MICROBIOLOGICAL STUDIES ON BACTERIAL COMMUNITIES IN SOME LOCAL FERMENTED MILK PRODUCTS (LABAN) IN SAUDI ARABIA AI-Harbiy, Nada; H. Z. Hassan and Kawther Abed Biology Dept., Fac. of Science, Princess Nora Bint Abdul Rahman Univ.

ABSTRACT:

The present investigation was conducted to identify and characterize bacterial communities including starter bacteria as well as pathogenic ones, on some local fermented milk products (Laban) produced in Saudi Arabia in order to ensure human health. The isolated strains were characterized by phenotypic, physiological and biochemical methods, including API 50 CHL kit. The results showed that lactic acid bacteria (LAB) were present in high counts . The dominated species found were Streptococcus salivaius ssp. thermophilic (47.7%), Lactobacillus dellberckii ssp. bulgricus (22.2%), Lactobacillus acidophilus (22.2%), Leuconostoc lactis (5.5%), Lactococcus lactis ssp. lactis (2.2%). The results obtained demonstrated that the pathogenic bacteria in five examined products are Escherickia coli, Enterococcus faecalis, Enterococcus cloacae, Enterococcus durens, Enterococcus casselifavus, Pasudomonus capacia, Streptococcus mitis, Bacillus cereus, and Corynebacterium ssp. .

Keywords: Lactic acid bacteria (LAB), pathogenic bacteria, fermented milk products (Laban), Saudi Arabia.

INTRODUCTION

Laban is a fermented milk product produced in Saudi Arabia and some Arab countries using traditional and industrial manufacturing practices (Chammas et al. 2006). Lactic acid bacteria (LAB) have long been consumed by people in several fermented foods such as dairy products (Khedid et al., 2006) Nowadays, LAB are a focus of intensive international research for their essential role in most fermented food, for their ability to produce various antimicrobial compounds promoting probiotic properties (Temmerman et al., 2003) reduction of serum cholesterol (Desmazeaud, 1996; Jackson et al.,2002) including antitumor activity (De Vuyst and Degeest,1999; Hilde et al., 2003) stimulation of the immune system (Isolauri et al., 2001); stabilization of gut microflora (Gibson et al., 1997). According to Tamime and Robinson (1999) laban is obtained through the lactic fermentation of heattreated milk by using thermophilic starters, composed of Streptococcus thermophilus and Lactobacillus delbrueckii subsp. bulgaricus strains. The thermophilic lactic acid bacteria are best known as starters for fermented milks. Several varieties of fermented milks originate from countries in Asia Minor and the Balkans, like Armenia, Turkey and Bulgaria. These products have emerged from spontaneous acidification of raw milk by indigenous organisms. Although these organisms have by no means been exhaustively characterised, they consist largely of thermophilic lactic acid bacteria, probably due to the relatively high incubation temperature determined by the prevailing climate. The first description of milk fermentations by these

Al-Harbiy, Nada et al.

bacteria can be found in the literature of some hundred years ago (Weigmann, 1905). Several attempts were made at that time to identify the bacteria dominating the flora in yoghurt-like products and they were given the names Bacillus bulgaricus and Diplostreptococcus. These spontaneous fermentations of milk into yoghurt have now been developed into microbiologically well-controlled industrial processes. The two most frequently used starter bacteria are now classified as Lactobacillus delbrueckii subsp. bulgaricus and Streptococcus salivarius subsp. thermophilus, generally shortened as Lb. bulgaricus and S. thermophilus, respectively. Furthermore, many researches have studied the behavior of pathogenic bacteria in fermented milks (Al Shaikli,1980; Salji et al, 1983; Nasser *et al.*, 2004; Ombui and Nduhiu, 2005; Aygun and Pehlivanlar, 2006; Zelalem *et al.*,2007).

The main goals of this study were to isolate and identify LAB and pathogenic bacteria from 60 fermented milk samples belonging to 5 commercial companies were gathered at 5 different sites of Riyadh city in Saudi Arabia.

MATERIALS AND METHODS

Samples collection:

A total of 60 samples of fermented milk products (Laban) belonging to 5 commercial companies were collected from 5 different sites in the north, east, south, west, and central regions of Riyadh city. The samples were obtained from both large trade centers and small markets at each site. All samples have the same production dates. The samples were immediately cooled and transported to the laboratory in icebox 4° C and analyzed for the content of LAB and pathogenic bacteria on the arrival.

