--|--|--|
| Characters | | inerals dry weig | ht) | Crude protein (mg/g dry | Total carbohydrates | | | | | | |
| Treatments | N P K | | K | weight) | % | | | | | | |
| Control(distilled water) | 3.85 | 0.38 | 0.18 | 24.06 | 50.82 | | | | | | |
| NAA at 100 ppm | 5.25 | 0.45 | 0.42 | 32.81 | 60.58 | | | | | | |
| 200 ppm | 5.60 | 0.52 | 0.30 | 35.00 | 60.86 | | | | | | |
| PP ₃₃₃ at 5 ppm | 4.06 | 0.53 | 0.22 | 25.38 | 61.47 | | | | | | |
| 10 ppm | 4.20 | 0.43 | 0.25 | 26.25 | 64.93 | | | | | | |
| Salicylic acid at 50 ppm | 4.90 | 0.45 | 0.25 | 30.63 | 59.29 | | | | | | |
| 100 ppm | 4.90 | 0.58 | 0.28 | 30.63 | 60.22 | | | | | | |
| Ascorbic acid at 50 ppm | 4.48 | 0.44 | 0.29 | 28.00 | 52.93 | | | | | | |
| 100 ppm | 4.55 | 0.57 | 0.34 | 28.44 | 55.58 | | | | | | |

In this respect, the increase in nitrogen and phosphorus due to applying the growth substances(NAA) may be the result of its role on regulating ions and may modify the uptake movement and metabolism of nutrients with in the plant tissues. The increase in total carbohydrate in response to treatment applications is supported by stimulation in photosynthetic pigments and the accumulation of the dry matter in the shoots of BA treated plants (Zewail, 2011).

The positive effect existed with pp₃₃₃ on total carbohydrates could be attributed to that increase of chlorophylls content and its reversion on improvement or enhancement of photosynthetic processes (Ismaeil,1995).

These results are in agreement with those of EI – Desouky *et al.*, (2001), EI – Shraiy and Hegazi (2009) Sharaf EI-Deen and Manaf (2009), Eaid (2010) and Zewail (2011)

REFERENCES

- Aloni B.; Karni L.; Zaidman Z. and A. A. Schaffer (1996): Changes of carbohydrates in pepper (*Capsicum annuum* L.) flowers in relation to their abscission under different shading regimes. Ann. Of Bot. 78: 163–168.
- A.O. A. C. (1990): Official Method of analysis, 10th Ed., Association of Official Analysis Chemists, Inc. USA.
- Deriaux, M.; Kerrest, R. and Y. Montalon (1973): Etude de la sulface foliare et de 1, activite photosynthetique chez quiques hybrids de mais. Ann. Amelior plantes, 23: 95-107.
- Downie A. and N. Brewin(2007): The *Rhizobium* legume symbiosis. Molecular Microbiology Department John Innes Centre, Norwich Research Park, Colney, Norwich, NR4 7UH UK
- Dubois, M.; Gilles, K. A.; Hamilton, J. K.; Rebens, P. A. and F. Smith (1956): Colorimetric method for determination sugars and related substances. Anal. Chem. Soc., 46: 1662-1669.
- Duncan, B. D. (1955). Multiple test range and multiple F tests. Biometries. 11-142.
- Eaid, R, S. M. (2010): Physiological studies on the effects of some bioregulators on sesame plant. Theses ,M.Sc., Bot. Dept., Fac. Of Agric., Benha Univ., Egypt.
- El-Desouky, S.A.; Khedr,Z.M.; Wanas,A.L. and H.S. Ahmed, (2001): response of the Egyptian cotton plant to foliar spray with some macro nutrients (NPK) and the growth regulator paclobutrazol pp₃₃₃ I- effects on vegetative growth, leaf anatomy and chemical components., Annals. Agric. Sci. Moshtohor, 39(4): 2087-2107.
- El-Shraiy, A.M.and M.A Hegazi, (2009): Effect of acetylsalicylic acid, indole-3- bytric acid and gibberellic acid on plantgrowth and yield of pea (*pisum sativum l.*), Australian Journal of Basic and Applied Sciences, 3(4): 3514-3523.
- Fletcher, R.A.; Gilley, A.; Davis, T.D. and N. Sankhla (2000):Triazoles as plant growth regulators and stress protectants. Hort. Rev., 24: 55-138.
- Gharib, F. A. E.L. (2006): Effect of Salicylic Acid on the Growth, Metabolic Activities and Oil Content of Basil and Marjoram. Int. J. Agri. Biol., 8(4): 485-492.
- Grossmann, K. (1992): Plant growth reterents: Their mode of action and benefit for physiologicalresearch, In: C.M. Karassan, L.C Van Loon. And D. Vereugdenhil (ed.). Progress in plant growth regulation. Kluwer Academic Publishers, Netherlands, pp: 788-797.
- Gutam Sridhar, R. V. K.; Chetti, M. B. and S. M. Hiremath (2009): Effect of naphthalene acetic acid and mepchloride on physiological components of in bell pepper(*Capsicum annuum* L.). J. Agric. Res., 47(1): 53-62.
- Gutiérrez-Coronado, M.; Trejo, C.L. and A. Larqué-Saavedra (1998): Effects of salicylic acid on the growth of roots and shoots in soybean, Plant Physiol. Biochem., 36(8): 563-565.