Isolation of the strains:

- LAB:

Aliquots of 10 ml of fermented milk sample was homogenized with 90 ml of peptone water to make an initial dilution (10^{-1}) . The suspension was used for making suitable serial dilution up to 10^{-9} by incorporating 1 ml into 9 ml of sterile peptone water in sterile tubes. Enumeration of LAB was determined using various selective media, MRS agar (pH 5.3) (Biokar, France) and M17 agar (pH 6.2) (Biokar, France). After incubation at 30° C (24 - 48 h) colonies were enumerated, recorded as colony forming units (CFU/g cheese). The colonies were randomly picked from plates with 30- 300 colonies, and transferred in 10 ml of appropriate broth. The selected colonies were purified by repeated streaking on the appropriate ager media. LAB strains were kept on media slant at 4°C and streaked every 4 weeks. Prior to use, LAB strains were activated in broth media at 30°C for 24 h, and subcultured in MRS agar at 30°C for 24 h.

- Pathogenic bacteria:

Ten ml of fermented milk sample was homogenized with 90 ml of saline water to make an initial dilution (10-1). The suspension was used for making suitable serial dilution up to 10-8 by incorporating 1 ml into 9 ml of

J. Agric. Chemistry and Biotechnology, Mansoura Univ., Vol. (7), July, 2010

sterile saline water in sterile tubes. Enumeration of pathogenic bacteria was determined using various selective media, Nutrient agar (The total bacterial count), MacConkey agar (Coliform), Bile Aesculin agar (Enterococci), Mannitol salt agar (Staphylococcus), and Bacillus cereus selective agar Base (Bacillus). After incubation at 37°C (24 - 72 h) colonies were enumerated, recorded as colony forming units (CFU /ml) of cheese. The colonies were picked from plates, and transferred in 10 ml of appropriate broth. The selected colonies were purified by repeated streaking on the appropriate ager media. Pathogenic bacteria strains were kept on slants of the suitable media at 4°C.

Identification of the studied bacterial isolates:

LAB strains were identified by morphological, physiological and biochemical techniques according to methods recommended by several authors (Facklam and Collins, 1989; Charteris *et al.*, 2001; and Klein, 2001). All strains were initially subjected to Gram staining, catalase test, growth at 10 - 45 °C in MRS and M17 broth, and gas production form glucose. All strains were tested for their sugar fermentation patterns using API 50 CHL, and duplicated for each isolate. Pathogenic bacteria strains were identified by morphological, physiological and biochemical techniques according to methods recommended by several authors (Van Netten, 1989; Martin *et al.*,1967; and Holbrook and Andersson, 1980). All strains were initially subjected to Gram staining and catalase test. Identification of bacteria was carried out using the methods recommended in Bergey's Manual of Systematic Bacteriology Vol. 1 (Holt, 1984).

RESULTS AND DISCUSSION

LAB counts in fermented milks:

The M17 agar medium was used to estimate the numbers of Streptococcus salivarius ssp. thermophilus while MRS agar medium was used to elucidate the numbers of Lactobacillus. The results obtained show that the estimated starter bacterial numbers on M17 agar media were ranged from 108 to 109 while those estimated on MRS agar media were ranged from 106 to 107 (CFU/ml) (Table 1) . Statistical analysis shows that there were no significantly differences between the means of the starter bacterial numbers on the M17 agar media for the samples in the large centers as compared with those in the small markets the beginning of the production. The results showed that there were decrease in the numbers of the starter bacteria on M17 agar media for the samples stored in the larger centers at the expire date as compared with those at the beginning. Further decrease was noticed in the small markets .