- Hegazi, A. M. and A. M. El-Shraiy (2007): Impact of Salicylic Acid and Paclobutrazol Exogenous Application on the Growth, Yield and Nodule Formation of Common Bean. Aust. J. Basic & Appl. Sci., 1(4): 834-840.
- Horneck, D. A. and D. Hanson (1998): Determination of potassium and sodium by Flam Emission Spectrophotometry. In Handbook of Referance Methods for plant analysis. Kalra, Y. P. (ed.):153-155.
- Horneck, D. A. And R. O. Miller (1998):Determination of total nitrogen in plant tissue. In Handbook of Referance Methods for plant analysis. Kalra, Y. P. (ed.): 75-83.
- Hungria, M. and M.A.T. Vargas (2000): Environmental factors affecting nitrogen fixation in grain legumes in the topics, with an emphasis on Brazil. *Field Crops Res.*, 65: 151–164
- Ishikawa, T.; Dowdle, J. and N. Smirnoff (2006): Progress in manipulating ascorbic acid biosynthesis and accumulation in plants. Physiol. Plant. 126: 343–355.
- Ismaeil, F. H. M.(1995): Effect of gamma radiation and paclobutrazol (P.G.R.) on Vicia faba from the botanical point of view. Thesis, M.Sc, Fac. Of Agric.,Zagazig Univ., Benha Branch, Egypt.
- Johanson, D.V. (1940): Plant microtechnique. New York and London McGrow- hill Book Co. Inc. pp.27 154.
- Khafaga, H. S.; Raeefa, A. H.; Hala M. M. and S. A. Alaa(2009): Response of two faba bean cultivars to application of certain growth regulators under salinity stress condition at Siwa oasis.1- Growth traits, yield and yield components. 4th Conference on Recent Technologies in Agriculture, 2009:236-
- Klessig, D.F. and J. Malamy(1994): The salicylic acid signal in plants. Plant Mol. Biol., 26: 1439-1458.
- Nornal, R. (1982): Formulae for determination of chlorophyllous pigments extracted with N, N- Dimethylformamide. Plant Physiology, 69:1371-1381.
- Piper C.S. (1950). Soil and Plant Analysis. Inter. Sci. pub., New York, 213-217.
- Qbal, M.; Qasim Khan, M.; Khalid, R. J. D. and M. Munir(2009): Effect of foliar application of NAA on fruit drop, yield and physio-chemical characteristics of guava(*Psidium guajava* L.) red flesh cultivar. J. Agric. Res., 47(3): 259-269.
- Rasmussen, J.B.; Hammerschmidt, R. and M.N. Zook (1991): Systemic induction of salicylic acid accumulation in cucumber after inoculation with Pseudomonas syringae pv. Syringae. Plant Physiol., 97: 1342-1347.
- Riad, G. S.; Ghoname, A.A.; Sadak, M. S. and A.M. Hegazi(2008): Alleviation of poor growth eggplant under newly reclaimed land by foliar application of biostiimulators and micronutrients. Res. J. of Agric. And Biol. Sci., 4(6): 964-972.
- Sakhabutdinova, A.; Fatkhutdinova and F. RandShakirova (2003): Salicylic acid prevents the damaging action of stress factors on wheat plants. Bulg. J. Plant physiol., special issue 2003, 314–319

Sandell, R. (1950): Colorimetric determination of traces of metal 2nd Ed. Interscience pub., Inc. New York.

- Sass, J. E. (1951): Botanical micro technique Iowa state college press, Ames, Iowa, pp.228.
- Shahine, A. H. (1961): Effect of gamma radiation of seeds on germination, growth and some economical characters of barley. Ph. D. thesis, Zagreb Univ. Yugoslavia.

Sharaf El-Deen, H. A. M. and H. H. Manaf (2009). Effect of some plant growth regulators on shedding physiology, antioxidant enzymes, growth and yield of faba bean. J. Biol.Chem.Environ.Sci.,4(3):21-46.

Snedecor, G.W. and W.G. Cochran (1972): Statistical Methods. 6th ed. The Iowa state. Univ. Press, Amer, Iowa, U.S.A. PP. 593.

Zewail, R. M. Y. (2011): Inducing cold tolerability in faba bean plants for maximizing its productivity. Thesis, Ph. D., Bot. Dept., Fac. Of Agric., Benha Univ., Egypt.

تأثير بعض منظمات النمو و مضادات الأكسدة على النمو، المحصول و التركيب الكيماوي للبذور في نباتات الفول فاتن حسن محمود إسماعيل و محمد محمد محمود عبد العال قسم النبات-كلية الزراعة- جامعة بنها

تمت هذه الدراسة فى مزرعة كلية الزراعة بمشتهر – جامعة بنها فى موسمى النمو ٢٠٠٩ م. تهدف هذه الدراسة الى دراسة تأثير بعض منظمات النمو و مضادات الأكسدة على الصفات المورفولوجية، التشريحية، عدد العقد البكتيرية، الإز هار، تساقط الأز هار،خصوبة حبوب اللقاح، المحصول و التركيب الكيماوي للبذور في نباتات الفول صنف جريدلى. حيث نقعت البذور فى التركيزات الآتية:

١- الكنترول، ٢- نفتالين حمض الخليك بتركيز ١٠٠ و ٢٠٠ جزء في المليون، ٣- الباكلوبيوتر ازول بتركيز ٥ و
 ١٠ جزء في المليون، ٤- حمض الساليسيلك بتركيز ٥٠ و ١٠٠ جزء في المليون و ٥- حمض الإسكوربيك بتركيز ٥٠ و ١٠٠ جزء في المليون و ٩٠٠ يوم من النباتات بنفس المعاملات عند ٤٠ و ٢٠ يوم من الزراعة.

أعطت كل المعاملات زيادة معنوية للصفات المورفولوجية، عدد العقد البكتيرية، خصوبة حبوب اللقاح، المحصول ، محتوى البذرة من النيتروجين، القوسفور، البوتاسيوم، البروتين و الكربوهيدرات.

أعطت المعاملات نفتالين حمض الخليك بتركيز ١٠٠ و ٢٠٠ جزء في المليون و حمض الساليسيلك بتركيز ٥٠ و ١٠٠ جزء في المليون يتبعها الباكلوبيوتر ازول بتركيز ٥ و ١٠ جزء في المليون و حمض الإسكوربيك بتركيز ٥٠ و ١٠٠ جزء في المليون أعلى تأثير مقارنةً بالكنترول. و أيضاً نفس المعاملات أدت الى تقليل النسبة المئوية للتساقط.