Also, there were no significantly differences between the means of the starter bacterial numbers on the MRS agar media for the samples in the large centers as compared with those in the small markets at the beginning of the production. Also, the recorded data showed that there were decrease in these numbers on the same media for the samples stored in large centers at

Al-Harbiy, Nada et al.

the expired date as compared with those at the production date. Additional decrease was also recorded in the small markets.

gi e tri e i miti agai ana mite agai meala					
Cheese products	M17 agar	MRS agar			
Product I	1.67×109	1.09×107			
Product II	1.64×109	1.6×106			
Product III	2.30×109	2.3×106			
Product IV	9.40×108	9.11×107			
Product V	2.19×108	0			

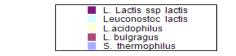
 Table 1: The mean numbers of starter bacteria from fermented milk

 grown on M17 agar and MRS agar media

Identification of isolates:

A total of 250 isolates were isolated from fermented milk samples gathered from 5 trade companies. A preliminary identification for these isolates was carried out on selective media. The identification for all isolates was continued by gram stain and catalase test.

The results of the identification revealed that the isolates grown on M17 agar medium have spherical shape to oval or short rods, usually present in pairs or chains, immotile, catalase negative and gram positive. While results of identification for the isolates grown on MRS agar medium have uniform rod shape. However, some isolates take short rod shape. They were gram positive, catalase negative and obligate anaerobic. A total of 90 isolates from these lactic acid n bacteria representing all the studied samples were randomly selected. Then they were identified by the identification ribbons API 50 CHL. The results recorded show that the lactic acid bacteria present in the samples belonging to the trade company-I were Streptococcus salivarius ssp. thermophilus (present in about 35%), Leuconostic lactis (15%) and Lactobacillus dellberckii ssp. bulgricus (50%) (Fig. 1). Streptococcus salivarius ssp. thermophilus and Lactobacillus acidophilis were recorded in the samples of the trade company-II with ratio of 50% for each type (Fig. 1). Considering bacterial types noticed in the samples of the trade company-III were Streptococcus salivarius ssp. thermophilus, Lactobacillus acidophilus and Leuconostoc lactis with ratios of 40, 50 and 10%, respectively (Fig.1). Also, Streptococcus salivarius ssp. thermophilus, Lactobacillus dellberckii ssp. bulgricus and Lactococcus lactis ssp. lactis were detected with ratios 45, 50 and 5%, respectively in the samples of the trade company IV (Fig. 1). The gained results revealed that Streptococcus salivarius ssp. thermophilus represents the most pronounced bacterial type followed by Lactococcus lactis ssp. lactis recognized in the samples of the trade company V. They were recorded with ratios 90 and 10%, respectively (Fig. 1).



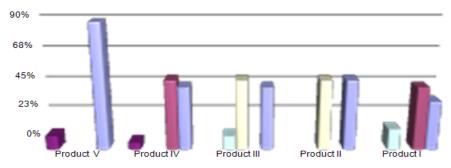


Fig. 1 Lactic acid bacteria species isolated from fermented milk products.

In general, mean ratios of *Streptococcus salivarius* ssp. thermophilus, Lactobacillus dellberckii ssp. bulgricus, Lactobacillus acidophilis, Leuconostoc lactis and Lactococcus lactis ssp. lactis in all samples gathered from the five trade companies were 47.77%, 22.22%, 22.22%, 5.5% and 2.2%, respectively (Table 2) (Fig. 2).

Table 2: Identification of the Lactic acid bacteria species isolated from fermented milk products

Isolates	Total of isolates	Present
Streptococcus salivarius ssp. thermophilus	43	47.77%
Lactobacillus dellberckii bulgricus	20	22.2%
Lactobacillus acidophilis	20	22.2%
Leuconostoc lactis	5	5.5%
Lactococcus Lactic ssp. lactis	2	2.2%

By considering their phenotypical characteristics, all strains isolated from fermented milk samples belonged to S. thermophilus and L. bulgaricus. This confirmed that laban is fermented milk product similar to yogurt, as already reported by Tamime and Robinson, 1999.

Counting and identification of pathogenic bacteria in the tested fermented milks samples:

In this study contamination of the fermented milk samples collected from the five trade companies were determined. The numbers of pollutants were counted using selective media at the beginning of production date as well as at the expired date. The results obtained revealed that only two out of the five sample collections have high contamination growth while the other

Al-Harbiy, Nada et al.

three collections did not show any contamination growth (Table 3). The results also show that contaminant types were Escherichia coli, Enterococcus faecalis, Enterobacter cloacae Enterococcus durens, Enterococcus casseliflavus, Pasudomonus capacia, Corynebacterium ssp., Streptococcus mitis, and Bacillus ceres.

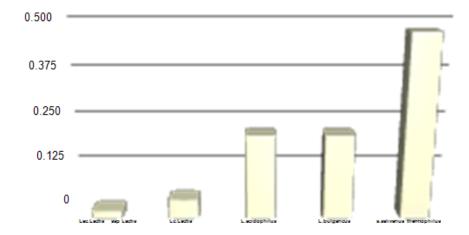


Fig. 2 Identification of the Lactic acid bacteria species isolated from fermented milk products

Cheese products	Nutrient Agar	Nutrient Agar 10 °C	MacConakey agar	Mannitol Salt agar	Bacillus Cereus Agar	Bile Aesculin Agar	Ss agar
I	1.3×108	0	3.28×104	6.9×102	0	4.24 ×107	0
II	1.8×107	0	1.27×105	0	1.9×104	1.8×102	0
III	3.3×105	0	0	0	0	0	0
IV	1.9×105	0	0	0	0	0	0
V	9.3×104	0	0	0	0	0	0

Table 3: Pathogenic bacteria in the tested fermented milk samples

In fact fermented food products are usually considered safe because of the low pH and production of antimicrobial substances by fermenting organisms.

REFERENCES

Al-Shaikhli, J.S. (1980). A Study on the market milk in the Riyadh area. Inciden of coliform contamination. Proceedings of the 4th Symposium on the Biological Aspects of Saudi Arabia. 69.

- Aygun, O. and Pehlivanlar, S. (2006). Listeria spp. in the raw milk and dairy products in Antakya, Turkey. Food Control, 17: 676- 679.
- Chammas, G. I.; Saliba, R.; Corrieu, G.; and Beal, C. (2006). Characterisation of lactic acid bacteria isolated from fermented milk Laban. Int. J. Food Microbiol., 110(1): 52- 61.
- Charteris, W.P.; Kelly, P.M.; Morelli, L.; and Collins, J.K. (2001). Quality control *Lactobacillus* strains for use with the API50 CH and API ZYM systems at 37 °C. J. Basic Microbiol., 41(5): 241-251.
- De Vuyst, L., and Degeest, B., (1999). Heteropolysaccharides from lactic acid bacteria. FEMS Microbiol. Rev., 23(2): 130- 135.
- Desmazeaud, M., (1996). Les bacteries lactiques dans l'alimentation humaine: utilisation et innocuite Cahiers Agric. 5: 331- 343.
- Facklam, R.R., and Collins, M.D. (1989). Identification of Enterococcus species isolated from human infections by a conventional test scheme. J. Clin. Microbiol., 27(4): 731-734.
- Gibson, G. R., Saavedra, J. M., Mac Farlane, S., and Mac Farlane, G. T., (1997). Probiotics and intestinal infections. In: Fuller, R. (Ed.), Probiotics. 2: Application and Practical Aspects. Chapman and Hall, New York, pp. 10 - 39.
- Ostille, H. M., Helland, M. H., and Narvhus, J. A., (2003). Growth and metabolism of selected strains of probiotic bacteria in milk. Int. J. Food Microbiol., 87(1-2): 17-27.
- Holbrook, R., and Andersson, J. M. (1980). An improved selective and diagnostic medium for the isolation and enumeration of Bacillus cereus in foods. Can. J. Microbiol., 26:753-759.
- Holt, J.G (1984). Bergey's Manual of Systematic Bacteriology. Gram-negative bacteria of general, medical or industrial importance Williams, Wilkins, Baltimore.
- Jackson, M. S., Bird, A. R., Mc Orist, A.L., (2002). Comparison of two selective media for the detection and enumeration of lactobacilli in human faeces. J. Microbiol. Methods, 51(3): 313-321.
- Isolauri, E., Sutas, Y., Kankaapaa, P., Arvilommi, H., and Salminen, S., (2001). Probiotics : effects on immunity. Am. J. Clin. Nutr., 73(2): 444-450.
- Khedid, K., Faid, M., Mokhtari, A., Soulaymani, A., and Zinedine, A., (2009). Characterization of lactic acid bacteria isolated from the one humped camel produced in Morocco. Microbiol Res,164(1) : 81-91.
- Klein, G., (2001). International committee of systematic bacteriology, subcommittee on the taxonomy of Bifidobacterium, Lactobacillus and related organisms minutes of the meeting. Int. J. Syst. Evol. Microbiol., 51: 259-261.
- Martin, J. E., Billings, T. E.; Hackney, J. F. and Thayer J. D. (1967). Primary isolation of N. gonorrhoeae with a new commercial medium. Public Health Rep., 82(4): 361-363.
- Nasser, L.A.; El-kersh, T.A. and Mejaly S.H. (2004). Enterococcal isolates from raw milk and dairy products in Riyadh region and their susceptibility to common antibiotics, Bull. Pharm. Sci. Assiut University, 27: 133-144.