بالإضافة الى أن نفثالين حمض الخليك بتركيز ٢٠٠ جزء فى المليون و حمض الساليسيلك بتركيز ١٠٠ جزء فى المليون أعطيا أقل نسبة مئوية للتساقط و عقم حبوب اللقاح و أعلى محصول من البذور و أدت الى اختلافات معنوية فى الصفات التشريحية.

قام بتحكيم البحث

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 كلية الزراعة – جامعة بنها

| Characters | Root length (cm) | Nodules number/ | Plant height | No. of Branches | No. of leaves | Total leaf area | Root dry weight | Stem dry weight | Leaf dry weight | | | |
|----------------------------|---------------------|--------------------|-----------------|--------------------|------------------|---------------------------|--------------------|--------------------|--------------------|--|--|--|
| Treatments | (0) | plant | (cm) | /plant | /plant | (cm ²)/ plant | (g)/plant | (g)/plant | (g)/plant | | | |
| | | 2009 season | | | | | | | | | | |
| Control(distilled water) | 13.17C | 20.00D | 38.33E | 3.33C | 46.00C | 3657.00D | 4.15B | 8.47E | 8.53D | | | |
| NAA at 100 ppm | 18.67AB | 33.33AB | 48.00B | 5.33AB | 48.33C | 6699.00A | 6.77A | 11.49AB | 11.16BC | | | |
| 200 ppm | 20.00A | 35.00A | 51.17A | 5.33AB | 54.33B | 6888.00A | 7.04A | 12.42A | 1181AB | | | |
| PP ₃₃₃ at 5 ppm | 15.67BC | 26.67ABCD | 43.67CD | 6.00AB | 59.67A | 5628.00ABC | 5.21AB | 9.54CDE | 12.56AB | | | |
| 10 ppm | 17.00AB | 26.67ABCD | 41.00DE | 6.67A | 61.67A | 5974.00ABC | 5.61AB | 9.79BCDE | 13.40A | | | |
| Salicylic acid at 50 ppm | 17.33AB | 28.33ABCD | 46.50BC | 5.67AB | 55.67B | 6407.00AB | 5.84AB | 11.11ABCD | 12.13AB | | | |
| 100 ppm | 18.00AB | 30.00ABC | 46.67B | 5.67AB | 59.33A | 6680.00A | 6.65A | 11.29ABC | 12.53AB | | | |
| Ascorbic acid at 50 ppm | 13.33C | 23.33CD | 44.00CD | 4.33BC | 47.00C | 5110.00C | 4.15B | 8.90E | 9.18D | | | |
| 100ppm | 13.33C | 25.00BCD | 46.50BC | 5.00ABC | 48.33C | 5288.00BC | 5.14AB | 9.39DE | 9.84CD | | | |
| | | | | 2 | 010 seaso | n | | | | | | |
| Control(distilled water) | 13.67E | 23.33D | 39.67F | 3.67C | 48.33C | 5563.00D | 5.59C | 8.75E | 8.79E | | | |
| NAA at 100 ppm | 20.67AB | 40.67AB | 52.50B | 6.33AB | 55.67B | 8427.00A | 7.59A | 13.08B | 12.24BC | | | |
| 200 ppm | 21.67A | 45.00A | 57.67A | 6.33AB | 56.00B | 8462.00A | 8.47A | 15.55A | 12.29BC | | | |
| PP ₃₃₃ at 5 ppm | 16.33CDE | 30.00CD | 45.00DE | 7.67AB | 62.67A | 7396.00ABC | 5.86BC | 11.03CD | 15.85A | | | |
| 10 ppm | 15.67DE | 33.33BC | 42.33EF | 8.33A | 63.33A | 7637.00AB | 6.01BC | 11.25BCD | 15.96A | | | |
| Salicylic acid at 50 ppm | 18.67BC | 35.00BC | 48.50CD | 6.67AB | 62.00B | 8137.00A | 6.03BC | 12.06BCD | 13.81B | | | |
| 100 ppm | 20.00AB | 36.67BC | 49.83BC | 7.00AB | 62.33A | 8237.00A | 7.27AB | 12.63BC | 15.69A | | | |
| Ascorbic acid at 50 ppm | 14.33E | 28.33CD | 45.50DE | 5.67BC | 50.33C | 6499.00CD | 5.75BC | 10.22DE | 10.19DE | | | |
| 100 ppm | 15.67DE | 30.00CD | 48.00CD | 6.33AB | 51.33C | 6661.00BC | 5.76BC | 10.88CD | 10.67CD | | | |

 Table (1): Effect of some growth regulators and antioxidants on morphological characters, bacterial nodules number and dry weight of faba bean plants during 2009 and 2010 seasons.

Table (4): Effect of some growth regulators and antioxidants on the mean counts and measurements of certain anatomical features of faba bean leaf at 85 days after sowing during 2010 seasons.

| anatonnical reactives of taba bean real at 05 days | | | | | |
|---|-------------------|---------|-------------------------|------------|---------------|
| Treatments | Control | NAA at | PP ₃₃₃ at 10 | | Ascorbic acid |
| Histological Characteristics (µ) | (distilled water) | 200 ppm | ppm | at 100 ppm | at 100 ppm |
| Thickness of upper epidermis cuticle layer | 16.50 | 14.40 | 18.00 | 15.30 | 10.80 |
| Thickness of Lower epidermis cuticle layer | 12.60 | 10.80 | 15.30 | 12.90 | 9.00 |
| Upper epidermis thickness | 36.90 | 49.50 | 55.35 | 43.20 | 41.40 |
| Lower epidermis thickness | 26.10 | 33.75 | 34.20 | 32.40 | 33.30 |
| Palisade tissue thickness | 117.90 | 148.50 | 163.35 | 181.80 | 137.70 |
| Spongy tissue thickness | 193.50 | 284.18 | 336.60 | 273.15 | 221.40 |
| Thickness of blade | 403.50 | 541.13 | 622.80 | 558.75 | 453.60 |
| Thickness of collenchyma layers below the upper epidermis at midrib | 45.90 | 105.30 | 98.10 | 77.40 | 63.90 |
| Thickness of collenchyma layers above the lower epidermis at midrib | 36.90 | 51.30 | 64.80 | 64.35 | 46.80 |
| Thickness of upper fibers layers in the vascular bundle | 112.50 | 153.90 | 150.30 | 156.60 | 231.30 |
| Thickness of xylem tissue | 201.60 | 244.80 | 225.10 | 211.80 | 215.20 |
| Number of xylem vessels in the vascular bundle | 36 | 52 | 59 | 47 | 37 |
| Thickness of widest xylem vessel in the vascular bundle | 31.50 | 47.70 | 39.15 | 37.80 | 41.85 |
| Thickness of phloem in the vascular bundle | 61.20 | 86.40 | 95.40 | 94.50 | 85.95 |
| Thickness of lower fibers layers in the vascular bundle | 100.35 | 109.80 | 120.60 | 108.00 | 117.10 |
| Length of midrib vascular bundle | 556.20 | 649.80 | 621.90 | 563.40 | 558.00 |
| No. of vascular bundle in midrib | 1 | 1 | 1 | 1 | 2 |
| Thickness of leaf midrib | 1058.40 | 1275.30 | 1374.60 | 1291.70 | 1251.10 |
| | | | | | |

Table (5): Effect of some growth regulators and antioxidants on flowering, fruit setting, flower abscission, pollen

| grains fertility. | , yield and y | vield com | ponents of | faba bean | plants during | g 2009 and 2010 seasons. |
|-------------------|---------------|-----------|------------|-----------|---------------|--------------------------|
| | | | | | | |