- Ombui, J.N. and Nduhiu, J.G. (2005). Prevalence of enterotoxigenic Bacillus cereus and its enterotoxins in milk and milk products in and around Nairobi. East Afr. Med J., 82 (6): 280-284.
- Salji, J.P.; Sawaya; W.N. and Ayaz, M. (1983). The yoghurt industry in the central province of Saudi Arabia. J. Cult. Dairy Prod., 18(4) : 14-19.
- Tamime, A. Y. and Robinson, R.K. (1999). Yoghurt Science and Technology(2nd Edition), pp. 11-16. woodhead Publishing, Cambridge.
- Temmerman, R., Pot, B., Huys, G., and Swings, J., (2003). Identification and antibiotic susceptibility of bacterial isolates from probiotic products, Int. J. Food Microbiol., 81(1): 1-10.
- Van Netten, P.; Perales, I.; van de Moosdijk, A.; Curtis, G.D.W.; Mossel D. A. A.(1989). Liquid and solid selective differential media for the detection and enumeration of L. monocytogenes and other Listeria spp. Int. J. Food Microbiol., 8 (4): 299-316.
- Weigmann, H. (1905). Die Garungen der Milch und der Abbau Ihrer Bestandteile. In F. Lafar (Ed.), Handbuch der Technischen Mykologie.
 2. Mykologie der Nahrungsmittelgewerbe pp. 48-104. Jena: Verlag von Gustav Fischer.
- Zelalem,Y.; Faye, B. and Loiseau, G. (2007). Occurrence and distribution of species of Enterobacteriaceae in selected Ethiopian traditional dairy products: A contribution to epidemiology. Food Control, 18 (11): 1397-1404.

دراسات ميكروبية للمجتمعات البكتيرية في بعض منتجات الالبان المتخمرة المحلية (اللبن) في المملكة العربية السعودية ندى الحربي ، حسام الدين زكي حسن و كوثر عابد قسم الأحياء - كلية العلوم - جامعة الأميرة نورة بنت عبدالرحمن

أجريت هذه الدراسة لتحديد وتوصيف مجتمعات البكتيريا بما في ذلك بكتيريا البادئ فضلا عن الأنواع الممرضة منها في بعض منتجات الألبان المتخمرة المحلية (اللبن)التي تنتج في المملكة العربية السعودية من أجل ضمان صحة الإنسان. تم تعريف السلالات المعزولة باستخدام الشكل الظاهري و الاختبارات الفسيولوجية والبيوكيميائية بالإضافة في (LAB) وأظهرت النتائج ارتفاع أعداد بكتيريا حمض اللاكتيك API 50 CHL. إلى استخدام نظام Streptococcus salivaius ssp. thermophilic (47.7%) العينات المختبره . و قد هيمنت الأنواع

Lactobacillus dellberckii ssp. bulgricus (22.2%), Lactobacillus acidophilus (22.2%), Leuconostoc lactis (5.5%), Lactococcus lactis ssp. lactis (2.2%)

كما أظهرت النتائج المتحصل عليها ايضا أن البكتيريا الممرضة في الخمسة المنتجات محل الدراسة هي Escherickia coli, Enterococcus faecalis, Enterococcus cloacae, Enterococcus durens, Enterococcus casselifavus, Pasudomonus capacia, Streptococcus mitis, Bacillus ceres, and Corynebacterium ssp

قام بتحكيم البحث

كلية الزراعة – جامعة المنصورة	أ.د / محمود محمد عوض الله السواح
كلية العلوم ـ جامعة المنصورة	اد / فتحي عواد منصور