| Characters | No. of flowers / | % of Abscissi | pollen fertili | | No. of setted | % of setted | Weight of | Pods yield(g)/ | Weight of seeds/ | Seed yield (g)\ | Seed |
|----------------------------|---------------------|------------------|-------------------|-----------|----------------|-------------|--------------|-------------------|---------------------|--------------------|----------|
| Treatments | plant | on | Fertility | Sterility | pods/ plant | pods | pod(g) | plant | pod | plant | index(g) |
| | | | | | | 2009 season | | | | | |
| Control(distilled water) | 58.00C | 58.59A | 49.46D | 50.54A | 22.74B | 39.38B | 2.13C | 56.80F | 1.647D | 54.61D | 82.33D |
| NAA at 100 ppm | 70.00AB | 43.58B | 94.12A | 5.88FG | 32.88AB | 53.56A | 3.75A | 139.20A | 3.253AB | 131.10A | 162.7AB |
| 200 ppm | 72.23A | 46.44B | 94.97A | 5.03FGH | 40.67A | 56.42A | 3.76A | 144.00A | 3.303A | 142.20A | 165.2A |
| PP ₃₃₃ at 5 ppm | 60.33C | 49.96B | 77.26C | 22.74C | 26.65B | 50.04A | 3.56A | 90.38DE | 2.803BC | 86.77C | 140.2BC |
| 10 ppm | 61.67C | 47.59B | 81.82AB | 18.18D | 31.23AB | 52.41A | 3.62A | 101.60CD | 3.003AB | 90.56C | 150.2AB |
| Salicylic acid at 50 ppm | 62.33C | 47.09B | 87.79AB | 12.21E | 31.24AB | 52.91A | 3.56A | 111.50BC | 3.130AB | 98.15BC | 162.5AB |
| 100 ppm | 63.67BC | 46.50B | 92.74A | 7.26F | 31.61AB | 53.50A | 3.62A | 118.90B | 3.259AB | 102.90B | 156.5AB |
| Ascorbic acid at 50ppm | 58.33C | 58.57A | 72.89C | 27.11B | 24.98B | 41.43B | 2.28C | 66.35F | 1.797D | 61.84C | 89.33D |
| 100 ppm | 58.33C | 58.50A | 74.70C | 25.30BC | 25.56B | 41.50B | 3.08B | 82.90E | 2.450C | 81.84C | 122.5C |
| | | | | | 2 | 2010 season | | | | | |
| Control(distilled water) | 60.33C | 56.63A | 56.90D | 49.10A | 26.15D | 43.37C | 2.26C | 81.59D | 1.767C | 64.84D | 88.33C |
| NAA at 100 ppm | 80.00A | 42.43BC | 94.38A | 5.62 EF | 44.08A | 57.57AB | 3.85A | 169.40A | 3.343A | 147.30A | 167.2A |
| 200 ppm | 81.33A | 40.63C | 94.06A | 5.94EF | 46.72A | 59.37A | 3.91A | 175.30A | 3.410A | 156.40A | 170.5A |
| PP ₃₃₃ at 5 ppm | 67.67B | 49.15ABC | 77.12C | 22.88BC | 35.67BC | 50.85ABC | 3.74A | 112.30BC | 3.237A | 94.80BC | 161.8A |
| 10 ppm | 69.67B | 46.82ABC | 84.21AB | 15.79CD | 35.75BC | 53.18ABC | 3.75A | 112.90BC | 3.250A | 105.70B | 162.5A |
| Salicylic acid at 50 ppm | 70.33B | 46.68ABC | 85.37AB | 14.63CD | 36.65BC | 53.32ABC | 3.77A | 134,10B | 3.270A | 139.40A | 163.5A |
| 100 ppm | 78.67A | 45.78ABC | 90.91A | 9.09E | 42.55AB | 54.22ABC | 3.84A | 166.30A | 3.343A | 145.00A | 167.2A |
| Ascorbic acid at 50ppm | 63.67BC | 52.70AB | 74.54C | 25.46B | 32.56CD | 47.30BC | 2.46C | 82.87D | 1.967C | 65.12D | 98.33C |
| 100 ppm | 65.33BC | 49.47ABC | 81.08AB | 18.92C | 33.07C | 50.53ABC | 3.18B | 97.68CD | 2.673B | 84.46C | 133.7B |

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